

RESEARCH ARTICLE

Open Access

Low prevalence of *Neospora caninum* and *Toxoplasma gondii* antibodies in dogs in Jilin, Henan and Anhui Provinces of the People's Republic of China

Yurong Yang^{1,2}, Qiongfang Zhang¹, Yangguang Kong¹, Yuqing Ying², Oliver Chun Hung Kwok², Hongde Liang^{1*} and Jitender Prakash Dubey²

Abstract

Background: *Neospora caninum* and *Toxoplasma gondii* are important pathogens of worldwide distribution. *N. caninum* is a major cause of abortion in cattle and dogs are main reservoirs because they excrete the environmentally resistant oocysts. Toxoplasmosis is a worldwide zoonosis and dogs are considered as sentinels for this parasite because of their close contact with people and cats; additionally dog meat is also used for human consumption in China. The aim of the present study was to assess the prevalence of *N. caninum and T. gondii* infection in dogs from China. A total of 425 countryside dog hearts in Jilin, Henan and Anhui provinces of the People's Republic of China were collected from slaughter houses in two batches; the first batch of 96 in October 2013, and the second batch of 329 in April 2014. Serum samples extracted from 96 dog hearts were tested for antibodies to *N. caninum* and from 425 dog hearts were tested for *T. gondii* antibodies in the modified agglutination tests (cut-off 1:25 for both), using respective antigens.

Results: Antibodies to *N. caninum* were 6 of 96 (6.25%) of dogs with titers of 1:25 in 2, 1:50 in 3, and 1:100 in 1. All seropositive dogs were more than 1 year old. Antibodies to *T. gondii* were found in 35 of 425 (8.24%) dogs with titers of 1:25 in 15, 1:50 in 14; and 1:100 in 6.

Conclusion: The results of the present study indicated low prevalence of *N. caninum* and *T. gondii* antibodies in dogs of China, compared with Europe and America. Identification of the risk factors that underlie these differences may help prevention of neosporosis and toxoplasmosis. This is the first report of *N. caninum* infection in dogs from China.

Keywords: *Neospora caninum, Toxoplasma gondii,* Seroepidemiology, Dogs, China, Modified agglutination test, *Neospora* agglutination test

Background

Neospora caninum and Toxoplasma gondii are related coccidians that until 1988 were considered the same organism [1]. N. caninum is now considered the most important abortifacient for cattle worldwide, including China [2,3]. Dog, wolf, coyote and dingo are the definitive hosts for N. caninum that shed environmentally

resistant oocysts. *Toxoplasma gondii* infection in dogs is important for following reasons. Dogs can be infected through contact with the *T. gondii*, which may be acquired from rooting in infected soil or from ingesting cat feces or from eating raw meat. Dogs can also mechanically transmit *T. gondii* oocysts to humans [4]. In China, dogs serve as food animals, the consumption of undercooked meat containing *T. gondii* tissue cysts can be a supplementary health risk to consumers.

Currently, there is no report of isolation *T. gondii* from dog in China, and little is known of *N. caninum* infections

Full list of author information is available at the end of the article



^{*} Correspondence: hdliang12@163.com

¹Laboratory of Veterinary Pathology, College of Animal Science and Veterinary Medicine, Henan Agriculture University, Zhengzhou 450002, PR China

in dogs in China. The objective of present study was to determine the seroprevalence of *N. caninum and T. gondii* infections in dogs from China, and to attempt isolate *T. gondii*.

Methods

Naturally infected dogs

A total of 425 countryside dogs were sampled from the slaughter house (Table 1). These countryside dogs in China are part of the farmer's household; they were mainly used for guarding. Their diet includes boiled rice, discarded raw food animal tissues and whatever dogs can forage. These dogs were sold for food. Hearts of dogs were selected for the present study because serum could be obtained from the heart. The first batch samples (96 dog hearts) were collected in October 2013; 50 dogs were younger than 1 year, and 46 were older than 1 year. Second batch samples (329 dog hearts) were collected in April 2014; they were older than 1 year. Blood was collected from heart 1 day (second batch) or three days (first batch) after slaughter, centrifuged at 2000 x g for 10 min, and sera were separated. Hearts samples from first batch were transported by air as part of personal baggage of the senior author from China to the Animal Parasitic Diseases Laboratory (APDL), United States Department of Agriculture, Beltsville, Maryland, USA within one week of killing. During interim, samples were kept cold. Permission papers from USA and China were obtained before transportation.

Ethical aspects

This study was approved by the institutional animal use protocol committee of the United States Department of Agriculture and the Henan Agriculture University, China.

Climatic conditions

The climate of Henan (Latitude 34.90°N, Longitude 113.50°E) is humid subtropical whereas the climate of Anhui province (Latitude 31.86°N, Longitude 117.28°E) is semi-humid, monsoonal, and the climate of Jilin province (Latitude 43.70°N, Longitude 126.20°E) is humid continental climate, winters are long (lasting from November to March), cold, and windy.

Serologic examination

Serum samples from 425 dogs were tested for antibodies to *T. gondii* using the modified agglutination test (MAT) [5]. Serum samples from first batch samples (96 dogs) were also tested for *N. caninum* antibodies by the *Neospora* agglutination test (NAT) [6]. A titer of 1:25 was considered as indicative of exposure to both parasites. Sera were diluted with phosphate-buffered saline and tested 1:25, 1:50, 1:100 and 1:200 dilutions for both parasites. In addition, for *T. gondii*, sera were also tested at 1:10 dilution.

Table 1 Seroprevalence of *Toxoplasma gondii* and *Neospora caninum* infection in dogs in Jilin, Henan and Anhui Provinces

| Chavastavisti | Dogs | Positive | no. in diffe | rent titres | | Total positive | Seroprevalence | 050/ 61 | |
|-----------------|------------|----------|--------------|-------------|-------|--------------------|----------------|-------------|--|
| Characteristics | tested no. | 1:10 | 1:25 | 1:50 | 1:100 | no. (cut-off 1:25) | (%) | 95% CI | |
| Toxoplasma gone | dii | | | | | | | | |
| Region | | | | | | | | | |
| Henan | 31 | 1 | 1 | 0 | 0 | 1 | 3.23 | <0.01-17.58 | |
| Anhui | 22 | 0 | 0 | 0 | 0 | 0 | 0 | - | |
| Jilin | 372 | 5 | 14 | 14 | 6 | 34 | 9.14 | 6.59-12.53 | |
| Age (years) | | | | | | | | | |
| ≤1 | 50 | 0 | 1 | 0 | | 1 | 2.00 | <0.01-11.47 | |
| >1 | 375 | 6 | 14 | 14 | 6 | 34 | 9.07 | 6.53-12.43 | |
| Total | 425 | 6 | 15 | 14 | 6 | 35 | 8.24 | 5.95-11.26 | |
| Neospora caninu | m | | | | | | | | |
| Region | | | | | | | | | |
| Henan | 31 | 0 | 0 | 0 | 0 | 0 | 0 | - | |
| Anhui | 22 | 0 | 0 | 0 | 0 | 0 | 0 | - | |
| Jilin | 43 | 0 | 2 | 3 | 1 | 6 | 13.95 | 6.18-27.64 | |
| Age (years) | | | | | | | | | |
| ≤1 | 50 | 0 | 0 | 0 | 0 | 0 | 0 | - | |
| >1 | 46 | 0 | 2 | 3 | 1 | 6 | 13.04 | 5.74-26.04 | |
| Total | 96 | 0 | 2 | 3 | 1 | 6 | 6.25 | 2.63-13.23 | |

Table 2 Prevalence of T. gondii antibodies in dogs in People's Republic of China

| Map region | Provinces/ cities | Year tested | Dog type | No. tested | No. positive | % positive | Serologic test | No. with terminal titer of | | | | | Reference |
|---------------|----------------------|----------------|-------------------|---------------|-----------------|------------|-------------------------|----------------------------|---------|---------|-----------|------|---------------|
| | | | | | | | | <25 | 25 ~ 49 | 50 ~ 99 | 100 ~ 199 | ≥200 | кетегепсе |
| I | Beijing | 1999-2005 | Pet | 534 | 128 | 24.0 | ELISAª/LAT ^b | unknown | | | | | [22] |
| II | Heilongjiang | 2010-2011 | Stray/pet | 124 | 14 | 11.3 | IHA^b | unknown | | | | | [23] |
| III | Liaoning | 2012 | Pet | 328 | 33 | 10.0 | IHA^b | unknown | | | | | [24] |
| | | 2012 | Police | 291 | 90 | 30.9 | MAT ^e | 45 | 27 | 0 | 12 | 6 | [25] |
| IV | Shangdong | 2010-2011 | Stray/pet | 143 | 14 | 9.8 | IHA^b | unknown | | | | | [23] |
| V | Henan | 2010-2011 | Stray/pet | 106 | 13 | 12.3 | IHA^b | unknown | | | | | [23] |
| | | 2013 | Countryside | 31 | 1 | 3.2 | MAT ^e | 1 | 1 | 0 | 0 | 0 | Present study |
| | | unknown | Stray/countryside | 231 | 93 | 40.3 | ELISAª | unknown | | | | [26] | |
| VI | Jiangshu | 2010 | Household | 288 | 62 | 21.5 | MAT ^e | 21 | 15 | 0 | 11 | 9 | [27] |
| | | | City pet | 1178 | 15 | 1.3 | | | | | | | |
| VII | Shanghai | 2009 | Town pet | 364 | 22 | 6.0 | IHA ^b | unknown | | | | | [28] |
| | | | Countryside | 194 | 19 | 9.8 | | | | | | | |
| VIII | Xinjiang | 2010-2011 | Stray/pet | 259 | 29 | 11.2 | IHA^b | unknown | | | | | [23] |
| IX | Gansu | 2010 | Pet | 259 | 28 | 10.8 | MAT ^e | 14 | 9 | 4 | 1 | 0 | [29] |
| Χ | Sichuan | 2010 | Household | 314 | 11 | 3.5 | IHA ^b | 0 | 0 | 4 | 3 | 4 | [30] |
| XI | Yunnan | 2011-2012 | Pet | 611 | 132 | 21.6 | IHA^c | 0 | 0 | 29 | 25 | 78 | [31] |
| XII | Guangdong | unknown | Stray | 36 | 12 | 33.3 | ELISA ^d | | | | | | [0.0] |
| | | | Household | 114 | 20 | 17.5 | | unkno | wn | | | | [32] |
| VIII | Jilin | 2013 | Countryside | 43 | 7 | 16.3 | AAATP | 5 | 6 | 1 | 0 | 0 | Present study |
| XIII | | 2014 | Countryside | 329 | 27 | 8.2 | MAT ^e | 0 | 8 | 13 | 6 | 0 | |
| XIV | Anhui | 2013 | Countryside | 22 | 0 | 0 | MAT ^e | 0 | 0 | 0 | 0 | 0 | Present study |

IHA: indirect hemagglutination test, ELISA: enzyme-linked immunosorbent assay, LAT: latex agglutination test.

^a:ELISA Kit produced by Zhuhai S.E.Z Haitai Pharmaceutlcals Co., Ltd, China.

b:Kit produced by Lanzhou Veterinary Institute, Academy of Agriculture and Science, China.

^c:Kit produced by Nanjing Veterinary research Institute, Jiangshu Academy of Agriculture and Science, China.

d:Kit produced by Combined Company, Shenzhen, China.

e:in-house.

Bioassay of tissues for T. gondii

Myocardium (50 g, or 30 g for young dog) from the first batch samples with MAT of T. gondii seropositive (MAT, \geq 10) dogs (n = 14) was digested individually in pepsin and bioassayed in mice as described [7]. Briefly, tissues were washed and homogenized in saline (0.85% NaCl), mixed with acidic pepsin, and incubated in a shaker water bath for 60 min at 37°C. The homogenate was filtered through two layers of gauze, centrifuged, sediment neutralized with sodium bicarbonate, centrifuged again, mixed with antibiotics, and the homogenate inoculated subcutaneously into three Swiss Webster (SW) outbreed albino mice, and two gamma interferon gene knockout (KO) mice [7]. All inoculated mice were observed daily for illness. Dead mice, or killed when ill, were examined for T. gondii by making impression smears from the lung and examined for tachyzoites. Survivors were bled on day 41 post-inoculation (p.i.) and 1:100 dilution of serum from each mouse was tested for T. gondii antibodies with the MAT. Mice were killed 47 or 48 days post infection and brains of all mice were examined for tissue cysts as a squash preparation as described [7].

Results and discussion

Six (6.25%) of 96 dogs were seropositive of *N. caninum* with titers of 1:25 in 2, 1:50 in 3, and 1:100 in 1, and all seropositive dogs were more than 1 year old (Table 1).

Antibodies to *N. caninum* were found in 6 (13.95%) of 43 dogs from Jilin but not in 53 dogs from the Henan and Anhui, because most of the dogs from other two regions were less than one year old. Antibodies to *T. gondii* were found in 35 of 425 (8.24%) dogs with titers of 1:25 in 15, 1:50 in 14; and 1:100 in 6, none was positive at 1:200 (Table 1). The age range of the *N. caninum* and *T. gondii* negative dogs was 1.06 ± 0.55 (years) and 2.90 ± 1.38 (years). The age range of the *N. caninum* and *T. gondii* positive dogs was 2.25 ± 1.42 (years) and 3.06 ± 2.16 (years). T. gondii was not isolated in mice inoculated with tissues of any dogs.

Hearts samples and serum were kept cold and transported by air from China to USA within one week of killing. One week antibodies are fairly stable. *T. gondii* antibodies were stable on dried filter papers for up to 45 days at room temperature [8] and for 6 months at 25°C when stored with silica gel [9]. In an unpublished experiment antibodies were still detectable in blood 7 days after they were accidently left in a water bath at 37°C (JP Dubey, unpublished 1971).

In the present study, antibodies to *N. caninum* were detected in dog sera for the first time from China. How dogs become infected with *N. caninum* in nature is unknown [1]. Congenital transmission and ingestion of infected tissues or oocysts are the three probable modes of transmission. Of these, the ingestion of infected tissues is

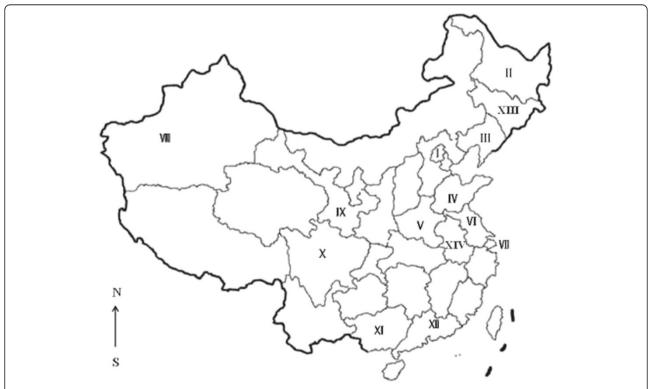


Figure 1 Seroepidemiology of *Toxoplasma gondii* in dogs in China. I: Beijing, II:Heilongjiang, III:Liaoning, IV:Shandong, V:Henan, VI: Jiangshu, VII:Shanghai, VIII:Xinjiang, IX:Gansu, X:Sichuan, XI:Yunnan, XII:Guangdong, XIII: Jilin, XIV: Anhui.

considered the most important because congenital transmission in dogs is relatively rare and fecal transmission has not been proven [3]. Little is known of the natural epidemiology of N. caninum in nature. In the USA, cattle and deer are commonly infected with N. caninum. Viable parasite has not been isolated from small mammals. Little is known of the epidemiology of neosporosis in China. In this study, antibodies to N. caninum were 6.25% (6/96) of first batch samples tested. We didn't test the second batch samples because of non availability of reagents for testing of N. caninum by NAT, or IFA, or cELISA in China. Compared with other countries [10-13], the prevalence of N. caninum antibody was low in East Asia [14,15]; also it is low in this study. Although a comparison of different serological tests (NAT, IFA, cELISA) for the detection of N. caninum antibodies in dogs has not been made, it is generally accepted that results by NAT and IFA are comparable because both tests assess antibodies directed against whole tachyzoites [1]. None of the serological tests, however, have been validated because it is very difficult to isolate *N. caninum* from asymptomatic hosts [1].

In this study, the prevalence of antibodies and titers to T. gondii were low compared to data from other parts of China (Table 2, Figure 1) and other country [16,17]. However, results are not strictly comparable because of different serological tests used and different cut-off used. In several of these previous reports, the serological tests used have not been critically evaluated. To facilitate further investigations we have summarized available reports on canine toxoplasmosis in Table 2. The MAT we used has been extensively used for the detection of T. gondii antibodies in many species, including humans and dogs [7]. Viable T. gondii was isolated from 51% (22/43) of dogs with MAT antibodies [18] and there is no evidence for any cross reactivity with other antigens in MAT [19]. The lack of isolation of *T. gondii* from the hearts of any of the dogs in the present study could be related to the loss of infectivity during transit from China and USA. However, viable T. gondii has been isolated in the USDA laboratory in Beltsville from tissues of animals that had been in transit for up to 10 days from several countries to USA. The survival of T. gondii tissue in meat at room temperature is largely unknown. However, viable T. gondii was isolated from a Hawaiian bird that was completely rotten and most carcasses had been eaten by maggots [20]. At 40°C, T. gondii tissue cysts survived up to 2 months [21]. Temperatures at which the dog tissues samples were stored varied from 4-20°C while in transit from China to USA. Therefore, we are uncertain of the effect of storage on the survival of tissue cysts.

Conclusions

Results of the present study indicated low prevalence of *N. caninum* and *T. gondii* infection in dogs in China,

compared with data from Europe and America. Identification of the risk factors that underlie these differences may help prevention of neosporosis and toxoplasmosis. Dog meat is consumed by local people in China. Therefore, dogs are at a risk of *T. gondii* infection and should be of public health concern.

Competing interests

None of the authors of this paper has a financial or personal relationship with other people or organisations that could inappropriately influence or bias the content of the paper.

Authors' contributions

YRY performed the laboratory tests, data analysis, and wrote the manuscript. QFZ participated in the collect samples, and the laboratory tests. YGK participated in the collect samples, obtained the general data of the dogs. YQY participated in the laboratory tests. OCHK participated in the laboratory tests and helped in the writing of the manuscript. HDL designed the study protocol, obtained the serum samples and general data of the dogs. JP D designed the study protocol, analyzed the results and helped in the writing of the manuscript. All authors read and approved the final version of the manuscript.

Acknowledgements

This research project was financed in part by China Scholarship Council.

Author details

¹Laboratory of Veterinary Pathology, College of Animal Science and Veterinary Medicine, Henan Agriculture University, Zhengzhou 450002, PR China. ²United States Department of Agriculture, Agricultural Research Service, Beltsville Agricultural Research Center, Animal Parasitic Diseases Laboratory, Beltsville, MD 20705-2350, USA.

Received: 7 September 2014 Accepted: 5 December 2014 Published online: 12 December 2014

References

- Dubey JP: Neosporosis in dogs. Commonwealth Agricultural Bureau Reviews 2013, 8:1–26.
- Yao L, Yang N, Liu Q, Wang M, Zhang W, Qian WF, Hu YF, Ding J: Detection of *Neospora caninum* in aborted bovine fetuses and dam blood samples by nested PCR and ELISA and seroprevalence in Beijing and Tianjin, China. *Parasitology* 2009, 136:1251–1256.
- Dubey JP, Schares G: Neosporosis in animals the last five years. Vet Parasitol 2011. 180:90–108.
- Lindsay DS, Dubey JP, Butler JM, Blagburn BL: Mechanical transmission of Toxoplasma gondii oocysts by dogs. Vet Parasitol 1997, 73:27–33.
- Dubey JP, Desmonts G: Serological responses of equids fed Toxoplasma gondii oocysts. Equine Vet J 1987, 19:337–339.
- Romand S, Thulliez P, Dubey JP: Direct agglutination test for serologic diagnosis of Neospora caninum infection. Parasitol Res 1998, 84:50–53.
- Dubey JP: Toxoplasmosis of animals and humans. In 2nd edition. Boca Raton, Florida, USA: CRC Press, Taylor & Francis Group; 2010:1–313.
- Bahia MT, Vitor RWA, Caldas R, Antunes CMF, Chiari CA: Use of filter paper in serodiagnosis of goat toxoplasmosis. Braz J Vet Res Anim Sci 1995, 32:83–88. Portuguese.
- Nogami S, Kamata H, Maruyama S, Furuya H, Inoue I: Preservation of feline anti-toxoplasma gondii antibody activity using blood absorbed on filter paper stored under different conditions. Res Ves Sci 1992, 52:387–388.
- Nazir MM, Maqbool A, Akhtar M, Ayaz M, Ahmad AN, Ashraf K, Ali A, Alam MA, Ali MA, Khalid AR, Lindsay DS: Neospora caninum prevalence in dogs raised under different living conditions. Vet Parasitol 2014, 204:364–368.
- de Sousa ME, Porto WJ, de Albuquerque PP, de Souza Neto OL, Pinheiro Júnior JW, Mota RA: Seroprevalence of antibodies to Neospora caninum in dogs in the state of Alagoas, Brazil. Rev Bras Parasitol Vet 2012, 21:287–290.
- Gavrea R, Mircean V, Pastiu A, Cozma V: Epidemiological survey of Neospora caninum infection in dogs from Romania. Vet Parasitol 2012, 188-382–385

- Kuruca L, Spasojevic-Kosic L, Simin S, Savovic M, Laus S, Lalosevic V: *Neospora caninum* antibodies in dairy cows and domesic dogs from Vojvodina, Serbia. *Parasite* 2013, 20:40.
- Kubota N, Sakata Y, Miyazaki N, Itamoto K, Bannai H, Nishikawa Y, Xuan X, Inokuma H: Serological survey of Neospora caninum infection among dogs in Japan through species-specific ELISA. J Vet Med Sci 2008, 70:869–872
- Nguyen TT, Choe SE, Byun JW, Koh HB, Lee HS, Kang SW: Seroprevalence of Toxoplasma gondii and Neospora caninum in dogs from Korea. Acta Parasitol 2012. 57:7–12.
- Alvarado-Esquivel C, Romero-Salas D, Cruz-Romero A, García-Vázquez Z, Peniche-Cardeña A, Ibarra-Priego N, Ahuja-Aguirre C, Pérez-de-León AA, Dubey JP: High prevalence of *Toxoplasma gondii* antibodies in dogs in Veracruz, Mexico. BMC Vet Res 2014, 10:191.
- Langoni H, Fornazari F, da Silva RC, Monti ET, Villa FB: Prevalence of antibodies against *Toxoplasma gondii* and *Neospora caninum* in dogs. *Braz J Microbiol* 2014, 44:1327–1330.
- El Behairy AM, Choudhary S, Ferreira LR, Kwok OC, Hilali M, Su C, Dubey JP: Genetic characterization of viable *Toxoplasma gondii* isolates from stray dogs from Giza, Egypt. *Vet Parasitol* 2013, 193:25–29.
- Dubey JP: Validation of the specificity of the modified agglutination test for toxoplasmosis in pigs. Vet Parasitol 1997, 71:307–310.
- Work TM, Massey JG, Rideout BA, Gardiner CH, Ledig DB, Kwok OC, Dubey JP: Fatal toxoplasmosis in free-ranging endangered 'Alala from Hawaii. J Wildl Dis 2000. 36:205–212.
- 21. Jacobs L, Remington JS, Melton ML: The resistance of the encysted form of *Toxoplasma gondii*. *J Parasitol* 1960, **46**:11–21.
- Yu J, Ding J, Xia Z, Lin D, Li Y, Jia J, Liu Q: Seroepidemiology of Toxoplasma gondii in pet dogs and cats in Beijing, China. Acta Parasitol 2008, 53:317–319.
- Liu Y, He G, Cheng Z, Qi Y, Liu J, Zhang H, Liu G, Shi D, Yang D, Wang S, Wang Z: Seroprevalence of *Toxoplasma gondii* in dogs in Shandong, Henan, and Heilongjiang Provinces, and in the Xinjiang Uygur Autonomous Region, People's Republic of China. *J Parasitol* 2012, 98:211–212.
- 24. Yang N, Mu M, Li H, Hu J, Gao W, Yang S, He J: Seroprevalence of *Toxoplasma gondii* infection in pet dogs in Shenyang, Northeastern China. *J Parasitol* 2013, **99:**176–177.
- Liu CW, Yang N, He JB, Mu MY, Yang M, Sun N, Li HK: Seroprevalence of Toxoplasma gondii infection in police dogs in Shenyang, northeastern China. Korean J Parasitol 2013, 51:579–581.
- Yan C, Fu LL, Yue CL, Tang RX, Liu YS, Lv L, Shi N, Zeng P, Zhang P, Wang DH, Zhou DH, Zhu XQ, Zheng KY: Stray dogs as indicators of *Toxoplasma gondii* distributed in the environment: the first report across an urban-rural gradient in China. *Parasit Vectors* 2012, 5:6.
- 27. Li Y, Liu Q, Li S, Wei F, Jin H, Yang M: Seroprevalence of *Toxoplasma gondii* infection in dogs in Jiangsu Province, eastern China. *J Parasitol* 2012, **98**:878–879.
- Wang Q, Jiang W, Chen YJ, Jing ZY: Prevalence of Toxoplasma gondii antibodies and DNA in dogs in Shanghai, China. J Parasitol 2011, 97:367–369.
- Wu SM, Huang SY, Fu BQ, Liu GY, Chen JX, Chen MX, Yuan ZG, Zhou DH, Weng YB, Zhu XQ, Ye DH: Seroprevalence of *Toxoplasma gondii* infection in pet dogs in Lanzhou, Northwest China. *Parasit Vectors* 2011, 4:64.
- Li B, Zhong N, Peng W, Shang L, Jin H, Liu Q: Seroprevalence of Toxoplasma gondii infection in dogs in Sichuan Province, southwestern China. J Parasitol 2012, 98:209–210.
- Duan G, Tian YM, Li BF, Yang JF, Liu ZL, Yuan FZ, Zhu XQ, Zou FC: Seroprevalence of *Toxoplasma gondii* infection in pet dogs in Kunming, Southwest China. *Parasit Vectors* 2012, 5:118.
- 32. Zhang H, Zhou DH, Chen YZ, Lin RQ, Yuan ZG, Song HQ, Li SJ, Zhu XQ: Antibodies to *Toxoplasma gondii* in stray and household dogs in Guangzhou, China. *J Parasitol* 2010, **96**:671–672.

doi:10.1186/s12917-014-0295-3

Cite this article as: Yang et al.: Low prevalence of Neospora caninum and Toxoplasma gondii antibodies in dogs in Jilin, Henan and Anhui Provinces of the People's Republic of China. BMC Veterinary Research 2014 10:295.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at www.biomedcentral.com/submit

