

Genomic Designing of Climate-Smart Oilseed Crops

Chittaranjan Kole
Editor

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 Springer

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ISBN 978-3-319-93535-5 ISBN 978-3-319-93536-2 (eBook)
<https://doi.org/10.1007/978-3-319-93536-2>

Library of Congress Control Number: 2018965428

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The registered company address is: Gewerbestrasse 11, 6330 Cham, Switzerland



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Previously Professor and Bascom Chair in
Agronomy in the University of Wisconsin at
Madison,*

*Where I worked with him on the Brassicaceae
system and learnt how to develop novel
concepts of plant molecular genetics and
side-by-side generate genetic resources for
crop improvement.*

Preface

The last 120 years have witnessed a remarkable evolution in the science and art of plant breeding culminating in quite a revolution in the second decade of the twenty-first century! A number of novel concepts, strategies, techniques, and tools have emerged from time to time over this period, and some of them deserve to be termed as milestones. Traditional plant breeding, immediately following the rediscovery of the laws of inheritance, has been playing a spectacular role in the development of innumerable varieties in almost all crops during this entire period. Mention must be made on the corn hybrids, rust-resistant wheat, and obviously the high-yielding varieties in wheat and rice that ushered the so-called green revolution. However, the methods of selection, hybridization, mutation, and polyploidy employed in traditional breeding during this period relied solely on the perceivable phenotypic characters. But most, if not all, of the economic characters in crops are governed by polygenes which are highly influenced by environment fluctuations, and hence, phenotype-based breeding for these traits has hardly been effective.

Historical discovery of DNA structure and replication in 1953 was followed by a series of discoveries in the 1960s and 1970s that paved the way for recombinant DNA technology in 1973 facilitating the detection of a number of DNA markers in 1980 onwards and their utilization in construction of genetic linkage maps and mapping of genes governing the simply inherited traits and quantitative trait loci controlling the polygenic characters in a series of crop plants starting with tomato, maize, and rice. Thus, new crop improvement technique called as molecular breeding started in later part of the twentieth century. On the other hand, genetic engineering made modification of crops for target traits by transferring alien genes, for example, the *Bt* gene from the bacteria *Bacillus thuringiensis*. A large number of genetically modified crop varieties have thus been developed starting with the commercialization of “flavr Savr” tomato in 1994.

Meantime, the manual DNA sequencing methodology of 1977 was being improved with regard to speed, cost-effectiveness, and automation. The first-generation sequencing technology led to the whole genome sequencing of *Arabidopsis* in 2000 and followed by rice in 2002. The next-generation sequencing technologies were available over time and used for sequencing of genomes of many

other model and crop plants. Genomes, both nuclear and organellar, of more than 100 plants have already been sequenced by now, and the information thus generated are available in public database for most of them. It must be mentioned here that bioinformatics played a remarkable role in handling the enormous data being produced in each and every minute. It can be safely told that the “genomics” era started in the beginning of the twenty-first century itself accompanying also proteomics, metabolomics, transcriptomics, and several other “omics” technologies.

Structural genomics has thus facilitated annotation of genes, enumeration of gene families and repetitive elements, and comparative genomics studies across taxa. On the other hand, functional genomics paved the way for deciphering the precise biochemistry of gene function through transcription and translation pathways. Today, genotyping-by-sequencing of primary, secondary, and even tertiary gene pools; genome-wide association studies; and genomics-aided breeding are almost routine techniques for crop improvement. Genomic selection in crops is another reality today. Elucidation of the chemical nature of crop chromosomes has now opened up a new frontier for genome editing that is expected to lead the crop improvement approaches in near future.

At the same time, we will look forward to replacement of genetically modified crops by cisgenic crops through transfer of useful plant genes and atomically modified crops by employing nanotechnology that will hopefully be universally accepted for commercialization owing to their human-friendly and environment-friendly nature.

I wish to emphatically mention here that none of the technologies and tools of plant breeding are too obsolete or too independent. They will always remain pertinent individually or as complementary to each other, and will be employed depending on the evolutionary status of the crop genomes, the genetic resources and genomics resources available, and above all the cost-benefit ratios for adopting one or more technologies or tools. In brief, utilization of these crop improvement techniques would vary over time, space, and economy scales! However, as we stand today, we have all the concepts, strategies, techniques, and tools in our arsenal to practice ‘genome designing’, as I would prefer to term it, of crop plants not just genetic improvement to address simultaneously ‘food, nutrition, energy, and environment security, briefly the FNEE security’, as I introduced the concept in 2013 and have been talking about for the last 5 years at different platforms.

Addressing FNEE security has become more relevant today in the changing scenario of climate change and global warming. Climate change will lead to greenhouse gas emissions and extreme temperatures leading to different abiotic stresses including drought or waterlogging, on the one hand, and severe winter and freezing, on the other hand. It will also severely affect uptake and bioavailability of water and plant nutrients and will adversely cause damage to physical, chemical, and biological properties of soil and water in cropping fields and around. It is also highly likely that there will be emergence of new insects and their biotypes and of new plant pathogens and their pathotypes. The most serious concerns are, however, the unpredictable crop growth conditions and the unexpected complex interactions among all the above stress factors leading to drastic reduction in crop yield and

quality in an adverse ecosystem and environment. Climate change is predicted to significantly reduce productivity in almost all crops. For example, in cereal crops, the decline of yield is projected at 12–15%. On the other hand, crop production has to be increased at least by 70% to feed the alarmingly growing world population, projected at over 9.0 billion by 2050 by even a moderate estimate.

Hence, the unpredictability of crop growing conditions and thereby the complexity of biotic and abiotic stresses warrant completely different strategies of crop production from those practiced over a century aiming mostly at one or the few breeding objectives at a time such as yield, quality, resistance to biotic stresses due to disease–pests, tolerance to abiotic stresses due to drought, heat, cold, flood, salinity, acidity, improved water and nutrient use efficiency, etc. In the changing scenario of climate change, for sustainable crop production, precise prediction of the above limiting factors by long-term survey and timely sensing through biotic agents and engineering devices and regular soil and water remediation will play a big role in agriculture. We have been discussing on “mitigation” and “adaptation” strategies for the last few years to reduce the chances of reduction of crop productivity and improve the genome plasticity of crop plants that could thrive and perform considerably well in a wide range of growing conditions over time and space. This is the precise reason for adopting genomic designing of crop plants to improve their adaptability by developing climate-smart or climate-resilient genotypes.

Keeping all these in mind, I planned to present deliberations on the problems, priorities, potentials, and prospects of genome designing for development of climate-smart crops in about 50 chapters, each devoted to a major crop or a crop group, allocated under five volumes on cereal, oilseed, pulse, fruit, and vegetable crops. These chapters have been authored by more than 250 of eminent scientists from over 30 countries including Argentina, Australia, Bangladesh, Belgium, Brazil, Canada, China, Egypt, Ethiopia, France, Germany, Greece, India, Ireland, Japan, Malaysia, Mexico, New Zealand, Kenya, Pakistan, Philippines, Portugal, Puerto Rico, Serbia, Spain, Sri Lanka, Sweden, Taiwan, Tanzania, Tunisia, Uganda, UK, USA, and Zimbabwe.

There are a huge number of books and reviews on traditional breeding, molecular breeding, genetic engineering, nanotechnology, genomics-aided breeding, and gene editing with crop-wise and trait-wise deliberations on crop genetic improvement including over 100 books edited by me since 2006. However, I believe the present five book volumes will hopefully provide a comprehensive enumeration on the requirement, achievements, and future prospects of genome designing for climate-smart crops and will be useful to students, teaching faculties, and scientists in the academia and also to the related industries. Besides, public and private funding agencies, policy-making bodies, and the social activists will also get a clear idea on the road traveled so far and the future roadmap of crop improvement.

I must confess that it has been quite a difficult task for me to study critically the different concepts, strategies, techniques, and tools of plant breeding practiced over the last 12 decades that also on a diverse crop plants to gain confidence to edit the chapters authored by the scientists with expertise on the particular crops or crop groups and present them in a lucid manner with more or less uniform outline of contents and formats. However, my experience gained over the last 7 years in the capacity of the Founding Principal Coordinator of the International Climate Resilient Crop Genomics Consortium (ICRCGC) was highly useful while editing these books. I have the opportunity to interact with a number of leading scientists from all over the world almost on regular basis. Organizing and chairing the annual workshops of ICRCGC since 2012 and representing ICRCGC in many other scientific meetings on climate change agriculture offered me a scope to learn from a large number of people from different backgrounds including academia, industries, policy-making bodies, funding agencies, and social workers. I must acknowledge here the assistance I received from all of them to keep me as a sincere student of agriculture specifically plant breeding.

This volume entitled *Genomic Designing of Climate-Smart Oilseed Crops* includes eight major crops including Soybean, Oilseed Rape, Groundnut, Sunflower, Flax, Rape and Mustard, Sesame, and Castor Bean. These chapters have been authored by 54 scientists from six countries including Australia, Canada, China, India, Serbia, and USA. I place on record my thanks for these scientists for their contributions and cooperation.

My own working experience on oilseed crops dates back to early 90s in the laboratory of Prof. Thomas C. Osborn in the Department of Agronomy of the University of Wisconsin-Madison. I must confess that this period of about 4 years through working on the Brassicaceae system in his lab and other two labs of his collaborating faculties including Prof. Paul H. Williams in the Department of Plant Pathology and Prof. Jiwan P. Palta in the Department of Horticulture had tailored my mind-set and enriched my expertise and helped me to grow as a science worker. Hence, I have dedicated this book to Prof. Osborn as a token of my respect, thanks, and gratitude.

New Delhi, India

Chittaranjan Kole

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Abbreviations

| | |
|-----------------------|--|
| ^{13}C | Isotope carbon 13 |
| $\Delta^{13}\text{C}$ | Carbon isotope discrimination |
| ABA | Abscisic acid |
| AC | Ash content |
| ADH | Alcohol dehydrogenase |
| AFLP | Amplified fragment length polymorphism |
| ALA | α -linolenic acid |
| ALS | Acetolactate synthase |
| AMOVA | Analysis of molecular variance |
| ANOVA | Analysis of variance |
| APX | Ascorbate peroxidase |
| AQP | Aquaporin |
| ARF | Auxin-response factor |
| ATP | Adenosine triphosphate |
| ATR | Atrazine |
| ATT | Acquired thermotolerance |
| BABA | β -Aminobutyric acid |
| BAC | Bacterial artificial chromosome |
| <i>Bar</i> | Bialaphos resistance gene |
| BC | Back cross |
| BH | Branching habit |
| BIM | Bayesian interval mapping |
| BL | Bayesian LASSO |
| BLAST | Basic local alignment search tool |
| BLUP | Best linear unbiased prediction |
| BPMV | Bean pod mottle virus |
| BRR | Bayesian ridge regression |
| BSA | Bulked segregant analysis |
| <i>Bt</i> | <i>Bacillus thuringiensis</i> |
| C10 | C+1 alleles with capsule open at tip |

| | |
|-----------------|--|
| CaMV | Cauliflower mosaic virus |
| Cas9 | CRISPR-associated 9 protein |
| CAT | Catalase |
| CC | Climate change |
| CDS | Coding DNA sequence |
| CEG | Core eukaryotic genes |
| CGMCP | Centre for Genetic Manipulation of Crop Plants |
| CGRIS | Chinese Genetic Resources Information System |
| CID | Carbon isotope discrimination |
| CIM | Common Information Model |
| CIM | Composite interval mapping |
| CL | Capsule length |
| cM | CentiMorgan |
| CNV | Copy number variant |
| CO ₂ | Carbon dioxide |
| CMS | Cytoplasmic male sterility |
| CN | Capsule number per plant |
| CNN | Capsule node number |
| CNPA | Centro Nacional de Pesquisa de Algodao |
| CNS | Capsule number per stem |
| CNV | Copy number variant |
| CR | Clubroot resistance gene |
| CRISPR | Clustered regularly interspaced short palindromic repeats |
| CRP | Coordinated research project |
| CRR | Charcoal rot resistance |
| <i>cryIAcF</i> | Delta-endotoxin of <i>Bacillus thuringiensis</i> gene (1AcF) |
| <i>cryIEC</i> | Delta-endotoxin of <i>Bacillus thuringiensis</i> gene (1EC) |
| CS | Climate smart |
| CTD | Canopy temperature depression |
| CWR | Crop wild relative |
| CZL | Capsule zone length |
| DAG | Diacylglycerols |
| DALP | Direct amplification of length polymorphism |
| DAP | Days after planting |
| DArT | Diversity arrays technology |
| DAS | Days after sowing |
| DDBJ | DNA Databank of Japan |
| DEG | Differentially expressed gene |
| DGAT | Diacylglycerol acetyltransferase |
| dgatA | Acyl-CoA:diacylglycerol acyltransferase A |
| DH | Doubled haploid |
| DI | Disease index |
| DMO | Dicamba monooxygenase |
| DREB | Dehydration responsive element binding (protein) |
| DREB2A | Drought responsive element binding protein 2A |

| | |
|----------------|--|
| DS | Determinate sesame |
| DSB | Double-stranded break |
| DSF | Days from sowing to flowering |
| dsRNA | Double-stranded RNA |
| Dt | Determinate |
| DTF | Days to flowering |
| Dw | Dwarf |
| <i>Dwf</i> | Dwarfing gene |
| ECP/GR | European Cooperative Programme for Crop Genetic Resources Network |
| EDB | European Brassica Database |
| ELS | Early leaf spot |
| EMF | Embryonic flower |
| EMS | Ethyl methanesulphonate |
| EPA | Eicosapentaenoic acid |
| EPA | Environmental Protection Agency (USA) |
| EPSP | 5-Enolpyruvylshikimate-3-phosphate |
| ESCORENA | European Co-operative Research Network on Flax and other Bast Plants |
| ESPS | 5-Enolpyruvylshikimate-3-phosphate synthase |
| EST | Expressed sequence tag |
| ETI | Effector-triggered immunity |
| F ₂ | Second filial generation |
| FA | Fatty acid |
| FA | Flowers per leaf axil |
| FAO | Food and Agriculture Organization |
| FC | Fiber content |
| FCL | Length of the lateral capsule |
| FCT | Thickness of the lateral capsule |
| FCW | Width of the lateral capsule |
| FDA | Food and Drug Administration (USA) |
| <i>FLC</i> | Flowering Locus C |
| FNEE | Food, nutrition, energy and environment |
| FNI | Fast neutron irradiation |
| FOS | <i>Fusarium oxysporum</i> f.sp. <i>sesami</i> |
| FSD | Fresh seed dormancy |
| FT | Flowering locus T |
| G × E | Genotype × environment |
| GAB | Genomics-assisted breeding |
| GBS | Genotyping-by-sequencing |
| GCA | General combining ability |
| GE | Genetically engineered |
| GEAC | Genetic Engineering Appraisal Committee (India) |
| GEBV | Genome-estimated breeding value |
| GFF | General feature format |

| | |
|-----------|---|
| GFP | Green fluorescent protein |
| GM | Genetically modified |
| GMHRA | Glyphosate acetyltransferase and modified soybean acetolactate synthase |
| GN | Grain number per capsule |
| GO | Gene ontology |
| GP | Gene pool |
| GR | Glyphosate resistant |
| GRD | Groundnut rosette disease |
| GRDC | Grains Research and Development Corporation |
| GRIN | Germplasm Resources Information Network (USA) |
| GRU | Germplasm Resources Unit |
| GS | Genomic selection |
| GSO | Seamless capsule open at tip |
| GSS | Genome survey sequence |
| GUS | β -glucuronidase gene |
| GWAS | Genome-wide association study |
| H | Index of genetic diversity |
| HAAS | Henan Academy of Agricultural Sciences |
| HDR | Homology-directed repair |
| He | Average expected heterozygosity per locus |
| HFC | Height to the first capsule |
| Hi-C | Chromosome conformation capture |
| HOA | High OA |
| HPPD | 4-Hydroxyphenylpyruvate dioxygenase |
| HR | Highly resistant |
| HS | Highly susceptible |
| Hsfs | Heat shock transcription factors |
| HSP | Heat shock protein |
| HSRC | Henan Sesame Research Center |
| I | Shannon's information index |
| IAEA | International Atomic Energy Agency |
| IBC | Institute of Biodiversity Conservation |
| IBPGR | International Bureau of Plant Genetic Resources |
| ICGR-CAAS | Institute of Crop Germplasm Resources, Chinese Academy of Agricultural Sciences |
| ICRISAT | International Crops Research Institute for the Semi-Arid Tropics |
| ID | Indehiscent |
| IFDB | International Flax Database |
| IFVCNS | Institute of Field and Vegetable Crops |
| IL | Internode length |
| ILs | Interspecific lines |
| IND | Improved nondehiscent |
| InDel | Insertion/Deletion |
| INTA | Instituto Nacional de Tecnología Agropecuaria |

| | |
|---------|---|
| IOD | Iodine value |
| IP | Intellectual property |
| IPCC | Intergovernmental Panel on Climate Change |
| ISSR | Inter-simple sequence repeat |
| ITPGRFA | The International Treaty for Plant Genetic Resources for Food and Agriculture |
| KASP | Kompetitive allele-specific polymerase chain reaction |
| KEGG | Kyoto Encyclopedia of Genes and Genomes |
| LD | Linkage disequilibrium |
| LD | Long day |
| LEA | Late embryogenesis abundant |
| LG | Linkage group |
| LIN | Linolenic acid |
| LIO | Linoleic acid |
| LIS-1 | Linum insertion sequence 1 |
| LLS | Late leaf spot |
| LN | Leaf number per plant |
| LncRNA | Long noncoding RNA |
| LOA | Low OA |
| LOD | Logarithm of odds |
| LRR | Leucine-rich repeat |
| MABC | Marker-assisted backcrossing |
| MAGIC | Multiparent advanced generation intercross |
| MAPKK | Mitogen-activated protein kinase kinase |
| MARS | Marker-assisted recurrent selection |
| MAS | Marker-assisted selection |
| MBB | Mexican bean beetle |
| MBPG | Multinational Brassica Genome Sequencing Project |
| MCL | Length of the central capsule |
| MCT | Thickness of the central capsule |
| MCW | Width of the central capsule |
| MDA | Malondialdehyde |
| MG | Maturity group |
| MIM | Multiple interval mapping |
| MLS | Multilateral System |
| MoEF&CC | Ministry of Environment, Forest and Climate Change (India) |
| MR | Moderately resistant |
| MSD | Main stem diameter |
| MSIL | Length of main stem internode |
| MSNN | Node number of main stem |
| MTA | Marker-trait association |
| MTA | Material Transfer Agreement |
| Na | Number of alleles |
| NAM | Nested association mapping |
| NARS | National Agricultural Research System (India) |

| | |
|--------------|--|
| NBPGR | National Bureau of Plant Genetic Resources (India) |
| NBS | Nucleotide-binding site |
| NCBI | National Center for Biotechnology Information |
| NDVI | Normalized difference vegetation index |
| Ne | Effective number of alleles |
| NGS | Next-generation sequencing |
| NHEJ | Non-homologous end joining |
| NIL | Near-isogenic lines |
| NN | Node number |
| <i>nptII</i> | Neomycin phosphotransferase II gene |
| NTG | N-methyl-N'-nitro-N-nitrosoguanidine |
| NUE | Nutrient use efficiency |
| OA | Osmotic adjustment |
| OA | Oxalic acid |
| OC | Oil content |
| OLE | Oleic acid |
| <i>OLP</i> | Osmotin-like protein gene |
| PAGE | Parametric analysis of gene set enrichment |
| PAL | Palmitic acid |
| PAT | Phosphinothricin acetyltransferase |
| PAV | Presence/absence variants |
| PCR | Polymerase chain reaction |
| PE | Paired end |
| PEG | Polyethylene glycol |
| PGR | Pod growth rate |
| PH | Plant height |
| PHB | Polyhydroxybutyrate |
| PIABS | Photosynthetic efficacy index |
| PIC | Polymorphic information content |
| PiHS | Population-based integrated haplotype score |
| <i>Pl</i> | Downy mildew resistance gene |
| PLCP | Papain-like cysteine protease |
| PLH | Potato leafhopper |
| PO | Protein content |
| POX | Peroxidase |
| PPO | Polyphenol oxidase |
| PR | Pathogenesis-related |
| PRH | Bearing height of primary raceme |
| PUFA | Polyunsaturated fatty acids |
| PVE | Phenotypic variation explained |
| QTL | Quantitative trait locus |
| QTLs | Quantitative trait loci |
| R | Resistance gene |
| RAD | Restriction site-associated DNA |
| RAPD | Random amplified polymorphic DNA |

| | |
|---------------|--|
| RCA | <i>R. communis</i> agglutinin |
| <i>RcPAL</i> | <i>Ricinus communis</i> phenylalanine ammonialyase gene |
| <i>RcPEPC</i> | <i>Ricinus communis</i> phosphoenolpyruvate carboxylase gene |
| <i>Rf</i> | Fertility restoration gene |
| RFLP | Restriction fragment length polymorphism |
| RGB | Red, green and blue |
| RGC | Resistance gene candidate |
| RHL | Residual heterozygous line |
| RIL | Recombinant inbred line |
| RNAi | RNA interference |
| ROD | Reduction of density |
| RR-BLUP | Ridge regression best linear unbiased prediction |
| RRGS | Reduced-representation genome sequencing |
| RRS | Reduced-representation sequencing |
| RSA | Root system architecture |
| RSAMPL | Random selective amplification of microsatellite polymorphic locus |
| RSLs | Recombinant substitution lines |
| RT-PCR | Real-time PCR |
| RT-PCR | Reverse transcription PCR |
| RXBS | Rongxian black sesame |
| SAM | Sequence alignment map |
| SAT | Semi-arid tropics |
| SBA | Soybean aphid |
| SbDV | Soybean dwarf virus |
| SBL | Soybean looper |
| <i>SbNHX1</i> | <i>Salicornia brachiata</i> reverse transporter protein gene |
| SC | Sesamin content |
| SCAR | Sequence-characterized amplified region |
| scFv | Single-chain variable fragment |
| SCMR | SPAD chlorophyll meter reading |
| SCoT | Start codon targeted polymorphism |
| SD | Short day |
| SDS | Sudden death syndrome |
| SEA | Singular enrichment analysis |
| SFW | Sesame Fusarium wilt |
| SG | Selective genotyping |
| SGMD | Soybean Genomics and Microarray Database |
| SGP | The Sesame Genome Project |
| SGWG | The Sesame Genome Working Group |
| SHA | Shattering |
| SIM | Simple interval mapping |
| SiNPs | Silicon nanoparticles |
| SLA | Specific leaf area |
| SLAF | Specific length amplified fragment |

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| SLAF-seq | Specific length amplified fragment sequencing |
| SMA | Single marker analysis |
| SMG | Suppressor with morphogenetic effects on genitalia |
| SMV | Soybean mosaic virus |
| SN | Seed number per plant |
| SNC | Seed number per capsule |
| SNP | Single nucleotide polymorphism |
| SOD | Superoxide dismutase |
| SoyGD | Soybean Genome Database |
| SR | Shatter resistant |
| SRAP | Sequence-related amplified polymorphism |
| SSCP | Single-strand conformational polymorphism |
| SSH | Semi-shattering |
| SSR | Simple sequence repeat |
| STE | Stearic acid |
| STF | Days from sowing to flowering |
| STS | Sequence tagged site |
| SUS | Super-shattering |
| TAG | Triacylglycerols |
| TALEN | Transcription activator-like effector nuclease |
| TE | Transpiration efficiency |
| TFL-like | Terminal flower-like |
| TIGR | The Institute for Genomic Research |
| TILLING | Targeting induced local lesions in genomes |
| TIR | Temperature induction response |
| TL | Tip length without the capsule |
| TP | Training population |
| TRAP | Target region amplification polymorphism |
| TRAP | Tartrate-resistant acid phosphatase |
| TSS | Total soluble sugars |
| TSW | Thousand seed weight |
| TSWV | Tomato spotted wilt virus |
| TT | Triazine tolerant |
| TUFGEN | Total Utilization Flax Genomics |
| UGM | Ungrouped matches |
| UPM | The Universidad Politécnica de Madrid |
| UPOV | International Union for the Protection of New Varieties of Plants |
| USDA | United States Department of Agriculture |
| UTRs | Untranslated regions |
| VBC | Velvet bean caterpillar |
| VIR | Vavilov Institute of Plant Industry |
| VNIIMK | All-Russia Research Institute of Oil Crops |
| VPD | Vapor pressure deficit |
| WGR | Whole genome re-sequencing |
| WGS | Whole genome shotgun |

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| WSC | Water soluble carbohydrates |
| WUE | Water use efficiency |
| ZFN | Zinc-finger nucleases |