RESEARCH NOTE

Open Access



Comparative therapeutic index, lethal time and safety margin of various toxicants and snake antivenoms using newly derived and old formulas

Saganuwan Alhaji Saganuwan^{*} 💿

Abstract

Objective: The assessment of clinical efficacy and toxicity is very important in pharmacology and toxicology. The effects of psychostimulants (e.g. amphetamine), psychotomimetics (e.g. Cannabis sativus) and snake antivenoms are sometimes unpredictable even at lower doses, leading to serious intoxication and fatal consequences. Hence, there is need to re-assess some formulas for calculation of therapeutic index, lethal time and safety margin with a view to identifying therapeutic agents with remarkable toxicity potentials.

Results: The therapeutic index formula $[T_1 = 3(W_a \times 10^{-4})]$ was derived from $T_1 = LD_{50}/ED_{50}$ and $ED_{50} = \frac{LD_{50}}{3} \times W_a \times 10^{-4}$. Findings have shown that, therapeutic index is a function of death reversal (s), safety factor (10⁻⁴) and weight of animal (Wa). However, the new safety margin formula $\left[MS = \sqrt[3]{\frac{LT_{50}}{LD_{50}}} \times \frac{1}{ED_{99}}\right]$ derived from $LT_{50} = \frac{LD_{50}}{D_1^2}$ and $MS = \frac{LD_{1}}{ED_{99}}$ shows that safety margin is a function of cube root of ratio between LT_{50} and LD_{50} and ED_{100th} . Concentration (k) of toxicant at the receptor $\left[K = \sqrt[3]{\frac{LT_{50}}{LD_{50}}} \times \frac{1}{T^n}\right]$ derived from $D_1 \times T^n = K$ and $LD_1 = \sqrt[3]{\frac{LT_{50}}{LD_{50}}}$ shows that therapeutic index, lethal time and safety margin is a function of drug or toxicant concentration at the receptor, the drug-receptor interaction and dose of toxicant or drug administered at a particular time.

Keywords: Therapeutic index, Safety margin, Efficacy, Toxicity, Weight, Reversal, Drug

Introduction

The important assessment of clinical efficacy and toxicity of drugs and chemicals cannot be overemphasized. Dose–response relationship can identify hazardous substance [1] with toxic or beneficial effect over time [2]. Examples of such substances are snake and scorpion venoms, plant extract, drug and chemicals that cause different kinds of toxic effects on various body systems [3–11]. Attempts were made to use structures of therapeutic

*Correspondence: pharn_saga2006@yahoo.com

Department of Veterinary Pharmacology and Toxicology, College of Veterinary Medicine, Federal University of Agriculture, P.M.B. 2373, Makurdi, Benue, Nigeria agents to predict their toxic effects [12, 13]. The predictive toxicity was based on active sites of compounds, such as the number of aromatic rings in polycyclic hydrocarbons, the number of chlorine atoms in chlorinated hydrocarbons and the number of hydroxyl groups. Such predictions have made some success but far from perfect [14–16]. In the past, many animals (40–100) were used for safety study until OECD introduced up-and-down procedure, limiting the number of animals for the study to 5-20 [17–19]. The use of large number of animals for determination of median lethal dose (LD₅₀) has been discouraged worldwide [20]. Hence, based on the principle of R3 (Reduction, Refinement and Replacement),



© The Author(s) 2020. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/

the number of animals for $\rm LD_{50}$ determination has been reduced to 2–6 animals [6]. The inherent variability, lack of predictive validity and lack of reliability of experimental animal models and conflicting clinical reports on therapeutic indices, safety margins and lethal times of some psychostimulants, psychotomimetics and snake antivenoms have necessitated the need to revise the current therapeutic index and safety margin formulas.

Main text

Methodology

Literatures from journals published by Elseviers, Springer, Springer Nature, Sage, Tailor and Francis, Wiley and other publishers were searched for reports on LD_{50} of amphetamine, dextroamphetamine, lysergic acid diethylamide, potassium permanganate, Abrus precatorius and tetrahydroxycannonbinol in dog, rabbit, mouse, human and rat, respectively. The d-tubocurarine has been reported to counteract their effects to some levels. However the reported LD_{50} and ED_{50} of some snake venoms and antivenoms were used for the study. The formulas used in determination of LD_{50} for snake venom with effective dose fifty (ED_{50}) divided by the denominator (3) as well as other related formulas, were incorporated into derived therapeutic index, lethal time and margin safety formulas [3–27]. The derivations are as follow:

Previously established formulas

Therapeutic index (TI)
$$= \frac{TD_{50}}{ED_{50}} = \frac{LD_{50}}{ED_{50}}$$
 (1)

Margin of safety
$$=$$
 $\frac{TD_1}{ED_{99}} = \frac{LD_1}{ED_{99}}$ (2)

Effective dose fifty (ED₅₀) for snake antivenom

$$=\frac{LD_{50}}{3} \times \mathrm{Wa} \times 10^{-4} \tag{3}$$

Newly derived formulas

$$If LD_{50} = TI \times ED_{50} \tag{4}$$

Substitute for LD_{50} in Eq. (3)

$$ED_{50} = \frac{TI \times ED_{50}}{3} \times Wa \times 10^{-4}$$
$$= \frac{\frac{TI \times ED_{50}}{3}}{ED_{50}} = Wa \times 10^{-4}$$
(5)

$$= \frac{TI \times ED_{50}}{3} \times \frac{1}{ED_{50}} = \text{Wa} \times 10^{-4}$$
(6)

$$=\frac{TI}{3} = Wa \times 10^{-4} \tag{7}$$

$$= TI = 3(Wa \times 10^{-4})$$
(8)

TI=Therapeutic index; LD_{50} =Median lethal dose; ED_{50} =Median effective dose; Wa=Weight of animal; 10^{-4} =Safety factor.

Integration of lethal time with margin safety formula

Median lethal time (LT₅₀) =
$$\frac{LD_{50}}{D^p}$$
 (9)

$$LD_{50} = \frac{LT_{50}}{LD_1^p}$$
(10)

whereas $LD_1 = Dose$ that kills one animal; p = Exponent (1/3).

Remember Eq. (2) for margin of safety (MS) = $\frac{LD_1}{FD_{00}}$.

$$LD_1 = MS \times ED_{99} \tag{11}$$

$$LD_1^P = \frac{LT_{50}}{LD_{50}}$$
(12)

But p = 1/3. Therefore

=

$$LD_1^{\frac{1}{3}} = \frac{LT_{50}}{LD_{50}} \tag{13}$$

$$LD_1 = \sqrt[3]{\frac{LT_{50}}{LD_{50}}}$$
(14)

So, integrate Eqs. (11) and (14)

$$LD_1 = MS \times ED_{99} = \sqrt[3]{\frac{LT_{50}}{LD_{50}}}$$
(15)

Hence,

$$MS = \sqrt[3]{\frac{LT_{50}}{LD_{50}}} \times \frac{1}{ED_{99}}$$
(16)

Therefore, margin of safety is a function of cube root of ratio between LT_{50} and LD_{50} and one-hundredth of ED.

| Toxicant | Experimental | ental | Weight | LD ₅₀ (mg/kg) | LD ₅₀ (mg/kg) ED ₅₀ (mg/kg) LT ₅₀ | LT ₅₀ | LD ₁ (mg/kg) | LD ₁ (mg/kg) ED ₉₉ (mg/kg) T ₁ (Control) T ₂ | T ₁ (Control) | Т2 | MS ₁ (Control) MS ₂ | MS ₂ |
|------------------------------|--------------|----------------------|---------------|--------------------------|--|------------------|-------------------------|--|--------------------------|-------|---|-----------------|
| | Animal | Animal Antidote | of animal(kg) | | | | | | | | | |
| Amphetamine | Dog | D-Tubocucarine 10 | 10 | 5.9 i.v | 2.0 | 2.9 s | 0.118 | 3.96 | 2.95 | 3.0 | 0.03 | 0.13 |
| Dextroamphetamine | Dog | D-Tubocucarine 10 | 10 | 11.8 i.v | 3.9 | 7.3 s | 0.236 | 7.72 | 3.02 | 3.0 | 0.03 | 0.11 |
| Methamphetamine | Dog | D-Tubocucarine 10 | 10 | 11.8 i.v | 3.9 | 7.3 s | 0.236 | 7.72 | 3.02 | 3.0 | 0.03 | 0.11 |
| Lysergic acid diethylamide | Rabbit | D-Tubocucarine 1.8 | 1.8 | 0.3 i.v | 0.02 | 0.6 s | 0.006 | 0.04 | 15.0 | 0.54 | 0.15 | 13.2 |
| Potassium permanganate Mouse | Mouse | D-Tubocucarine 0.02 | 0.02 | 1499.7 oral | 1.0 | 1.3 h | 30.0 | 1.98 | 1499.7 | 0.003 | 15.1 | 1.58 |
| Abrus precatorius | Human | Human D-Tubocucarine | 60 | 197 i.m | 394 | 5.2 min | 3.94 | 780.1 | 0.5 | 18.0 | 0.005 | 0.002 |
| Tetrahydroxy cannabinol | Rat | D-Tubocucarine 0.15 | 0.15 | 29 i.v | 0.15 | 24.2 min | 0.58 | 0.30 | 193.3 | 0.05 | 1.9 | 12.2 |
| | | | | | | | | | | | | |

| ğ |
|---------|
| £ |
| Ð |
| Ĕ |
| pla |
| ld bl |
| ar |
| als |
| ij. |
| em |
| ÷ |
| b ər |
| ē |
| is of s |
| so |
| argin: |
| arç |
| Ĩ |
| Ę |
| afe |
| ŝ |
| Ĕ |
| ŝ |
| i. |
| pd |
| 5 |
| euti |
| pe |
| era. |
| ĥ |
| Ξ. |
| e |
| Tabl |
| Ë |

Integration of time of exposure with toxic or lethal dose

Concentration of toxicant (K) =
$$D \times T^n$$
 (17)

whereas D=Daily dose; T=Time of exposure; K=constant which is the concentration of toxicant causing toxicity; n=power of exponent.

Therefore,

$$D = \frac{T^n}{K}$$
(18)

But if D can kill one animal as shown in Eq. (15) and related to Eq. (18), it would be referred to as TD_1

$$\therefore LD_1 = TD_1 = \sqrt[3]{\frac{LT_{50}}{LD_{50}}} = \frac{T''}{K}$$
(19)

Therefore,

$$K = \sqrt[3]{\frac{LT_{50}}{LD_{50}}} \times \frac{1}{T^n}$$
(20)

The formulas were used to calculate LD_{50} , ED_{50} , LT_{50} , LD_1 , ED_{99} , therapeutic index (TI) and safety of margin for all the reported antidotes for snake envenomation, *Abrus precatorius*, lysergic acid diethylamide, tetrahydroxy-cannabinol, amphetamine, methamphetamine, dextro-amphetamine and potassium permanganate poisoning. All the LT_{50} in hour and minute should be converted to second.

Results

The LD_{50} , ED_{50} , LT_{50} , LD_1 , ED_{99} , dose of toxicants, therapeutic index and safety margin of amphetamine, dextroamphetamine, methamphetamine, lysergic acid diethylamide, tetrahydroxycannabinol, potassium permanganate and *Abrus precatorius* are presented in Table 1.The LD_{50} , ED_{50} , LD_1 , ED_{99} , therapeutic index and safety margin of snake venoms and antivenoms are presented in Table 2.

Discussion

Side effects, adverse drug reactions, untoward effects, side toxicity and idiosyncratic effects associated with drugs may be due to normal dose, under dose or drug over dose [5, 28]. The calculated therapeutic index of amphetamine (2.95), dextroamphetamine and amphetamine (3.02) using the previously established formula as compared to therapeutic index of 3.0 for the three drugs using the new formula show that, the newly developed formula can be used for calculation of therapeutic index of some psychomimetic and psychotomimetic drugs. However, the previously established formula yielded very

high therapeutic index for LSD (15.0), potassium permanganate (1499.7), Abrus precatorius extract (0.5) and tetrahydroxycannabinol (193.3) as compared to 0.54, 0.003, 18.0 and 0.05 yielded by the newly developed formula, respectively. The findings agree with the report indicating that the conventional formula for calculation of therapeutic index is not a truthful measure of safety of a drug in clinical setting [10]. The low therapeutic index of 0.05 for tetrahydroxycannabinol agrees with the report that most biologically active molecules of Cannabis sativa have no therapeutic uses [24]. Very low therapeutic index (0.003) of potassium permanganate yielded by the newly derived formula agrees with the report indicating that the chemical is highly toxic [4]. The associated toxicity signs are rapid shallow respiration, diarrhea, gastroenteritis, liver and kidney damage and death.

The low therapeutic index (0.5) of A. precatorius shows that the plant is very toxic. This may be due to presence of toxic principle called abrin [29]. However, the relatively high therapeutic index of 18.0 calculated using the new formula agrees with the report that the plant may have some degrees of therapeutic safety [21]. The therapeutic index for LSD using the conventional (15) and new formula (0.54) corroborates the findings that the pharmacology of LSD is complex and its mechanism of actions is not understood [25]. A. precatorius extract is more toxic when given intraperitoneally as compared to oral route [11]. However d-tubocurarine can alleviate toxicity effects of amphetamine, dextroamphetamine, methamphetamine [26], Abrus precatorius [30], tetrahydroxycannabinol [31], potassium permanganate [32], and lysergic acid diethylamide [33].

The dose-toxicity response pattern in graded fashion may culminate in LD₅₀ and could be counteracted by therapeutic dose 50. This explains individual variation of susceptibility to doses of toxicants as proven by low therapeutic index (0.0007) of Micrarus fulvius antivenom (Table 2) as compared to high toxicity potential of M. *fulvius* venom [11]. The low to high therapeutic indices of all the snake antivenoms in the present study indicate that, treatments of snake envenomation is by toxin neutralization, using specific antidotes for specific snakes [8]. The obtaining of therapeutic index (0.006–1499.7) in the present study disagrees with the report of Stanley indicating that therapeutic index could be 33,000:1 [34]. Therefore one vial of the relevant antivenom is sufficient for the circulating venom, but recovery time may be delayed, because many clinical and laboratory effects are not reversed immediately [35]. Hence clinical trials of antivenoms are potentially more important and useful [36]. Pain score of more than two (2) requires additional antivenom and patient should be frequently assessed [37] for improvement. Therefore, there is need for clinicians

Table 2 Therapeutic indices and safety margins of the antidotes of some snake venoms

| Species | LT ₅₀ (hr) | LD ₅₀ (mg/kg) | ED ₅₀ (mg/kg) | LD ₁ (mg/kg) | ED ₁ (mg/kg) | ED ₉₉ (mg/kg) | T ₂ | MS ₁ (Control) |
|--|-----------------------|--------------------------|--------------------------|-------------------------|-------------------------|--------------------------|----------------|---------------------------|
| Crotalus durissus terrificus (Tropical rattle snake) | 0.43 | 0.13 | 4.02 | 0.026 | 0.0804 | 7.96 | 0.69 | 0.003 |
| Crotalus scutulatus scutulatus (Mojave rattle snake) | 0.52 | 0.17 | 4.40 | 0.034 | 0.088 | 8.71 | 0.85 | 0.004 |
| Crotalus horridus africaudatus (Cane- brake rattle snake) | 1.61 | 0.92 | 7.72 | 0.184 | 0.1544 | 15.29 | 4.60 | 0.012 |
| Crotalus adamanteus (Eastern dia- mond back rattle snake) | 2.08 | 1.35 | 8.77 | 0.27 | 0.1754 | 17.36 | 6.75 | 0.016 |
| <i>Crotalus durissus durissus</i> (Central American rattle snake) | 2.51 | 1.79 | 9.64 | 0.358 | 0.1928 | 19.09 | 8.95 | 0.019 |
| Agkistrodon piscivorus piscivorus (Eastern cotton mouth) | 3.84 | 3.38 | 11.91 | 0.676 | 0.2382 | 23.58 | 16.90 | 0.029 |
| <i>Croatlus viridus helleri</i> (Southern pacific rattle snake) | 3.92 | 3.48 | 12.03 | 0.696 | 0.2406 | 23.82 | 17.40 | 0.029 |
| Crotalus molossus molossus (Northern black-tailed rattle snake) | 4.17 | 4.42 | 13.03 | 0.884 | 0.2606 | 25.80 | 22.10 | 0.034 |
| <i>Sistrurus miliarius barbourin</i> (Southern pygmy rattle snake) | 4.91 | 4.87 | 13.45 | 0.974 | 0.269 | 13.45 | 24.35 | 0.072 |
| Agkistrodon contortrix contortrix (Southern Copperhead) | 5.0 | 4.99 | 13.56 | 0.998 | 0.2712 | 13.56 | 24.95 | 0.073 |
| Crotalus horridus horridus (Timber rattle snake) | 5.85 | 6.32 | 14.61 | 1.264 | 0.2922 | 14.61 | 31.60 | 0.087 |
| Micrarus fulvius | 0.79 | 0.32 | 4.77 | 0.064 | 0.0954 | 9.44 | | 0.007 |

Consroe et al. established baseline LD₅₀ values for crotalid anti venin FAB prepared from sheep immune globulin (IgG)

and laboratory toxicologists to improve therapeutic knowledge of snake envenomation [38]. Cardio-respiratory distress, coagulopathy and swelling in the first hours of admission are poor prognostic signs associated with weak therapeutic response to snake envenomation [39]. Effective dose 99 (7.96-23 mg/kg) agrees with the report indicating that, there are many recommended therapeutic interventions, which are ineffective and may be harmful [40]. Therefore, more purified and specific antivenoms are required to avoid post-treatment reactions [41], suggesting that polyvalent antivenom may be less effective against neurotoxic snake bite [42], translating to 1:2 required 30 vials of antivenom [43]. Paraspecific neutralization of snake venom by antivenom could induce coagulopathy in the affected patients [44]. Efficient, safety and thermal stability have been reported for freeze-dried trivalent antivenom for snake bites in larger phase III trial [45]. Russell's viper injects 63-70 mg of venom during the first bite and each vial of polyvalent antivenom neutralizes 6 mg of the venom, 8-10 vials are required in majority of the cases [46]. Neither antivenom nor time of its administration affects venom-induced coagulopathy [35]. Low dose of 20-220 ml reduced the hospital stay as compared to 40-550 ml dose, suggesting that the lower the dose of snake antivenom the more effective the antivenom. Fatality rates of 15.4% and 17.6% for 2 and 4 vials of antivenom as compared 223% have been reported [47]. Protection of snake antivenom against *Echis ocellatus* is 21–99% in Nigeria [36]. Hence, the number of animals for similar study can be reduced [5]. The LT_{50} (0.065–24.2 min) of all the animal, plant and chemical toxins in the present study shows the importance of dose-time-response relationship in identification of hazards [1].

Conclusion

The newly derived formulas yielded low and safer values for therapeutic indices and standard safety margins of drugs, toxicants, venoms, antivenom and other xenobiotics. But the safety of therapeutic agent is dependent on dose, lethal time, body weight, frequency and time of administration and safety factor of the drug.

Limitations

The calculations were based on the findings from experiments conducted in various laboratories across the globe. All the lethal times have to be converted to seconds. The derived formulas were applied on different species of toxic animals and plants.

Abbreviations

 LT_{50} : Median lethal time; LD_{50} : Median lethal dose; ED_{50} : Median effective dose; LD_1 : Lethal dose per one animal; ED_1 : Effective dose per one animal; ED_{99} : Effective dose per 99 animal; LD_1 : Lethal dose 1; T_1 : Therapeutic index for the

Acknowledgements

I sincerely thank Williams Yusuf of Federal University of Agriculture Makurdi and Kehinde Ola Emmanuel of National Open University all in Nigeria for typing the work.

Authors' contributions

SAS designed and carried out the study, analyzed the data, wrote and proof read the manuscript. The author read and approved the final manuscript.

Funding

The study was carried out using my monthly emoluments.

Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Ethics approval and consent to participate

Not applicable, because neither animals nor humans were used for the study; the data were generated from earlier few data established in clinics and laboratories.

Consent to publish

Not applicable.

Competing interest

The author declares that he has no competing interest.

Received: 15 May 2020 Accepted: 10 June 2020 Published online: 16 June 2020

References

- Tennekes HA. The importance of dose-time-response relationships for hazard identification and limitation of animal experiments. Open Acc J Toxicol. 2017;1(5):1–4.
- Druckrey H. Quantitative Grundiagen der Krebserzeugung. Klin Wochenschr. 1943;22:532.
- Saganuwan SA. A modified arithmetical method of Reed and Muench for determination of a relatively ideal median lethal dose. Afr J Pharm Pharmacol. 2011;5(12):1543–6.
- Saganuwan SA. Acute toxicity studies of potassium permanganate in Swiss albino mice. Nig J Physiol Sci. 2008;23(1–2):31–5.
- 5. Saganuwan SA. Arithmetic geometric-harmonic (agh) method of rough estimation of median lethal dose (LD_{50}) using up-and-down procedure. J Drug Metab Toxicol. 2015;6(2):1–3.
- Saganuwan SA. Arithmetic rough estimation of median lethal dose (LD₅₀) using up-and-down procedure. Toxicol Lett. 2014;229:5127.
- Saganuwan SA, Onyeyili PA. Comparative toxicology effects of orally and intraperitoneally administered aqueous extracts of Abrus precatorius leaf in Mus musculus. Herba Polon. 2011;57(3):32–44.
- Saganuwan SA. Calculation of effective dose fifty of antivenom for American pit viper envenomation. Comp Clin Pathol. 2018;27:1321–5.
- Saganuwan SA. Determination of median effective dose fifty (ED₅₀) of scorpion antivenom against scorpion envenomation using a newly developed formula. Animal Mod Exp Med. 2018;1:228–34.
- Saganuwan SA. Principles of Pharmacological Calculations. Zaria: Ahmadu Bello University Press; 2012. p. 529.
- 11. Saganuwan SA. The New algorithm for calculation of median lethal dose (LD_{50}) and effective dose fifty (ED_{50}) of Micrarus fulvius venom and antivenom in mice. Int J Vet Sci Med. 2016;4:1–4.
- Golberg L. structure-activity correlation as a predictive tool in toxicological study. Fundamentals, methods, and application, hemisphere. Washington: OSA; 1983.
- Rosenkranz HS, Takihi N, Klopman G. structure activity—based predictive toxicity: an efficient and economical method for generating non-cosmeric data bases. Nutagenesis. 1991;6:391–4.

- 14. Saganuwan SA. Proxicam: source for Synthesis of Central Nervons (CNS) acting drugs. CNSAMC. 2017;17(2):135–40.
- Saganuwan SA. Biomedical application of polymers: a case study of non-CNS drugs becoming cns acting drug. CNSAMC. 2018;18(1):32–8.
- Saganuwan SA. Chemistry and effects of bramistem acting drugs. CNSAMC. 2019;19(3):180–6.
- OECD. OECD Guideline for the Testing of Chemicals, Test Guideline 401, Acute Oral Toxicity. OECD Paris (1987) http://www.oecd.org//ehs/test/ health.htm.
- OECD. Gindan Document on Acute Oral Toxicity. Environmental Health and Safety Minograph Series on Testing and Assessment No. 24, 2000.
- OECD. Harmonized Integrated Classification System for Human Health and Environmental Hazards of Chemical Substances and Mixtures. ENV/ JM/MONO, OECD Paris 2001:6. http://www.oecd.org/ehs/class/hcl6.htm.
- 20. Saganuwan SA. Toxicity studies of drugs and chemicals in animals: an overview. Bulg J Vet Med. 2017;20(4):291–318.
- Saganuwan SA, Onyeyili PA. The paradox of human equivalent dose formula: a canonical case study of Abrus precatorius aqueous leaf extract in monogastric animals. Mac Vet Rev. 2016;39(1):23–32.
- Saganuwan SA. The use of body surface area for determination of age, body weight, urine creatinine clearance: the reliable canonical method of assessing renotoxicity in animals. Comp Clin Pathol. 2018;2018:1–6.
- 23. Saganuwan SA. Toxicology: the basis for development of antidotes. Toxicology. 2015;1(1):1–2.
- 24. Gabriel G. Toxicology and pharmacology of Cannabis sativa with special references to Δ 9-THC. UNODC 2018: 1–33.
- Passie T, Halperin JH, Stichtenoth DO, Enrich HM, Hintzen A. The pharmacology of lysergic acid diethylmide: a review. CNS Neurosci Therapeut. 2008;14:295–314.
- 26. Zalis EG, Kaplan G, Lundberg GD. Acute lethality of the amphetamines in dogs and its antagonism by curate. Exp Biol Med. 1965;118(2):557–61.
- 27. Timbrell JA. Principles of Biochemical Toxicology. Informa Healthcare: New York; 2009. p. 453.
- Du W, Chong S, Mclachlan AJ, Luo L, Glasgon N, Gnjidic D. Adverse drug reactions due to opioid analgesic use in New South Wales, Australia: a spatial-temporal analysis. BMC Pharmacol Toxicol. 2019;20(55):1–11.
- 29. Hart M. Jecquirity bean poisoning. N Engel J Med. 1963;268:885–6.
- Karthikeyan A, Amalnath SD. Abrus precatorius poisoning: a retrospective study of 112 patients. Ind J Critical Care Med. 2017;21(4):224–5.
- Whalley BJ, Lin H, Bell L, Hill T, Patel A, Gray RA, Roberts CE, Devinsky O, Bazelot M, Williams CM, Stephens GJ. Species-specific susceptibility to cannabis-induced convulsions. BJP. 2019;176:1506–23.
- Cameron CB, Gregory GA, Rudolph AM, Heymann MA. Cardiovascular effects of d-tubocurarine and pancuronium in newborn lambs during normoxia and hypoxia. Pediatr Res. 1986;20(3):246–52.
- Burish MJ, Thoren KL, Madou M, Toossi S, Shah M. Hallucinogens causing seizures? A case report of the synthetic amphetamine 2,5-dimethoxy-4-chloroamphetamine. Neurohospitalist. 2015;5(1):32–4.
- 34. Stanley TH. Anesthesia for 21st century. Proceeding. 2000;13(1):7–10.
- Isbister GK, Brown SGA, Page CB, McCoubrie DL, Greene SL, Burkley NA. snake bite in Australia: a practical approach to diagnosis and treatment. Med J Aust. 2013;199(11):763–8.
- 36. Williams DJ, Habib AG, Warrell DA. Clinical studies of the effectiveness and safety of antivenoms. Toxicon. 2018;150:1–32.
- Kang S, Noon J, Chun B. Does the traditional snake bite seventy score correctly classify envenomated patients? Clin Exp Emerg Med. 2016;3(1):34–40.
- Warrell DA, Gutierrez JM, Calvete JJ, Williams D. New approaches and technologies of venomics to meet challenge of human envenoming by snakebites in India. Indian J Med Res. 2013;138:38–59.
- Dadpour B, Shatahi A, Monzavi SM, Zavav A, Afshari R, Khoshdel AR. Snakebite prognostic factors: leading factors of weak therapeutic response following snakebite envenomation. APJMT. 2012;1(1):27–33.
- 40. Peterson ME. Snakebite: coral snake. Clin Techn Small Anim Pract. 2006;21:83–6.
- Morais VM, Massaldi H. Snake antivenoms: adverse reactions and production technology. J Venom Anim Toxins Med Trop Dis. 2009;15(1):2–18.
- Pore SM, Ramanand SJ, Patil PT, Gore AD, Pawar MP, Gaidhankar SC, Ghanghas RR. A retrospective study of use of polyvalent anti-snake venom and risk factors for mortality from snakebite in a tertiary care setting. Indian J Pharmacol. 2015;47(3):270–4.

- 43. Warrel DA. Snakebite. Lancet. 2010;375:77-88.
- 44. Ainsworth S, Slagboom J, Alomran N, Pla D, Alhamdi Y, King SI, Balton FMS, Gutierreg JM, Vonk FJ, Toh CH, Calvete JJ, Kool J, Harrison RA, Casewell NR. The paraspecific neutralization of snake venom induced coagulopathy by antivenoms Commun Biol. 2018;1(34):1–14.
- Mendonca-da-Silva I, Tavares AN, Sachett J, Sardinha JF, Zaparolli L, Santos MFG, Lacerda M, Monteiro WM. Safety and efficacy of a freeze-dried trivalent antivenom for snake bites in the Brazilian Amazon: an open randomized controlled phase 116 clinical trial. Plos Neglet Trop DIS. 2017;11(11):1–21.
- Hazra A. Poisonous snakebites in India. Community Dev Med Unit Ration Drug Bull. 2003;30:1–12.
- Ramos-Cerrillo B, deRoodt AR, Chippaux JP, Olgivin L, Casasola A, Guzman G, Paniagua-Solis J, Alagon A, Slock RP. Characterization of a new polyvalent antivenom (Antivipnyn African) against African vipers and elapids. Toxicon. 2008;52(8):881–8.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

