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# Identification of an unusual *Brucella* strain (BO2) from a lung biopsy in a 52 year-old patient with chronic destructive pneumonia

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## Abstract

**Background:** Brucellosis is primarily a zoonotic disease caused by *Brucella* species. There are currently ten *Brucella* spp. including the recently identified novel *B. inopinata* sp. isolated from a wound associated with a breast implant infection. In this study we report on the identification of an unusual *Brucella*-like strain (BO2) isolated from a lung biopsy in a 52-year-old patient in Australia with a clinical history of chronic destructive pneumonia.

**Results:** Standard biochemical profiles confirmed that the unusual strain was a member of the *Brucella* genus and the full-length 16S rRNA gene sequence was 100% identical to the recently identified *B. inopinata* sp. nov. (type strain BO1<sup>T</sup>). Additional sequence analysis of the *recA*, *omp2a* and *2b* genes; and multiple locus sequence analysis (MLSA) demonstrated that strain BO2 exhibited significant similarity to the *B. inopinata* sp. compared to any of the other *Brucella* or *Ochrobactrum* species. Genotyping based on multiple-locus variable-number tandem repeat analysis (MLVA) established that the BO2 and BO1<sup>T</sup> strains form a distinct phylogenetic cluster separate from the other *Brucella* spp.

**Conclusion:** Based on these molecular and microbiological characterizations, we propose that the BO2 strain is a novel lineage of the newly described *B. inopinata* species.

## Background

Brucellosis is primarily a zoonotic disease, caused by members of the genus *Brucella*, which currently constitutes several species based on pathogenicity, host preferences and phenotypic characteristics: *B. abortus* (cattle), *B. canis* (dogs), *B. melitensis* (goats), *B. suis* (pigs), *B. ovis* (rams), *B. neotomae* (desert rats), *B. ceti* and *B. pinnipedialis* (marine mammals), and *B. microti* (common vole) [1-6]. Recently, a novel species, *Brucella inopinata*, associated with a human infection has been recognized as the newest member of the genus *Brucella* [7,8]. In early 1985, whole genome hybridization analysis studies revealed a high degree of genetic homology among the *Brucella* species, which led to the proposal that the genus *Brucella* was a mono-specific species with *B. melitensis* being the primary species and all others as

sub-species and biovars [9-11]. However, due to the limited acceptability of the one-species concept, the traditional classification of *Brucella* spp. based on phenotypic characteristics has been re-instated by the *Brucella* Taxonomy Subcommittee in 2006 [3].

*Brucella* are facultative intracellular pathogens that infect many organs and soft tissues, including mammary glands. Infection frequently results in abortion, low milk production and fetal death in animals [2,12-16]. Brucellosis in humans is mostly caused by *B. abortus*, *B. melitensis*, *B. suis*, and sometimes *B. canis* [14,17-19], and is commonly associated with the consumption of unpasteurized dairy products, meat from infected animals and exposure to infected animal tissues or laboratory transmission [1,2,20]. Human brucellosis is a chronic debilitating infection with a very broad clinical picture potentially affecting any major organ, including the lung, causing varying respiratory symptoms [20]. Respiratory infections in humans caused by *Brucella* spp. is a rare manifestation with reports describing

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multifocal abscesses or nodules, hilar adenopathy and hemorrhagic pleural effusion with resolution by antimicrobial therapy and lung decortications [21-26]. Most pulmonary brucellosis cases were found in farmers handling infected meat or travelers who consumed raw infected animal meat or unpasteurized milk products while visiting countries endemic for brucellosis [26,27].

We report the isolation and identification of an unusual gram-negative, non-motile *Brucella*-like coccoid bacillus (BO2) isolated from a lung biopsy in a 52-year-old male in Australia with a history of chronic destructive pneumonia. The patient traveled worldwide but denied any common risk factors associated with brucellosis. Both biochemical and molecular characteristics of the BO2 strain have demonstrated unique similarity with a recently described *B. inopinata* strain (BO1<sup>T</sup>) associated with a breast implant wound of a 71-year-old patient from Portland, Oregon with clinical signs of brucellosis [7,8].

## Results

### Phenotypic characterization

BO2 cells grown on SBA or RBA at 35-37°C with or without 5% CO<sub>2</sub> for 24 to 48 h were circular, convex, entire, smooth and opaque. The organisms were gram-negative, generally stained uniformly; and appeared coccoid to short coryneform rods. Colonies of the BO2 strain ranged in size from punctuate to 1.5 mm in diameter and they were non-motile, mucoid colonies on MacConkey agar; positive for oxidase and catalase, exhibited nitrate reduction with production of gas and rapid urease production (< 5 min). Hydrogen sulfide production by the BO2 strain was observed by the development of a dark gray color on lead acetate paper suspended above the heart infusion agar slant.

Subculture of individual colony types produced similar profiles and no hemolytic reaction was observed on SBA plates after overnight incubation at 37°C. The BO2 cells grew in the presence of thionine (1:25,000, 1:50,000 and 1:100,000 dilutions) and basic fuchsin (1:50,000 and 1:100,000 dilutions) dyes within 24 to 48 h. Both the acriflavin and gel formation tests were negative. However, lysis by Tbilisi phage specific for detection of *Brucella* spp. in two routine test dilutions (1× and 4× RTD) appeared incomplete [7,8,28] and agglutination of the BO2 cells with either monospecific anti-M or anti-A antisera were very weak.

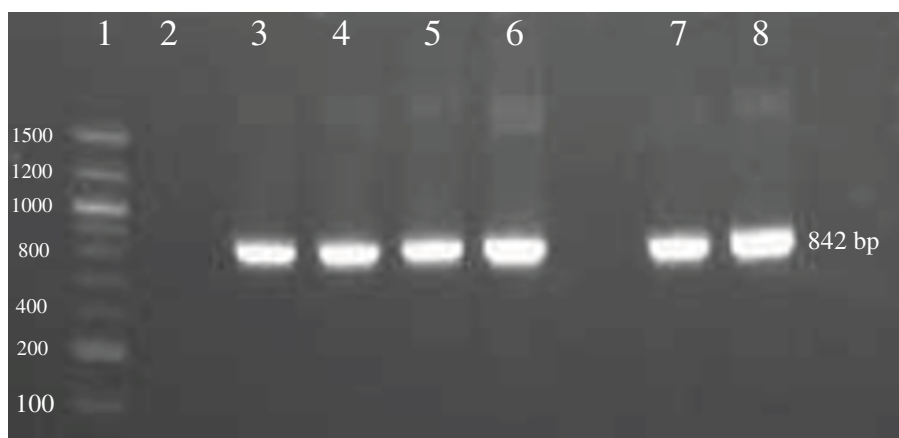
### Antimicrobial susceptibility test

The antimicrobial susceptibility profile of the BO2 strain was compared with a set of 93 other *Brucella* spp. strains (74 *B. melitensis*, 14 *B. suis* and 5 *B. abortus*) along with BO1<sup>T</sup> based on CLSI interpretive requirements for *Brucella* spp. [8,29,30]. Both strains had very similar MIC patterns to all *Brucella* reference strains tested previously [8,30] (Table 1). BO1<sup>T</sup> and BO2 strains grew well in cation-adjusted Mueller-Hinton broth (CAMHB) after just 20 hours of incubation, unlike other *Brucella* spp. (e.g., *B. abortus*, *B. melitensis*, and *B. suis*) which do not routinely grow very well in CAMHB and require 48 hours of incubation in *Brucella* broth for MIC testing [30]. Our standard phenotypic characterization, including the antimicrobial susceptibility profiles, suggested that the BO2 strain more closely resembled the BO1<sup>T</sup> strain of the *B. inopinata* sp. than the other classical *Brucella* spp.

### Molecular characterization

#### Detection of IS711 element by PCR

The *Brucella* specific insertion sequence (IS711) PCR was performed amplifying an 842-bp repetitive element using BO2 genomic DNA. The IS711 profile observed in



**Figure 1** IS711 profiles of PCR amplified products analyzed by gel electrophoresis on a 2% E-Gel displaying the following: molecular weight marker (lane 1), no template control (lane 2), *B. abortus* ATCC 23448 (lane 3), *B. melitensis* 16 M (lane 4), *B. suis* ATCC 23444 (lane 5), *B. ovis* ATCC 25840 (lane 6), BO1<sup>T</sup> (lane 7), and BO2 (lane 8).

strain BO2 was approximately the same size as that of the BO1<sup>T</sup> strain and the classical *Brucella* spp. including *B. ovis* (ATCC 25840) (Figure 1). The BO2 strain also generated several large amplicons (>1000 bp) similar to BO1<sup>T</sup> and other *Brucella* strains with low intensity as reported earlier [8].

#### Real-Time PCR for BO1<sup>T</sup>/BO2

A TaqMan PCR assay targeting conserved regions of the BO1<sup>T</sup> and *Brucella* spp. 16S rRNA gene sequence was designed for rapid differentiation of potential *B. inopinata*-like strains from all other classical *Brucella* and *Ochrobactrum* spp. This real-time PCR assay, using two hybridization probes: BI-P specific for *B. inopinata* spp. and BRU-P specific for *Brucella*/*Ochrobactrum* spp., gave average crossing threshold (Ct) values in the range of 15 to 20 (strong positive). The BI-P probe demonstrated perfect agreement for both BO1<sup>T</sup> and BO2 strains as did the BRU-P probe for all other *Brucella* or *Ochrobactrum* spp. respectively. Both probes showed no cross reactivity against the other non-*Brucella* strains tested to date [31] demonstrating very high specificity of the target sequences in the PCR assay. Both the BO1<sup>T</sup>/BO2 and the *Brucella*/*Ochrobactrum* specific probes were capable of optimal detection of template down to 10 fg/μl concentration of genomic DNA template (data not shown).

#### 16S rRNA gene sequence analysis

Rapid identification of the BO2 strain as *B. inopinata*-like by the BO1 PCR assay led to sequence analysis of the full-length 16S rRNA gene (1,412 bp) of the BO2 strain. Full sequence alignment with the 16S rRNA gene sequences of BO1<sup>T</sup>, reference *Ochrobactrum* spp. strains, and the *Brucella* spp. consensus sequence confirmed that the BO2 strain shared 100% 16S rRNA gene sequence identity to that of BO1<sup>T</sup> and 99.6% identity with other *Brucella* spp. (Table 2).

#### Omp2a/2b genes sequence analysis

We also analyzed two highly homologous outer membrane porin genes (*omp2a* and *omp2b*) of the BO2 and BO1<sup>T</sup> strains and compared their full-length sequences

with that of other *Brucella* species available in GenBank. The phylogenetic relationships derived by neighbor-joining clustering analysis of the BO2 *omp2a* (1093 bp) and *omp2b* (~1212 bp) genes with the NCBI sequences of other *Brucella* strains and the *Ochrobactrum anthropi* LMG 3331 reference strain demonstrated considerable intra- and inter-species variability (Figure 2). The BO2 *omp2a* and *omp2b* genes are 84.6% homologous to each other. Neighbor-joining clustering analysis of both *omp2a* and *omp2b* nucleotide sequences shows that BO2 clusters closest to BO1<sup>T</sup> and an atypical *B. suis* 83-210 strain [32]. The *omp2a* gene of BO2 is only 1.0% divergent from that of BO1<sup>T</sup>. The *omp2b* gene is characteristically more diverse within the *Brucella* spp. and is also evident with the BO2 *omp2b* gene which was 95.3% and 94.1% identical to the BO1<sup>T</sup> and *B. suis* 83-210 strains, respectively (Figure 2, Table 2). Clustering analysis demonstrates that BO1<sup>T</sup>, BO2 and the *B. suis* 83-210 strains form consistent sub-groups based on their *omp2a* and *omp2b* gene homology [32].

#### RecA gene sequence analysis

The *recA* gene (948 bp) of strain BO2 was compared to those of BO1<sup>T</sup>, the classical *Brucella* spp. (n = 8) and several representative *Ochrobactrum* spp. [31,33]. Within the genus *Brucella*, the *recA* gene is highly conserved with 100% nucleotide sequence identity among the different species. Interestingly, the BO2 *recA* nucleotide sequence reveals 99.2% identity to the *Brucella* consensus *recA* sequence due to 8 nucleotide substitutions. However, the BO2 *recA* gene has a lower identity (98.2%) when compared to the BO1<sup>T</sup> *recA* sequence differing by 17 nucleotides. Phylogenetic analysis of BO1<sup>T</sup> and BO2 strains with other *Brucella* and *Ochrobactrum* spp. shows that the *Brucella* spp. clade including BO2 and BO1<sup>T</sup>, are distantly similar to the *Ochrobactrum* spp. with approximately 85% sequence identity (Figure 3).

#### Multiple Locus Sequence Analysis

Multiple locus sequence analysis (MLSA) of nine *Brucella* spp. house-keeping genes has been used to

**Table 1 MIC results for 5 antimicrobial agents tested against BO1<sup>T</sup>, BO2 strains and 93 *Brucella* strains**

Antimicrobial agent	BO1 <sup>T</sup> MIC (μg/ml)			BO2 MIC (μg/ml)			Brucella spp. <sup>a</sup> in Brucella broth 48 h	
	CAMHB <sup>b</sup>	Brucella Broth	Brucella Broth	CAMHB	Brucella Broth	Brucella Broth	MIC Range (μg/ml)	MIC <sub>90</sub> (μg/ml)
Antimicrobial agent	20 h	20 h	48 h	20 h	20 h	48 h	(μg/ml)	(μg/ml)
Doxycycline	0.25	0.25	0.5	0.25	0.25	0.5	0.06 - 1	0.25
Gentamicin	1	2	2	1	2	2	0.5 - 2	1
Streptomycin	4	4	4	2	4	4	1 - 8	4
Tetracycline	0.25	0.5	1	0.12	0.25	0.25	0.12 - 1	0.5
Trimethoprim-sulfamethoxazole	0.5/9.5	0.25/4.75	0.5/9.5/0.25	0.5/9.5	0.25/4.75	0.5/9.5	0.12/2.38 - 0.5/9.5	0.5/9.5

<sup>a</sup> Ninety-three *Brucella* isolates (74 *B. meli*tensis, 14 *B. suis*, and 5 *B. abortus*) were tested [30].

<sup>b</sup> CAMHB = cation-adjusted Mueller-Hinton broth.

**Table 2 Comparative percent identity based on pair-wise analysis of five genes of BO2 with BO1<sup>T</sup> and classical *Brucella* spp. using MEGA4.**

BO2 genes	<i>B. inopinata</i> BO1 <sup>T</sup> (%)	<i>Brucella</i> spp. (%)
16S rRNA	100.0	99.6
<i>RecA</i>	98.2	99.2
MLSA	98.7	98.3-98.6
<i>Omp2a</i>	99.0	85.4-98.4
<i>Omp2b</i>	95.3	83.8-95.3

differentiate *Brucella* spp. into distinct sequence types (ST). BO1<sup>T</sup> was determined to be 1.67% divergent from ST1 and to possess novel alleles at all nine loci [8]. BO2 has shown similar divergence (1.5%) from ST1 by MLSA also with novel alleles in all nine loci. Neighbor-joining phylogenetic analysis clearly shows how divergent the BO1<sup>T</sup> and BO2 species are from the classical *Brucella* sequence types (Figure 4). Throughout the 4,396-bp sequence examined, the BO1<sup>T</sup> and BO2 genomes have 32 common SNPs while there are 30 BO1<sup>T</sup> and 26 BO2 specific nucleotide changes that further characterize the divergence of these two strains at these highly conserved loci in the *Brucella* genus.

#### Multiple-Locus Variable-Number Tandem Repeat Analyses

Both BO2 and BO1<sup>T</sup> strains were also investigated by multiple-locus variable-number tandem repeat (VNTR) analysis (MLVA) using fifteen VNTR loci by capillary electrophoresis. Results were compared with a panel of well-characterized *Brucella* strains (n = 209) representing known species from our collection [31]. Our MLVA-15 typing analysis of both BO2 and BO1<sup>T</sup> strains demonstrated unique VNTR profiles in which both strains have six *Brucella*-loci with the same alleles (VNTR 2, -3, -14, -20, -21 and -25); and seven loci with variable VNTR amplicons (VNTR1, -7, -27, -29, -30, -31 and -33). All VNTRs successfully amplified in both BO1 and BO2 with the exception of VNTR16 and -28 in BO1<sup>T</sup>. MLVA-15 analysis revealed that both BO2 and BO1<sup>T</sup> had distinct VNTR profiles in comparison to each other and other *Brucella* strains (Figure 5).

## Discussion

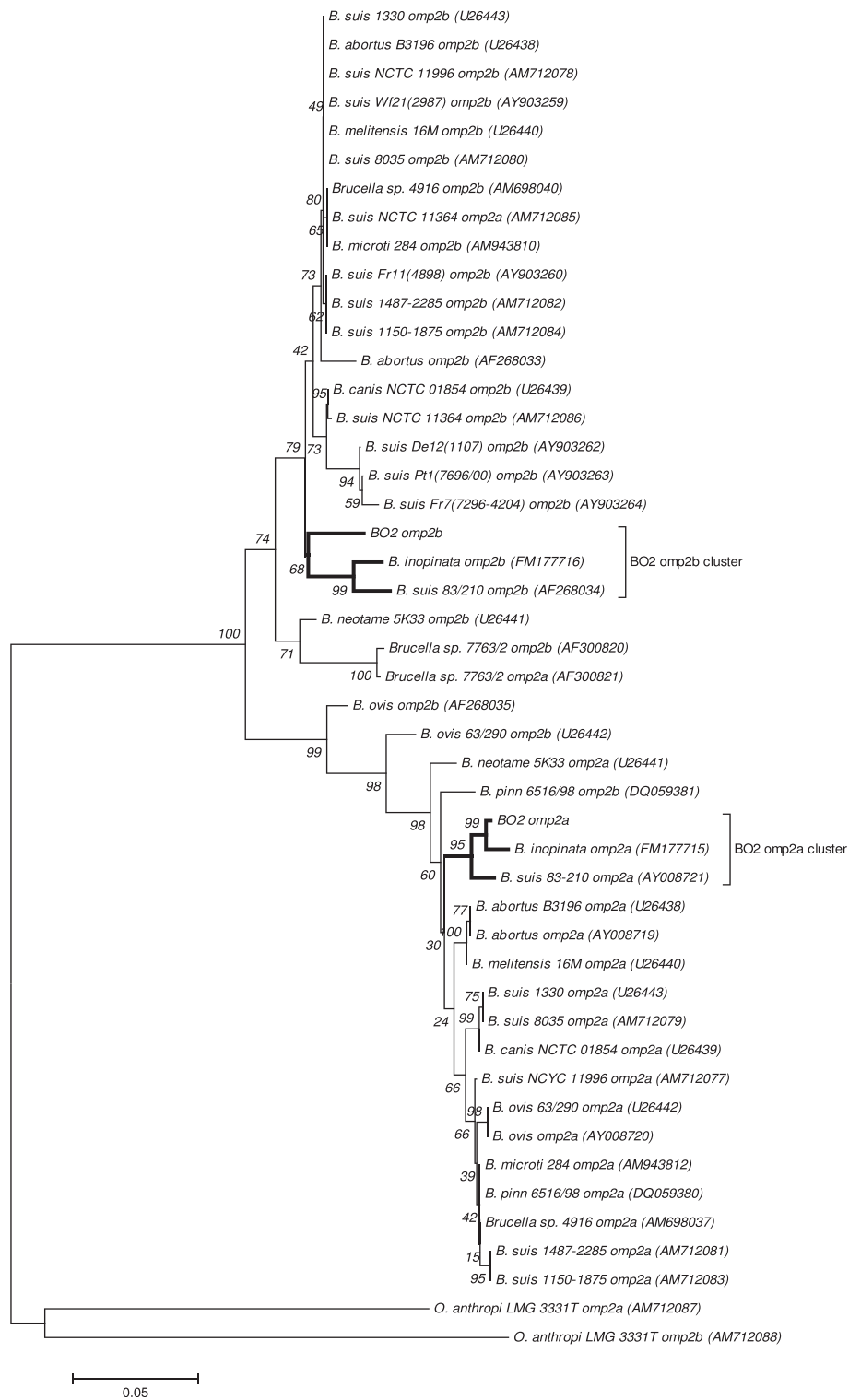
In this paper we present the identification of an atypical *Brucella*-like strain (BO2) isolated from the lung biopsy of a 52-year-old patient. As a young adult he lived in Oregon on two occasions (1981 and 1985-1987), and experienced an unexplained 'liver failure' and then severe pneumonia (with pleurisy) from which he recovered with multiple courses of antimicrobial therapy as reported by the patient to his physicians in Australia. This patient was originally misdiagnosed because of the misidentification of the BO2 strain as *O. anthropi* on an API 20NE system. It is a common practice for clinical

labs to attempt rapid identification of gram-negative *cocobacillus* organisms like *Brucella* spp. from blood culture using automated systems. However, the *Brucella* spp. are often misidentified due to their similar phenotypic characteristics to closely related organisms such as *Ochrobactrum* spp. [34,35]. Though the patient was initially treated for both *Ochrobactrum* and *Brucella* infections due to the difficulties in diagnosis, he recovered with an extended course of combination oral antimicrobial therapy.

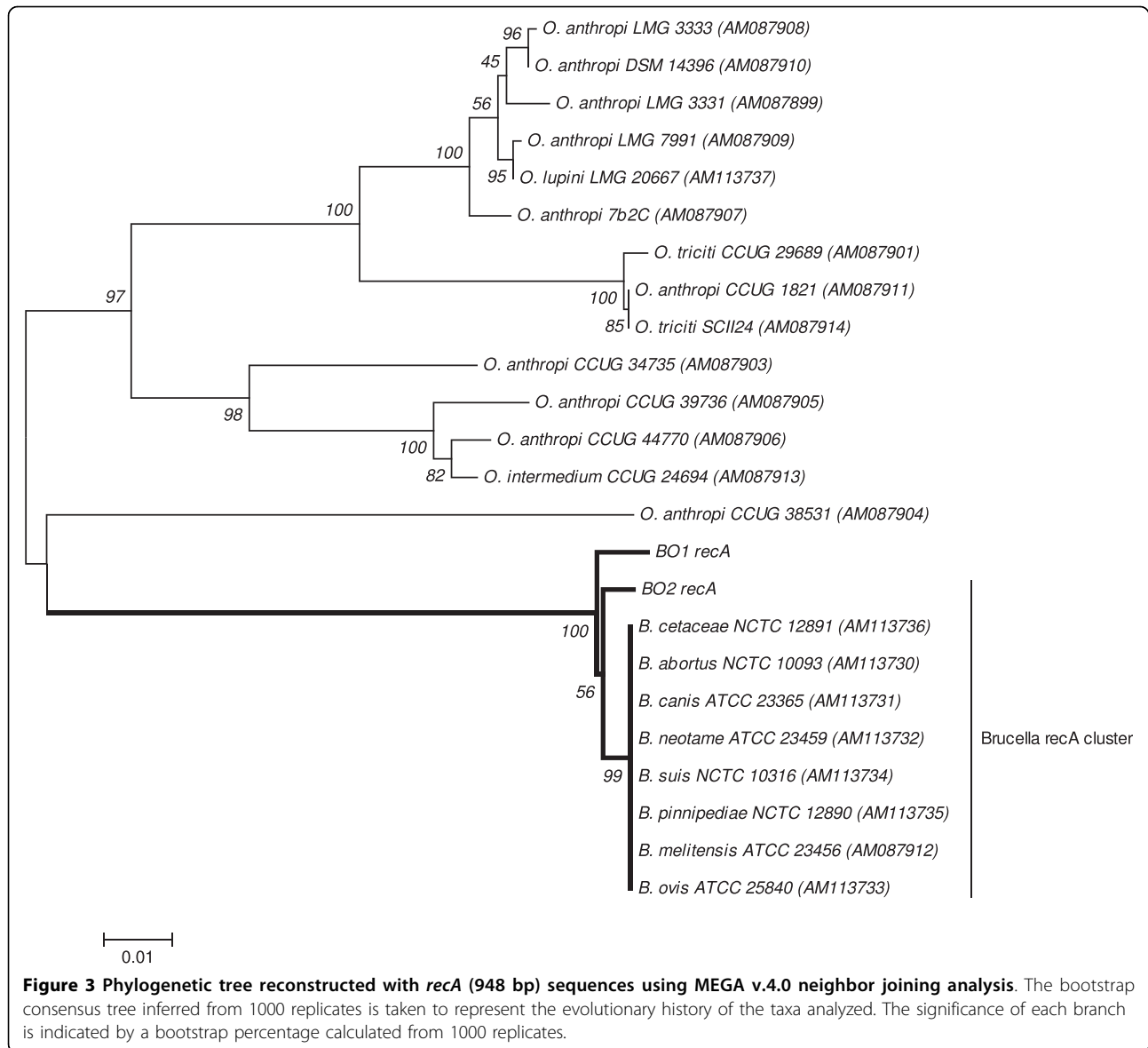
This BO2 strain is phenotypically and molecularly similar to the recently identified *B. inopinata* type strain (BO1<sup>T</sup>) recovered from a patient from Oregon, which was also originally misidentified as *O. anthropi* by the API 20E and API 20NE [7,8]. Both these strains share common colony morphology and biochemical characteristics including rapid urease and positive H<sub>2</sub>S production, inability or very weak agglutination with *Brucella* specific antisera for the lipopolysaccharide-O-antigens or acriflavin. Neither the BO1<sup>T</sup> or BO2 strains supports gel formation or exhibits growth inhibition to the dye media as shown by common members of the genus *Brucella*. BO2 also exhibited incomplete lysis by Tbilisi phage and had very similar antimicrobial susceptibility profiles to BO1<sup>T</sup> in comparison to other *Brucella* reference strains.

Insertion sequence (IS) fingerprinting in the *Brucella* species has shown that the genomic localization and copy number of the IS711 insertion element (also called IS6501) is species-specific and could have an association with specific pathogenicity for a preferred host [36-38]. The presence of multiple copies of BO1<sup>T</sup>-like IS711 insertion sequences suggest not only that BO2 is a member of the *Brucella* genus (Figure 1) but that the BO2-IS711 amplification pattern specifically resembles that of the newly described *B. inopinata* species [8]. Positive identification of the BO2 strain as a member of *B. inopinata* by our real-time BO1 PCR assay was significant. Both BO1<sup>T</sup> and BO2 strains were the cause of distinct and unusual forms of human brucellosis. Atypical clinical isolates of this nature can often be misdiagnosed by automated systems as was the case with BO1<sup>T</sup> and the BO2 strain described here [8,35]. The availability of the real-time TaqMan assay served as a reliable first-line tool for determining *B. inopinata*-like species.

These initial findings led to further characterization and sequence-based typing which provided additional supporting evidence that this new BO2 strain most resembles the *B. inopinata* sp. within the *Brucella* genus. Using broad-range eubacterial primers, Gee et al. effectively demonstrated the advantage of 16S rRNA gene sequencing to identify *Brucella* isolates reporting 100% identity in all the strains examined [31]. Interestingly, the full-length 16S rRNA gene sequence of BO2



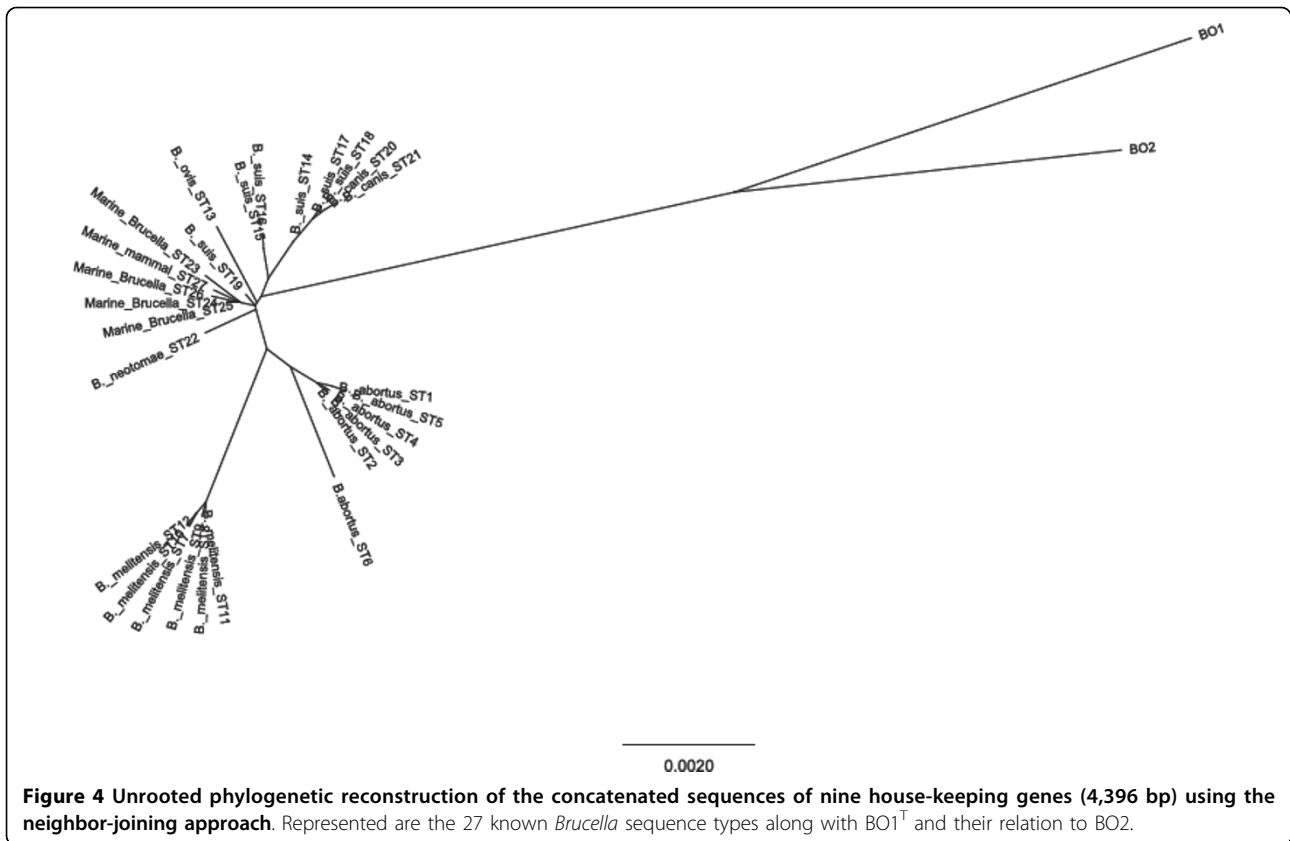
**Figure 2** Phylogenetic tree reconstructed with *omp2a* (1093 bp) and *omp2b* (~1211 bp) sequences using MEGA v.4.0 neighbor joining analysis. The bootstrap consensus tree inferred from 1000 replicates is taken to represent the evolutionary history of the taxa analyzed. The significance of each branch is indicated by a bootstrap percentage calculated from 1000 replicates.



was 100% identical to that of BO1<sup>T</sup> and 99.6% identical to the *Brucella* spp. consensus 16S rRNA gene sequence. The high sequence identity of the BO2 16S rRNA sequence to the recently described *B. inopinata* sp. is remarkable and represents the first recognized *Brucella* species to have a divergent 16S rRNA sequence [8].

The *recA* gene has been investigated as an alternative phylogenetic marker for several bacterial genera due to its highly conserved nature and ubiquity in prokaryotes [33,39,40]. Unlike the high sequence homology of the *recA* gene within the *Brucella* genus [41], we identified unique variability in the *recA* gene sequences of BO2 and BO1<sup>T</sup>. Sequence analysis revealed that the *recA* nucleotide sequence of the BO2 strain shared greater

similarity with the *Brucella* spp. *recA* consensus sequence than to BO1<sup>T</sup>. Both BO2 and BO1<sup>T</sup> *recA* sequences are distanced by 8 and 11 unique SNPs, respectively, from the *Brucella* spp. *recA* consensus sequence, and share only one common transversion at the 517 nucleotide position. Translation of the *recA* gene sequences of BO1<sup>T</sup>, BO2 and the *Brucella* spp. consensus sequence shows that all base pair changes were synonymous substitutions having no effect on protein structure or function. The *Brucella* outer membrane proteins have been studied extensively for their function in virulence, pathogenicity, bacteriophage reception, antigenic factors and antibacterial evasion [42-45]. The genetic variability among the *omp* genes within the *Brucella* spp. has proven effective at



characterizing *Brucella* spp. and strain types and is often used for higher resolution molecular typing [4,32,43,45]. The *omp2a/2b* genetic analysis we report here is very interesting in that BO2 consistently associates with not only BO1<sup>T</sup> but the atypical *B. suis* 83-210 strain that was isolated from a rodent in Australia [32]; and thus further investigation may be warranted into rodents as a possible natural reservoir for these novel *Brucella* species.

Investigation of the nine housekeeping genes by multi locus sequencing analysis demonstrates that BO2 is genetically distinct from BO1<sup>T</sup> yet exhibits remarkably similar divergence (1.5%) from the classical *Brucella* sequence types as shown in Figure 4. The relative similarity of the nucleotide sequences of BO1<sup>T</sup> and BO2 by MLSA demonstrates uniquely distant sequence types within the currently characterized *Brucella* spp. and should be considered as a new group of STs within the *Brucella* genus. They also exhibit distinct allelic profiles by MLVA although all alleles in both the BO1<sup>T</sup> and BO2 allelic profiles have been observed in other *Brucella* spp. Furthermore, the phylogenetic analysis shown in Figure 5 demonstrates that these strains form a single separate cluster from the classical *Brucella* spp. [8].

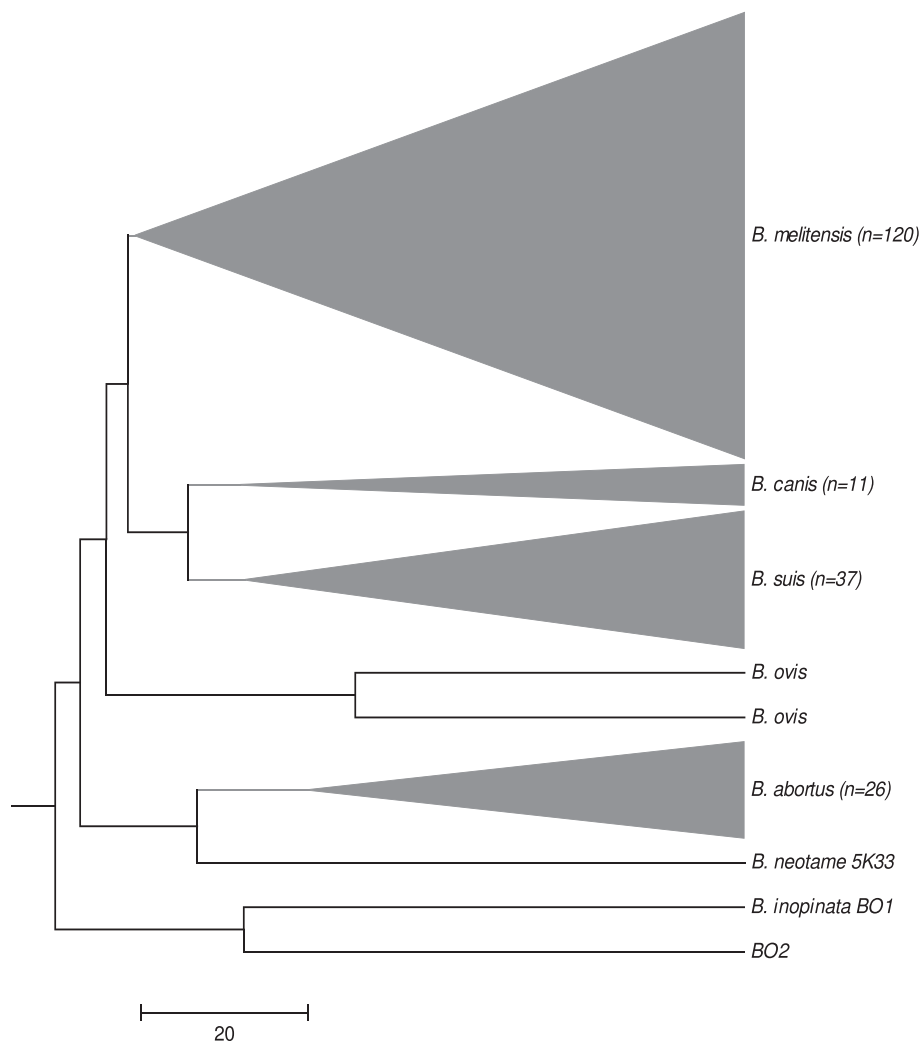
The molecular and microbiological characteristics presented here provide supporting evidence that strain BO2

is most closely associated with the BO1<sup>T</sup> strain and should be considered as a novel lineage of *B. inopinata* sp. Attempting to understand the evolutionary origin of these two strains is somewhat confounded by the interesting and disparate medical histories of the case patients (who both happened to have lived in Portland, Oregon) from whom these strains were isolated and suggests that there are new and emerging *Brucella* strains capable of causing unusual presentation of human brucellosis.

## Conclusion

Phenotypic and genomic analysis of the unusual *Brucella* strain (BO2) from a lung biopsy have established it as a lineage of the recently identified novel *B. inopinata* sp. type strain BO1<sup>T</sup>, which was isolated from a wound associated with a breast implant. This is the first report of a human brucellosis case associated with chronic destructive pneumonia caused by an atypical *Brucella* strain. An interesting finding from our molecular analysis reveals that both strains BO1<sup>T</sup> and BO2 appeared to be closely related to a less-characterized *B. suis* strain 83-210 (isolated from a rodent in Australia) by their *omp2a/2b* genes, which may suggest a common ancestor and may also provide insight into the ecological niche, and host reservoir for these novel *Brucella* strains causing unusual human infections.





**Figure 5** Condensed unweighted pair group method analysis (UPGMA) dendrogram of multiple-locus variable number tandem repeat analysis (MLVA) genotypes of BO1<sup>T</sup>, BO2 strains along with 209 characterized *Brucella* strains.

## Methods

### Patient

The patient was born in Malta in 1956 and immigrated to Australia at age two, where he would continually return and eventually settle throughout extensive worldwide travel including the Western region of the United States. Between 2003 and 2007, the patient was hospitalized multiple times in different hospitals in Australia for abnormal liver function, community acquired pneumonia, anterior chest wall abscess and sinus infection. In September 2007 a percutaneous lung biopsy was performed and a gram-negative organism was isolated from a broth culture of the fine needle aspirate of the patient's lung and identified as *Ochrobactrum anthropi* on an API20NE system. The testing laboratory was aware of the possibility of *Brucella* sp. being misidentified as *Ochrobactrum anthropi* [35] and the isolate was

referred for further testing. The patient was treated with combination therapy of doxycycline and rifampicin for twelve months and ciprofloxacin for three months (the latter was ceased after molecular testing confirmed *Brucella* species). The culture was initially tested according to standard microbiological and molecular procedures and then forwarded to the Centers for Disease Control and Prevention (CDC), Atlanta, GA, for further characterization. This gram-negative organism was designated as BO2 and stored at -70°C in defibrinated rabbit blood until further evaluation.

### Phenotypic analysis

The BO2 strain was routinely maintained on Trypticase soy agar with 5% defibrinated sheep blood agar (SBA) or rabbit blood agar (RBA) (BBL Microbiology Systems, Cockeysville, MD). Phenotypic identification of the BO2 strain was performed according to the laboratory



techniques in brucellosis described by Alton *et al.* in the World Health Organization monogram [7,8,28].

#### **Antimicrobial susceptibility analysis**

The antimicrobial susceptibility testing of the BO2 strain was performed by the broth microdilution method in CAMHB and *Brucella* broth in accordance with the Clinical and Laboratory Standards Institute (CLSI) protocol as described previously [8,29]

#### **Molecular analysis**

##### **Detection of IS711**

To detect the *Brucella*-specific insertion sequence IS711 element (842 bp) [37], cell lysate DNA templates from strains BO2, BO1<sup>T</sup>, *B. abortus* (ATCC 23448), *B. suis* 1330 (ATCC 23444), *B. ovis* (ATCC 25840) and *B. melitensis* 16 M (ATCC 23456) were amplified and the amplicons were analyzed by 2% E-Gel agarose gel electrophoresis as mentioned previously [8].

##### **Real-Time PCR assay**

A real-time TaqMan PCR assay was developed targeting a four base nucleotide substitution within the 16S rRNA gene sequence of BO1<sup>T</sup> (positions 145 to 148; GenBank accession no. EU053207). We designed two PCR primers 16SF (5'-CGGGCCGATCATTTGC-3') and 16SR (5'-AACTCAGGGAACTTGTGCTAATACC-3') to amplify a 72-bp region of the 16S rRNA *Brucella* consensus sequence and two hybridization probes, BI-P (5'-AAATCTTCCCCTTTCGGGCAC-3') and BRU-P (5'-AAATCTTCCCCGAAGGGCAC-3'), targeting a 4-bp polymorphic region within the 72-bp amplicon. Both probes were synthesized with a 6-carboxyfluorescein reporter molecule attached at the 5' end and Black Hole Quencher 1 on the 3' end. Each final PCR reaction mix contained 2 µl of DNA template and 18 µl of PCR master mixture containing 1 × LightCycler Faststart DNA Master HybProbe mix (Roche Applied Sciences, Indianapolis, IN), 4 mM MgCl<sub>2</sub>, 0.4 µM of each primer and 0.2 µM of probe. The LightCycler thermal cycling conditions were 95°C for 8 min followed by 45 cycles of 95°C for 5 sec and 60°C for 5 sec ending in a 45°C hold for 1 min 15 sec. A panel of 54 well characterized *Brucella* strains and 28 near-neighbors, including 5 *Ochrobactrum* strains [31] were evaluated by the assay. Positive results are expressed in log scale as crossing threshold values (Ct) of fluorescence released above the no-template control baseline of 0.01 following each amplification as described by the manufacturer.

##### **16S rRNA gene analysis**

The full length amplicon of 16S rRNA gene was generated using the BO2 cell-lysate DNA and sequenced using the BigDye terminator cycle 3.1 sequencing kit (ABI, Foster City, CA) as described previously [31]. A comparative full-length sequence analysis of BO2 was performed with the consensus 16S rRNA gene sequence of *Brucella* spp. [31], and the *Ochrobactrum*

*intermedium* type strain (GeneBank accession no. AM114411T) along with that of the *B. inopinata* BO1<sup>T</sup> strain (GeneBank accession no. EU053207) using the GCG Wisconsin software package (version 10.2; Accelrys, San Diego, CA) and MEGA 4.0 [31,46].

##### **Omp2a/2b and recA genes analysis**

The full-length outer membrane porin genes *omp2a* and *omp2b*, and also the *recA* gene of BO2 were sequenced [33,45], and compared with sequences of BO1<sup>T</sup> and other *Brucella* and *Ochrobactrum* spp. available in GenBank. Contigs were assembled and edited before multiple sequence alignments were constructed in the DNASTAR Lasergene 8 genetic analysis software suite (DNASTAR Inc., Madison, WI). Neighbor-joining consensus trees inferred from 1000 bootstrap replicates were constructed using MEGA version 4.0 [46].

##### **MLSA typing**

To assess the relation of BO2 with other classical *Brucella* spp. and BO1<sup>T</sup>, the multi locus sequence analysis (MLSA) primer sets were used to amplify and sequence nine discrete house-keeping genes as described previously [47]. Multiple sequences were aligned and neighbor-joining phylogenetic trees were constructed as described above.

##### **Sequence identities**

Similarity values reported throughout the text and in Table 2 were calculated from estimates of evolutionary divergence between the sequences represented in the corresponding dendograms. All results are based on the pairwise analysis of inclusive sequences using the Maximum Composite Likelihood method in MEGA 4.0 [46]. All positions containing gaps and missing data were eliminated from the dataset.

##### **MLVA typing**

Molecular typing of the BO2 strain based on multiple-locus variable-number tandem repeat (VNTR) analysis (MLVA) was investigated by examining fifteen *Brucella* spp. VNTR genetic markers (MLVA-15) [48,49], and a distance tree was generated in BioNumerics v.5.1 (Applied Maths, Saint-Martens-Latem, Belgium) by clustering analysis using the unweighted-pair group method with arithmetic averages (UPGMA) and saved in newick format. Tree manipulations and labeling were done in MEGA 4.0 [46].

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#### Authors' contributions

SG, SCB, AJ, JB CC participated in the clinical diagnosis, isolation and initial characterization of the strain BO2 and also contributed in drafting the manuscript. RVT, JEG, DRL, ARH, BKD performed both biochemical and molecular studies and drafted the manuscript. All authors read and approved the final manuscript.

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#### References

1. Boschiroli ML, Foulongne V, O'Callaghan D: **Brucellosis: a worldwide zoonosis.** *Curr Opin Microbiol* 2001, **4**(1):58-64.
2. Corbel MJ: **Brucellosis: an overview.** *Emerg Infect Dis* 1997, **3**(2):213-221.
3. Osterman B, Moriyon I: **International Committee on Systematics of Prokaryotes; Subcommittee on the taxonomy of *Brucella*: Minutes of the meeting, 17 September 2003, Pamplona, Spain.** *Int J Syst Evol Microbiol* 2006, **56**:1175.
4. Cloeckaert A, Verger JM, Grayon M, Paquet JY, Garin-Bastuji B, Foster G, Godfroid J: **Classification of *Brucella* spp. isolated from marine mammals by DNA polymorphism at the omp2 locus.** *Microbes Infect* 2001, **3**(9):729-738.
5. Jahans KL, Foster G, Broughton ES: **The characterisation of *Brucella* strains isolated from marine mammals.** *Vet Microbiol* 1997, **57**(4):373-382.
6. Scholz HC, Hubalek Z, Sedlacek I, Vergnaud G, Tomaso H, Al Dahouk S, Melzer F, Kampfer P, Neubauer H, Cloeckaert A, et al: ***Brucella microti* sp. nov., isolated from a common vole *Microtus arvalis*.** *Int J Syst Evol Microbiol* 2008, **58**(Pt 2):375-382.
7. Scholz HC, Nockler K, Gollner C, Bahn P, Vergnaud G, Tomaso H, Al Dahouk S, Kampfer P, Cloeckaert A, Maquart M, et al: ***Brucella inopinata* sp. nov., isolated from a breast implant infection.** *Int J Syst Evol Microbiol* .
8. De BK, Stauffer L, Koylass MS, Sharp SE, Gee JE, Helsel LO, Steigerwalt AG, Vega R, Clark TA, Daneshvar M, et al: **Novel *Brucella* strain (BO1) associated with a prosthetic breast implant infection.** *J Clin Microbiol* 2008, **46**(1):43-49.
9. Verger JM, Grimont F, Grimont PAD, Grayon M: ***Brucella*, a monospecific genus as shown by deoxyribonucleic acid hybridization.** *Int J Syst Evol Microbiol* 1985, **35**:292-295.
10. Verger JM, Grimont F, Grimont PA, Grayon M: **Taxonomy of the genus *Brucella*.** *Ann Inst Pasteur Microbiol* 1987, **138**(2):235-238.
11. Grimont F, Verger JM, Cornelis P, Limet J, Lefevre M, Grayon M, Regnault B, Van Broeck J, Grimont PA: **Molecular typing of *Brucella* with cloned DNA probes.** *Res Microbiol* 1992, **143**(1):55-65.
12. Cerri D, Ebani VV, Pedrini A, Nuvoloni R, Renzoni G, Andreani E, Farina R: **Epididymitis by *Brucella ovis*: experimental infection in virgin ram lambs.** *New Microbiol* 1999, **22**(3):227-231.
13. Davis CE, Troy SB: **Brucellosis.** *N Engl J Med* 2005, **353**(10):1071-1072, author reply 1071-1072.
14. Fenkci V, Cevrioglu S, Yilmazer M: **Ovarian abscess due to *Brucella melitensis*.** *Scand J Infect Dis* 2003, **35**(10):762-763.
15. Pappas G, Akritidis N, Bosilkovski M, Tsianis E: **Brucellosis.** *N Engl J Med* 2005, **352**(22):2325-2336.
16. Troy SB, Rickman LS, Davis CE: **Brucellosis in San Diego: epidemiology and species-related differences in acute clinical presentations.** *Medicine (Baltimore)* 2005, **84**(3):174-187.
17. El-Olemy GM, Atta AA, Mahmoud WH, Hamzah EG: **Brucellosis in man-II. Isolation of the causative organisms with special reference to blood picture and urine constituents.** *Dev Biol Stand* 1984, **56**:573-578.
18. El-Olemy GM, Atta AA, Mahmoud WH, Hamzah EG: **Brucellosis in man. I. Serological diagnosis.** *Dev Biol Stand* 1984, **56**:565-572.
19. Quaipe RA: **Brucellosis in man.** *J Med Lab Technol* 1969, **26**(4):349-357.
20. Corbel MJ: **Recent advances in brucellosis.** *J Med Microbiol* 1997, **46**(2):101-103.
21. Al-Anazi AR, Aziz S, Fouda MA: **Brucellosis: haemorrhagic pleural effusion.** *Med Princ Pract* 2005, **14**(2):118-120.
22. Hatipoglu CA, Bilgin G, Tulek N, Kosar U: **Pulmonary involvement in brucellosis.** *J Infect* 2005, **51**(2):116-119.
23. Ohshimo S, Theegarten D, Totsch M, Moege J, Peitgen K, Guzman J, Costabel U: **Esophageal sarcoidosis presenting as pseudodiverticulum.** *Sarcoidosis Vasc Diffuse Lung Dis* 2008, **25**(1):64-67.
24. Olukman O: **Pulmonary involvement in childhood brucellosis: a case report.** *Vector Borne Zoonotic Dis* 2008, **8**(2):245-248.
25. Theegarten D, Albrecht S, Totsch M, Teschler H, Neubauer H, Al Dahouk S: **Brucellosis of the lung: case report and review of the literature.** *Virchows Arch* 2008, **452**(1):97-101.
26. Webb WA, Thoroughman JC: **Solitary pulmonary nodule due to *Brucella suis*. Report of a case.** *Dis Chest* 1966, **49**(2):222-224.
27. Park KW, Kim DM, Park CY, Kim HL, Jang SJ, Choi YS, Park MY, Song HJ, Lee SH: **Fatal systemic infection with multifocal liver and lung nodules caused by *Brucella abortus*.** *Am J Trop Med Hyg* 2007, **77**(6):1120-1123.
28. Alton GG, Jones LM, Pietz DE: **Laboratory techniques in Brucellosis.** *Monogr Ser World Health Organ* 1975, **55**: 1-163.
29. Institute/NCCLS CLSI ed.: **Performance standards for antimicrobial susceptibility testing-19th informational supplement-M100-S19** Wayne, PA: CLSI 2009.
30. Jevitt LA, Weigel LM, De B, Popovic T, Patel JB: **Development of a broth microdilution procedure for antimicrobial susceptibility testing of *Brucella* spp., abstr. C-357. Abstr. 105th Gen Meet Am soc Microbiol 2005: 2005; Atlanta, GA American Society for Microbiology, Washington, D.C. 2005.**
31. Gee JE, De BK, Levett PN, Whitney AM, Novak RT, Popovic T: **Use of 16S rRNA gene sequencing for rapid confirmatory identification of *Brucella* isolates.** *J Clin Microbiol* 2004, **42**(8):3649-3654.
32. Paquet JY, Diaz MA, Genevrois S, Grayon M, Verger JM, de Bolle X, Lakey JH, Letesson JJ, Cloeckaert A: **Molecular, antigenic, and functional analyses of Omp2b porin size variants of *Brucella* spp.** *J Bacteriol* 2001, **183**(16):4839-4847.
33. Scholz HC, Al Dahouk S, Tomaso H, Neubauer H, Witte A, Schloter M, Kampfer P, Falsen E, Pfeiffer M, Engel M: **Genetic diversity and phylogenetic relationships of bacteria belonging to the *Ochrobactrum-Brucella* group by *recA* and 16S rRNA gene-based comparative sequence analysis.** *Syst Appl Microbiol* 2008, **31**(1):1-16.
34. Batchelor BI, Brindle RJ, Gilks GF, Selkon JB: **Biochemical mis-identification of *Brucella melitensis* and subsequent laboratory-acquired infections.** *The Journal of hospital infection* 1992, **22**(2):159-162.
35. Elsaghir AA, James EA: **Misidentification of *Brucella melitensis* as *Ochrobactrum anthropi* by API 20NE.** *J Med Microbiol* 2003, **52**(Pt 5):441-442.
36. Cloeckaert A, Grayon M, Grepinet O: **An IS711 element downstream of the bp26 gene is a specific marker of *Brucella* spp. isolated from marine mammals.** *Clin Diagn Lab Immunol* 2000, **7**(5):835-839.
37. Halling SM, Tatum FM, Bricker BJ: **Sequence and characterization of an insertion sequence, IS711, from *Brucella ovis*.** *Gene* 1993, **133**(1):123-127.
38. Maquart M, Zygmunt MS, Cloeckaert A: **Marine mammal *Brucella* isolates with different genomic characteristics display a differential response when infecting human macrophages in culture.** *Microbes and infection/ Institut Pasteur* 2009, **11**(3):361-366.
39. Gurtler V, Mayall BC: **Genomic approaches to typing, taxonomy and evolution of bacterial isolates.** *Int J Syst Evol Microbiol* 2001, **51**(Pt 1):3-16.
40. Thompson CC, Thompson FL, Vandemeulebroecke K, Hoste B, Dawyndt P, Swings J: **Use of *recA* as an alternative phylogenetic marker in the family *Vibrionaceae*.** *Int J Syst Evol Microbiol* 2004, **54**(Pt 3):919-924.
41. Scholz HC, Tomaso H, Dahouk SA, Witte A, Schloter M, Kampfer P, Falsen E, Neubauer H: **Genotyping of *Ochrobactrum anthropi* by *recA*-based comparative sequence, PCR-RFLP, and 16S rRNA gene analysis.** *FEMS Microbiol Lett* 2006, **257**(1):7-16.
42. Cloeckaert A, Grayon M, Verger JM, Letesson JJ, Godfroid F: **Conservation of seven genes involved in the biosynthesis of the lipopolysaccharide O-side chain in *Brucella* spp.** *Res Microbiol* 2000, **151**(3):209-216.
43. Cloeckaert A, Grepinet O, Salih-Alj Debbah H, Zygmunt MS: **Overproduction of the *Brucella melitensis* heat shock protein DnaK in *Escherichia coli* and its localization by use of specific monoclonal antibodies in *B. melitensis* cells and fractions.** *Res Microbiol* 1996, **147**(3):145-157.
44. Cloeckaert A, Jacques I, Grillo MJ, Marin CM, Grayon M, Blasco JM, Verger JM: **Development and evaluation as vaccines in mice of *Brucella melitensis* Rev.1 single and double deletion mutants of the bp26 and**

- omp31 genes coding for antigens of diagnostic significance in ovine brucellosis. *Vaccine* 2004, **22**(21-22):2827-2835.
45. Cloeckaert A, Verger JM, Grayon M, Grepinet O: Restriction site polymorphism of the genes encoding the major 25 kDa and 36 kDa outer-membrane proteins of *Brucella*. *Microbiology* 1995, **141**(Pt 9):2111-2121.
46. Kumar S, Nei M, Dudley J, Tamura K: MEGA: a biologist-centric software for evolutionary analysis of DNA and protein sequences. *Brief Bioinform* 2008, **9**(4):299-306.
47. Whatmore AM, Perrett LL, MacMillan AP: Characterisation of the genetic diversity of *Brucella* by multilocus sequencing. *BMC Microbiol* 2007, **7**:34.
48. Huynh LY, Van Ert MN, Hadfield T, Probert WS, Bellaire BH, Dobson M, Burgess RJ, Weyant RS, Popovic T, Zanecki S, *et al*: Multiple Locus Variable Number Tandem Repeat (VNTR) Analysis (MLVA) of *Brucella* spp. identifies species specific markers and insights into phylogenetic relationships. *National Institute of Allergy and Infectious Disease, NIH: Frontiers in Research* 2008.
49. Tiller RV, De BK, Boshra M, Huynh LY, Van Ert MN, Wagner DM, Klena J, MT S, El-Shafie SS, Keim P, *et al*: Comparison of two multiple locus variable number tandem repeat (VNTR) analysis (MLVA) methods for molecular strain typing human *Brucella melitensis* isolates from the Middle East. *Journal of Clinical Microbiology* 2009, **47**(7):2226-2231.

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