

REVIEW

Open Access



X-ray cross-complementing family: the bridge linking DNA damage repair and cancer

Qiang Liu^{1,2,3†}, Qiu Peng^{1†}, Bin Zhang^{1,4*} and Yueqiu Tan^{1,2,3*}

Abstract

Genomic instability is a common hallmark of human tumours. As a carrier of genetic information, DNA is constantly threatened by various damaging factors that, if not repaired in time, can affect the transmission of genetic information and lead to cellular carcinogenesis. In response to these threats, cells have evolved a range of DNA damage response mechanisms, including DNA damage repair, to maintain genomic stability. The X-ray repair cross-complementary gene family (XRCC) comprises an important class of DNA damage repair genes that encode proteins that play important roles in DNA single-strand breakage and DNA base damage repair. The dysfunction of the XRCC gene family is associated with the development of various tumours. In the context of tumours, mutations in XRCC and its aberrant expression, result in abnormal DNA damage repair, thus contributing to the malignant progression of tumour cells. In this review, we summarise the significant roles played by XRCC in diverse tumour types. In addition, we discuss the correlation between the XRCC family members and tumour therapeutic sensitivity.

Keywords X-ray repair cross-complementary gene, XRCC, Cancer, DNA damage repair, Genomic instability

Background

Genomic instability, a hallmark of cancer, ensues from a complex interplay involving DNA damage, tumour-specific flaws in DNA repair, and the inability to halt

or impede the cell cycle prior to transmitting damaged DNA to daughter cells [1, 2]. Human DNA is exposed to tens of thousands of instances of damage each day, arising from both endogenous and exogenous factors, such as metabolites, ionising radiation (IR), ultraviolet (UV) light, and DNA damage resulting from replication errors [3–5]. Unrepaired DNA damage can significantly elevate the risk of various cancers, including breast, ovarian, prostate, and glioma, among others [6–9]. To maintain genome stability, cells adopt several measures to repair damaged DNA.

DNA damage repair (DDR) is one of the most critical biological responses in living organisms. The DNA repair pathway is usually a multi-step, nonlinear reaction involving a series of repair factors that work together in a time-series [10]. The DDR system contains five major repair pathways: base excision repair (BER), homologous recombination (HR), mismatch repair (MMR), nucleotide excision repair (NER), and non-homologous end-joining

[†]Qiang Liu and Qiu Peng contributed equally to the work.

*Correspondence:

Bin Zhang

1352009538@qq.com

Yueqiu Tan

tanyueqiu@csu.edu.cn

¹ Hunan Cancer Hospital and the Affiliated Cancer Hospital of Xiangya School of Medicine, Central South University, 283 Tongzipo Road, Changsha 410013, Hunan, China

² NHC Key Laboratory of Human Stem Cell and Reproductive Engineering, School of Basic Medical Sciences, Institute of Reproductive and Stem Cell Engineering, Central South University, Changsha 410078, Hunan, China

³ Clinical Research Center for Reproduction and Genetics in Hunan Province, Reproductive and Genetic Hospital of CITIC-Xiangya, Changsha 410008, Hunan, China

⁴ Department of Histology and Embryology, Xiangya School of Medicine, Central South University, Changsha 410013, Hunan, China



(NHEJ) [11]. Among all types of DNA damage, DNA double-strand breaks (DSB) are the most severe type of damage, and their efficient repair is essential for maintaining genome stability. There are two major DSB repair pathways in eukaryotes: HR and NHEJ [12, 13]. Mutations or aberrant expression of DDR-related genes result in compromised DNA damage repair functions, thereby reducing the capability of cells to repair damages caused by endogenous and exogenous stimuli. This fosters the accumulation of genetic alterations, ultimately leading to tumorigenesis [14]. DNA damage and abnormal DDR function not only contribute to tumorigenesis but also present opportunities and targets for tumour treatment. Many antitumour drugs operate in close association with the DNA damage and repair systems [15].

The DNA repair system is a vast and intricate network closely intertwined with all aspects of life, yet it remains inadequately understood. To date, several repair-related genes have been identified; however, their specific functions are not well understood. Among these, X-ray cross-complementing (XRCC) genes are some of the most studied DNA repair genes, and their abnormal expression has been reported to be associated with the development of various malignancies [16–21]. The XRCC gene

family comprises 11 main members (XRCC1–11), primarily responsible for maintaining chromosome stability by participating in DNA single-strand break repair [22, 23]. Among them, XRCC1–6 is a recognized member of the XRCC family, highly expressed in various tumour tissues and exhibiting multiple mutations in pan-cancer (Figs. 1 and 2). In addition, they play different biological functions in different cancer types (Table 1). In this review, we comprehensively elucidate the functions of the XRCC gene family in DNA damage repair, delving into their underlying mechanisms, and exploring their significant roles in tumour progression. In addition, we discuss the role of the XRCC gene family in the context of therapeutic sensitivities.

Structure and biological properties of the XRCC gene family

The XRCC family constitutes an essential group of DNA double-stranded break repair-related genes, responsible for encoding proteins involved in homologous recombination, which is indispensable for maintaining chromosomal stability and accomplishing DNA damage repair [24]. When DNA damage occurs, different XRCC genes participate in distinct DNA damage repair

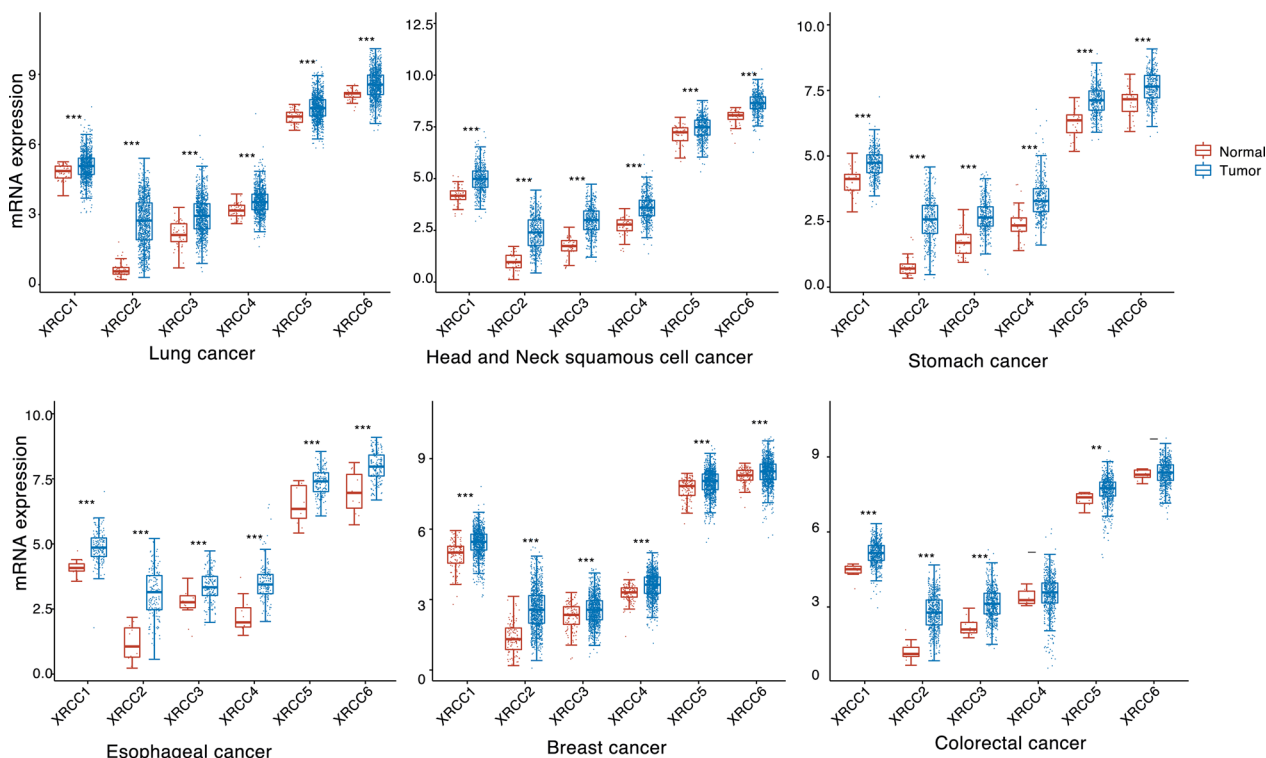


Fig. 1 XRCC1-XRCC6 is abnormally expressed in a variety of tumours. The RNA-seq data of the tumours shown in the figure were obtained using The Cancer Genome Atlas (TCGA) database, and the expression levels of XRCC1–6 in tumour tissues and normal tissues were analysed, where the horizontal coordinates represent different genes and the vertical coordinates represent the gene expression distribution. Different colours represent different groups. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$



Fig. 2 Genetic alterations of XRCC1–6 in pan-cancer. Analysis of XRCC1–XRCC6 mutations in pan-cancer using the cBioPortal database (<https://www.cbioportal.org/>)

pathways. In the context of double-stranded DNA damage repair, XRCC2, 3, and 11 operate through the HR pathway, whereas XRCC4, 5, 6, and 7 operate through the NHEJ pathway [23]. Notably, among the 11 members of the XRCC family, the probability of XRCC7 (PRKDC), XRCC8, XRCC9 (FANCG), XRCC10, and XRCC11 (BRCA2) belonging to this family remains controversial [22].

XRCC1 is located on chromosome 19q13.2–13.3, exhibits a total length of approximately 33 kb and contains 17 exons [25]. The *XRCC1* encodes a protein with three functional domains: the N-terminal domain, the BRCA1 carboxyl-terminal (BRCT) I domain, and the C-terminal BRCT II domain (Fig. 3), which interact with DNA polymerase beta, DNA ligase III, and poly(ADP-ribose) polymerase (PARP) to form a complex that acts as a “scaffolding protein” in the base excision repair process [23]. Human *XRCC1* was the first isolated mammalian repair gene reported to be associated with the repair of DNA damage caused by ionising radiation. In 1990, *XRCC1* was cloned by Thompson et al. from the gene library of EM9 cells [26]. In EM9 cells, the DNA ligase activity is reduced. Exposure to ionising radiation or ethyl methanesulfonate (EMS) led to impaired DNA strand breakage ligation and an elevated frequency of sister chromatid exchange (SCE). However, the introduction of *XRCC1* rectified the deficiency in the DNA repair capacity of this particular cell line.

Human *XRCC2* is located at 7q36.1 and contains three exons [27]. *XRCC2* is a newly discovered member of the

RecA/Rad51 family of recombinant repair proteins. It is highly conserved in mammals and humans and encompasses the characteristic ATP-binding region typical of the Rad51 family [28]. The functions of *XRCC2* include recruitment of the core protein Rad51 to the broken end of DNA, enhancement of Rad51 activity, maintenance of chromosome stability, and repair of DNA damage [29]. The loss of *XRCC2* expression can result in a defect in the core protein RAD51, leading to a significant reduction in the homologous recombination repair (HRR) function, particularly concerning DNA double-strand breaks. As a consequence, DNA damage cannot be effectively and timely repaired, giving rise to a considerably increased risk of chromosomal aberrations and abnormal chromosomal separation [30].

Human *XRCC3* is located on chromosome 14 q32.3, and the protein it encodes is involved in the recombination repair process of DNA double-strand breaks. The function of *XRCC3* was first identified in irs1SF cells, a Chinese hamster ovary (CHO) cell line. Transfection of the cloned *XRCC3* cDNA into irs1SF cells significantly improved chromosomal instability and reduced the sensitivity of irs1SF cells to various mutagens [31]. Liu et al. sequenced *XRCC3* and found homology with RAD51, a repair and recombination gene in eukaryotic cells; they further demonstrated the interaction between the two encoded proteins through a series of basic experiments. This indicates that the *XRCC3* protein belongs to the RAD51-related protein family and plays a key role in the homologous recombination process, essential for

Table 1 The list of XRCCs and their biological functions in different cancer types

XRCCs	Cancer type	Biological function	Mechanism	References
XRCC1	Glioma	Proliferation, migration, invasion, and angiogenesis	Targeting MMP-2, cyclin D1, VEGF, and p16	[139]
XRCC1	Gastric cancer	Induction of cisplatin resistance	Targeting thioredoxin-like protein 1 (TXNL1)	[127]
XRCC1	Pancreatic cancer	Induction of apoptosis	Targeting base excision repair pathway	[140]
XRCC1	Clear cell renal cell carcinoma	Regulating tumour metastasis	Regulating the expression of MMP-2, MMP-9	[74]
XRCC1	Lung cancer	Tumour metastasis	Regulating the expressions of E-cadherin, N-cadherin, and vimentin	[113]
XRCC2	Hepatocellular carcinoma	Proliferation	Repairing mitochondrial DNA damage	[16]
XRCC2	Colorectal cancer	Cell growth, cell cycle progression, and apoptosis	Regulating bcl-2 expression	[141]
XRCC3	Glioma	Temozolomide resistance	Promoting DNA double-strand break repair	[18]
XRCC3	Esophageal squamous cell carcinoma	Improvement in radiotherapy effect	Promoting DNA damage repair and/or enhancing Telomere stability	[134]
XRCC3	Breast cancer	Induction of cisplatin resistance sensitization of chemotherapeutic	Stimulating Rad51-related recombinational repair	[132]
XRCC4	Retinoblastoma	Drugs development	Regulating DNA damage repair	[135]
XRCC4	Medulloblastomas	Tumour growth	Regulating Myc-family or Cyclin D2	[142]
XRCC5	Colorectal cancer	Cancer stemness and aggressiveness	Promoting cyclooxygenase-2 expression. In cooperation with p300	[143]
XRCC5	Colorectal cancer	Proliferation	Activating cyclooxygenase-2 expression and enhanced prostaglandin E2 production	[144]
XRCC6	Osteosarcoma	Proliferation and metastasis	Promoting β -catenin/Wnt signalling pathway	[145]
XRCC6	Hepatocellular carcinoma	Promotion of the transformation of pre-cancerous hepatocytes and hepatocellular carcinoma development	Regulating the Wnt/ β -catenin pathway	[146]
XRCC6	Hepatocellular carcinoma		Inducing an effective autophagic degradation	[17]

MMP matrix metalloproteinase, VEGF vascular endothelial growth factor, XRCC X-ray repair cross-complementing

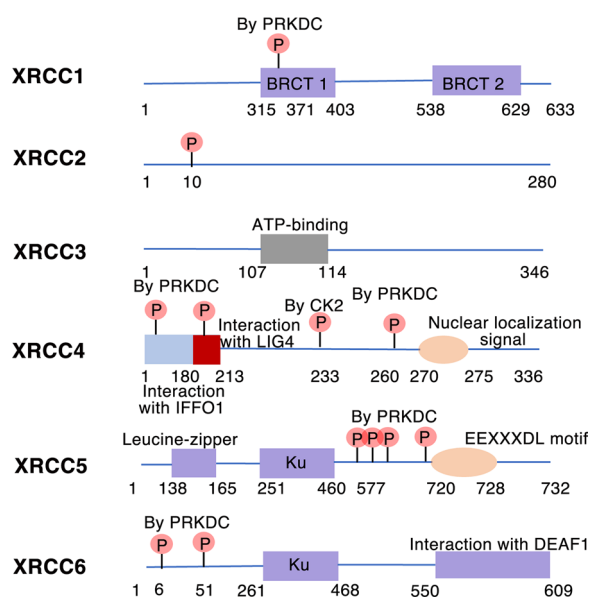


Fig. 3 List and domain structures of XRCC1–6

preserving chromosome stability and repairing DNA damage [32].

Human *XRCC4* is located on chromosome 5q11.2–13.3 and encodes a 336 amino acid protein (Fig. 3). It exhibits a spherical N-terminal head structural domain comprising seven peptide chains folded into a flared β -barrel, which is further connected to a long helix tail. The process of polymerisation involves the association of the two head regions and the initial segments of their helix tails [33]. *XRCC4* is an important NHEJ regulatory protein that directly interacts with Ku70/Ku80 in the repair pathway by preventing the degradation of free damaged DNA ends [34, 35]. *XRCC4* can form a complex with DNA ligase IV and XLE, and then form an elastic link between Ku70/Ku80 and DNA ligase IV, guiding the damaged DNA ends to join each other, so that DNA can be repaired [36, 37].

Human *XRCC5* is located at 2q33–34 and encodes a 732-amino acid protein with a molecular mass of approximately 86 kDa [38] (Fig. 3). *XRCC5*, also known as Ku80,

together with XRCC6 (Ku70) constitutes the XRCC5/XRCC6 heterodimer (Ku80/Ku70), which is a DNA-dependent protein kinase complex [37, 38]. The XRCC5/XRCC6 dimer binds to DNA double-stranded break ends and serves as an essential component of DNA nonhomologous end-joining repair [39].

Single nucleotide polymorphisms (SNPs) in the XRCC family and tumour susceptibility

Single nucleotide polymorphisms (SNPs) are alterations in DNA sequence that are caused by variations in a single base at the genomic level. As the most common form of genetic variation, SNPs are commonly found in the human genome and constitute more than 90% of all variations in human genomic DNA, with an average of one genotypic polymorphic SNP per thousand bases [40, 41]. SNPs may be found in both the coding and non-coding sequences of genes. SNPs located in the coding regions of genes, specifically those genes encoding immune response factors, have the potential to impact differences in gene expression or alter the structure of proteins they encode [42, 43]. Numerous studies have highlighted the potential function of SNPs, such as their impact on gene or protein modifications, promoter activity, and the modification of transcription factor binding sites. Moreover, SNPs can also influence the subcellular localisation of RNA and/or proteins. In addition, SNPs are associated with certain human traits and can influence an individual's susceptibility to specific diseases. Therefore, conducting an in-depth study of disease-associated SNPs and disease-susceptibility genes, along with analysing their functions, can significantly improve disease prevention strategies [44–47]. SNPs in the XRCC family of proteins

play a significant role in causing individual variations in DNA damage repair ability, which in turn determines an individual's susceptibility to tumours. Consequently, it is imperative to investigate genetic polymorphisms and tumour susceptibility and to explore specific molecular markers for the early diagnosis and treatment of tumours (Table 2).

Extensive research on XRCC1 SNPs has unequivocally established their correlation with tumour risk, treatment response, and survival outcomes in diverse malignancies, including lung cancer and gastric cancer [48–51]. Several SNPs have been detected within the coding region of *XRCC1* that result in corresponding amino acid changes in the encoding protein. The C→T base transition in exon 6 of *XRCC1* results in the conversion of the amino acid encoded by codon 194 from Arg to Trp, leading to the formation of the XRCC1 Arg194Trp gene polymorphism; the G→A base transition in exon 10 of *XRCC1* results in the conversion of the amino acid encoded by codon 399 from Arg to Gln, resulting in the formation of Arg399Gln gene polymorphism. Furthermore, the G→A base transition in exon 9 at position 27,466 results in the formation of Arg280His gene polymorphism [52]. *XRCC2* gene polymorphisms can potentially lead to alterations in the primary structure of XRCC2 or abnormal protein expression, resulting in impaired repair of DNA damage and increased susceptibility to cancer. Polymorphisms in *XRCC2* are associated with the development of various cancers, including lung, gastric, cervical, colon, breast, and others. Gok et al. reported that the Arg188His locus polymorphism of *XRCC2* was significantly associated with the development of gastric cancer. Furthermore, Perez et al. demonstrated that the rs3218536 locus

Table 2 The relationship between SNPs of XRCC1–6 and tumour

Genes	Variants	Position	Cancer types/functions	References
XRCC1	G>C	c.1517	Increased risk of hepatocellular carcinoma development	[147]
	Arg399Gln	exon 10	Genetic biomarker of squamous cell carcinoma of the head and neck	[148]
	Arg399Gln	exon 10	Increased childhood risk of acute lymphoblastic leukemia	[149]
	G>A	c. 1196	Influence of colorectal cancer on the clinical outcomes of patients	[150]
XRCC2	R188H	rs3218536	Influence breast cancer risk and survival	[151]
	C>T	rs718282	Increased the cancer risk of endometrial cancer	[152]
XRCC3	Thr241Met	rs861539	Associated with the survival of glioblastoma multiforme patients	[153]
	A>G	rs1799796	Increased risk of prostate cancer	[154]
	A>G	rs1799794	Modulates the risk of head and neck cancer	[155]
XRCC4	G>T	c.1394	Associated with breast cancer development	[65]
	S110P	rs79561451	Influence the susceptibility of individuals to breast cancer	[156]
	A>G	rs1805377	Genetic markers of hepatocellular carcinoma	[157]
XRCC5	G>A	rs207906	Increased susceptibility to leukaemia	[158]
XRCC6	C>G	c.-1310	Associated with breast cancer risk and oestrogen exposure	[159]

SNP single nucleotide polymorphisms, XRCC X-ray repair cross-complementing

polymorphism of *XRCC2* was substantially associated with the risk of cervical cancer pathogenesis. In addition, Sirisena and Kluzniak reported that SNPs in *XRCC2* are associated with the risk of breast cancer pathogenesis [53–57]. *XRCC3* possesses multiple SNPs, and certain *XRCC3* SNPs have been inextricably linked to tumorigenesis, cancer progression, and susceptibility to treatment. These SNPs have the potential to serve as molecular indicators for predicting tumorigenesis and prognosis [58]. Several studies have demonstrated that the Thr241Met SNP of *XRCC3* is associated with susceptibility to various cancers, including lung, bladder, endometrial, and laryngeal cancers [59–63]. The SNP of *XRCC4* G1394T has been reported to be associated with colorectal carcinogenesis and susceptibility to lung and prostate cancer [64]. Furthermore, the c.1394G>T SNP in *XRCC4* is associated with the development of breast cancer in Filipinos [65]. This study suggests that SNPs of *XRCC5* are associated with the development and progression of various tumours. Liu et al. observed that rs828704, rs3770502, and rs9288516 SNPs in *XRCC5* are associated with an increased risk of glioma susceptibility [66]. Hayden et al. observed that individuals carrying the TT genotype exhibited a reduced risk of myeloma compared with those carrying the *XRCC5* rs2440 CC genotype [67]. The structure and function of *XRCC6* are regulated by multiple SNPs and are closely associated with the development and progression of several tumours. Numerous studies have reported that SNPs of *XRCC6* are associated with genetic susceptibility to various cancers, including head and neck, bladder, lung, kidney, prostate, oral, and gastric cancers [68–71]. In addition, *XRCC7* SNP at allele 3434Thr has been reported to be associated with the risk of thyroid cancer in Iranian patients [72].

Role of XRCC in tumour metastasis

Metastasis refers to the process by which malignant tumour cells spread and establish secondary growths at distant sites from the primary tumour. The dissemination occurs through various means, including the lymphatic vessels, blood vessels, or body cavities from the primary site. Metastasis of malignant tumours is a major cause of death in cancer patients and a crucial factor affecting patient prognosis [73]. The XRCC family has been reported to regulate tumour metastasis by employing a variety of mechanisms. For instance, *XRCC1* is expressed at low levels in clear cell renal cell carcinoma (ccRCC) tissues in contrast to normal tissues. The ccRCC tissues with low *XRCC1* expression exhibit a positive correlation with lymph node metastasis and are associated with an unfavourable prognosis. Mechanistically, *XRCC1* inhibits tumour cell invasion and metastasis by regulating the expression of tissue

inhibitors of matrix metalloproteinase-2 (TIMP-2) and TIMP-1, leading to the suppression of the expression of metastasis-related markers matrix metalloproteinase-2 (MMP-2) and MMP-9 [74]. Additionally, the inhibition of *XRCC1* expression is associated with the progression of primary and metastatic melanoma [75]. The meta-analysis conducted by Bashir et al. revealed a significant downregulation of *XRCC2* in breast cancer tissues as opposed to non-cancerous healthy tissues. They also observed a significant correlation between *XRCC2* expression, lymph node status, and metastatic status in patients with breast cancer. These findings suggest that dysregulation of *XRCC2* in breast cancer could be utilized as a predictive indicator for lymph node metastasis and may serve as a therapeutic role in patients with breast cancer who are at risk of metastasis [76]. In colorectal cancer, the Thr241Met polymorphism of *XRCC3* is associated with time-to-metastasis and may potentially play a biological role in accelerating the metastatic process [77]. In breast cancer, scoring *XRCC4* expression using immunohistochemistry has proven to be effective in predicting postoperative breast cancer metastasis. In addition, the combined diagnosis of *XRCC4*, PARP1, and excision repair cross-complementation group 1 (ERCC1) has demonstrated considerable predictive capability in assessing the risk of breast cancer metastasis [78]. *XRCC5*, a downstream gene of miRNA-188-5p, was reported to be upregulated in glioma samples. In contrast, miRNA-188-5p was down-regulated in these samples, and patients with glioma exhibiting low miRNA-188-5p expression levels showed higher rates of distant metastasis. In addition, it is observed that miRNA-188-5p regulates glioma cell metastasis by suppressing *XRCC5* expression [79]. In hepatocellular carcinoma, there is a positive correlation between *XRCC5* expression level and the migration and invasion abilities of hepatocellular carcinoma cells. Inhibition of *XRCC5* expression leads to a significant reduction in the migration and invasion abilities of hepatocellular carcinoma cells. Additionally, high *XRCC5* expression is associated with tumour size, microvascular invasion, and lower overall survival time in the clinical samples of patients with hepatocellular carcinoma. Mechanistically, *XRCC5* regulates the expression of CTNNB1 (beta-catenin 1) and MMP9, which are key downstream target molecules of the Wnt/ β -catenin signalling pathway. Through this regulatory function, *XRCC5* promotes the progression of hepatocellular carcinoma [80]. Luo et al. reported that testicular expression 10 (TEX10) may potentially regulate cancer cell proliferation and metastatic processes through *XRCC6*, thereby controlling the Wnt/ β -catenin signalling pathway and DNA repair channels

[81]. These data suggest that the XRCC gene family plays an crucial role in tumour metastasis via multiple mechanisms.

Role of XRCC in tumour immunity

At present, tumour immunotherapy is the most promising strategy for cancer treatment. It is used to treat tumours by harnessing the body's immune system, enabling it to actively combat tumours, eradicate tumour cells, and establish sustained immune memory. Unlike targeted therapy, which focuses on specific targets, immunotherapy eliminates tumour cells by activating the body's immune system and utilising immunoactive substances and immune cells produced by the body [82, 83]. Several immune checkpoints associated with tumour immunity have been identified, including cytotoxic T-lymphocyte antigen 4 (CTLA-4), programmed death 1 (PD-1), programmed death ligand 1 (PD-L1), T-cell immunoglobulin and mucin-domain containing protein-3 (TIM3), and lymphocyte activating 3 (LAG3), among others [84–87]. Damaged DNA repair and associated genomic instability not only elevate mutagenicity and oncogenicity but also augment the neoantigenic load on the surface of tumour cells, thereby increasing their immunogenicity [88, 89]. The XRCC family is closely associated with tumour immunity.

In colorectal cancer samples, mutations of *XRCC1* were significantly correlated with adenomas. Aberrant *XRCC1* expression and mutations contribute to adenoma carcinogenesis. Moreover, PD-1/PD-L1 expression and CD4+ intraepithelial lymphocytes (IELs) are associated with tumour progression in patients possessing the wild-type *XRCC1*, suggesting that *XRCC1* expression is correlated with patient survival, tumour-infiltrating lymphocytes, and immune marker expression [90]. Using bioinformatics analysis, Li et al. observed that in breast cancer *XRCC2* and *XRCC3* are associated with the infiltration of immune cells, such as B cells, CD4+ T cells, CD8/CD4+ T cells, neutrophils, and dendritic cells, as well as the prognosis of patients with breast cancer [91]. In head and neck, lung and cervical cancers, the methylation status of *XRCC3* is associated with the expression of immune checkpoint molecules and inflammatory markers [92]. Guo et al. reported that retinoic acid-inducible gene I (RIG-I) can potentially be recruited to double-strand breaks (DSB) and inhibit NHEJ. Mechanistically, RIG-I hinders the formation of the XRCC4/LIG4/XLF complex on DSB by interacting with XRCC4, thereby disrupting DNA repair and rendering cancer cells sensitive to radiation therapy. XRCC4 enhances RIG-I oligomerization and ubiquitination to promote RIG-I signalling, thereby inhibiting RNA viral replication in host cells, indicating the crucial role played by XRCC4 in the innate

immune response [19]. The cGAS-STING pathway has emerged as a potential mechanism for the induction of inflammation-mediated tumorigenesis [93, 94]. Qi et al. reported that *XRCC5* and *XRCC6* are associated with the cGAS-STING pathway. Overexpression of *XRCC5* and *XRCC6* was significantly associated with the clinical stage and pathological grade of hepatocellular carcinoma. Moreover, they observed a significant correlation between the expression of *XRCC5* and *XRCC6* and the infiltration of B cells, CD4+ T cells, CD8+ T cells, macrophages, neutrophils, and dendritic cells in hepatocellular carcinoma [95]. In addition, the toll-like receptor 4 (TLR4)-mediated lack of immune activity inhibits the expression of *XRCC5* and *XRCC6* in response to damage by the carcinogen diethylnitrosamine (DEN). This effect leads to the impairment of DNA repair, facilitating the transformation of precancerous hepatocytes and the progression of HCC. In contrast, *XRCC6* expression prevents the development and progression of HCC by restoring the cellular senescence response and activating the immune network, thereby inducing efficient autophagic degradation, scavenging accumulated reactive oxygen species (ROS), reducing DNA damage, and attenuating proliferation [17, 96].

Role of XRCC in tumour metabolism

The abnormal metabolism of tumour cells is an important feature of tumours. As normal cells gradually develop into tumour cells, they acquire several hallmark capabilities. Abnormal alterations in energy metabolism are one of the primary hallmarks of malignancy [97]. Tumour cells perform several biosynthetic processes and metabolic activities in a metabolic reprogramming manner, providing energy and multiple substrates to support their rapid proliferation and survival [98]. The activation of oncogenes or inactivation of tumour suppressors drives the metabolic reprogramming of cancer cells, and the XRCC family plays a critical role in the tumour metabolic process.

In a recent study, Anurag et al. observed that proteomic analysis of pretreatment patient biopsies uniquely revealed metabolic pathways associated with drug resistance, including oxidative phosphorylation, lipogenesis, and fatty acid metabolism. Interestingly, proteogenomic analysis of somatic copy number aberrations identified a resistance-associated deletion in 19q13.31–33, which corresponded with the location of *XRCC1* [99]. Aldehyde dehydrogenase 2 (*ALDH2*) is also involved in lipid metabolism. Chen et al. found that the interaction between the base excision repair proteins, XRCC1 and ALDH2, was indicative of overall survival in patients diagnosed with lung and liver cancer [100]. Folic acid metabolism is associated with

the efficacy of platinum compounds [101, 102]. Folate metabolism involves DNA methylation mediated by the enzymes, tetrahydrofolate methylene reductase (MTHFR) and methionine synthase (MTR). Polymorphisms in *XRCC1* and folate metabolism genes can affect the prognosis of patients with non-small cell lung cancer [103]. In addition, polymorphisms in DNA repair genes (including *XRCC1*, *XRCC2*, and *XRCC3*) and steroid metabolism genes in patients undergoing prostate cancer radiotherapy are associated with clinically advanced toxicity [104].

Role of XRCC in autophagy

Autophagy is a process by which self-damaged organelles and proteins are separated into autophagic vesicles and transported to lysosomes for catabolism [105]. Autophagy is closely associated with various diseases and plays a complex role in tumours. Particularly, autophagy plays an oncogenic role in early-stage tumours. Additionally, stressors such as nutritional deficiency, DNA damage, and cytotoxic effects can potentially induce cellular autophagy and promote malignant tumour progression in advanced-stage tumours or during antitumour therapy. Recent studies have shown that autophagy plays a dual regulatory role in promoting and inhibiting tumour cell growth; thus, targeting autophagy may significantly affect the efficacy of antitumour therapy [105].

Ma et al. conducted a comprehensive analysis including a cohort of 47 patients with advanced or metastatic oesophageal cancer who underwent next-generation sequencing (NGS) between May 2017 and February 2020. This study resulted in the identification of 227 mutated genes. Among them, *XRCC1* exhibited a substantial number of mutations and was associated with autophagy [106]. Demirbag-Sarikaya et al. observed that the autophagy-related molecule autophagy-related protein 5 (ATG5) interacts with both *XRCC5* and *XRCC6*. This interaction is primarily mediated by *XRCC6*. They also found the interaction to be dynamic and enhanced under genotoxic stress. Moreover, they found that the interaction between ATG5 and *XRCC6* is essential for DNA repair and effective recovery from genotoxic stress. These results demonstrate a novel, direct, dynamic, and functional interaction between ATG5 and *XRCC6*, which are proteins that play critical roles in DNA repair under genotoxic stress conditions [107]. In addition, Wang et al. showed that the restoration of immunity supporting hepatocyte senescence and autophagy through *XRCC6* repair of DNA damage reverses the progression of TLR4-deficient deteriorating hepatocellular carcinoma [17, 96].

The influence of non-coding RNAs on XRCC

Non-coding RNA (ncRNA) is an emerging biomarker that exhibits correlations with tumorigenesis and possesses oncogenic or tumour-suppressing properties. It can be detected in serum, plasma, and other biological fluids, making it a promising therapeutic and prognostic target for tumours, due to its non-invasive nature, traumatic, high sensitivity, and specificity [108–110]. The ncRNAs, including long ncRNAs (lncRNAs), microRNAs (miRNAs), and circular RNAs (circRNAs), are extensively involved in tumour pathogenesis. The ncRNAs play a pivotal role in the biological processes of tumours by regulating cell growth and survival, EMT and metastasis, maintenance of tumour stem cells, metabolism, autophagy, chemoresistance, and angiogenesis [111, 112]. Several studies have reported that ncRNAs modulate tumour progression by regulating *XRCC*. In lung cancer, the circular RNA FLNA acts as a sponge for miR-486-3p and promotes tumour cell proliferation, migration, and invasion by regulating *XRCC1* expression [113]. Li et al. observed that miR-3940-5p enhances homologous recombination repair after DSB by down-regulating *XRCC2* expression [114]. In oesophageal cancer, microRNA-127-3p enhances the chemosensitivity of phenanthroline-dione derivatives by targeting *XRCC3* [115]. In glioma cells, the long non-coding RNA SBF2-AS1 acts as a ceRNA for miR-151a-3p, which in turn regulates the expression of *XRCC4*, thereby enhancing DSB repair [116]. Furthermore, in hepatocellular carcinoma cells, lncRNA NIHCOLE promotes the ligation efficiency of DSB by regulating *XRCC4* [117]. CircXRCC5 acts as a sponge for miR-490-3p and regulates the expression of the downstream target gene, *XRCC5*, thereby activating *CLC3* transcription and promoting glioma progression [118]. In breast cancer, miR-623 inhibits cell proliferation, migration and invasion by targeting *XRCC5* by down-regulating cell cycle protein-dependent kinases and inhibiting phosphatidylinositol-3 kinase (PI3K)/Akt and Wnt/ β -Catenin pathways [119]. The correlation between microRNA-379-5p and premature ovarian insufficiency has been reported to be mediated by PARP1 and *XRCC6* [120].

Role of XRCC in tumour therapeutic sensitivity

Platinum-based combination chemotherapy represents the first-line standard of care for numerous types of tumours. The primary mechanism of action of platinum-based drugs is the formation of platinum–DNA adducts binding to guanine, adenine, and cytosine on DNA. This process leads to the creation of inter-strand or intra-strand DNA cross-links, ultimately causing DNA damage and cell death [121]. Differences in DNA repair ability directly lead to inter-individual differences

in the sensitivity of tumour cells to DNA-related cytotoxic drugs [122]. Therefore, the relationship between DNA repair genes and tumour susceptibility to platinum-based chemotherapy may be crucial for guiding individualised clinical treatments. Similarly, the biological mechanism of killing tumour cells by radiation therapy is primarily based on direct genomic damage caused by radiation, resulting in the loss of the proliferative ability of tumour cells. Therefore, the clinical effect of radiation therapy depends on the responsiveness of tumour cells to radiation damage and their ability to repair the damage. However, tumour cells are highly capable of damage repair and can selectively recognise damage and initiate repair pathways, leading to tumour cell tolerance to radiation therapy and other antitumour drugs. Studies have demonstrated that DNA damage repair mechanisms protect tumour cells from radiation therapy-induced cell death, indicating that repair pathway proteins may play a potential role in enhancing tumour cell radiosensitivity. Exploring new approaches to more effectively inhibit repair proteins is crucial for enhancing tumour radiosensitivity [123].

DNA repair ability is associated with the Gln399Arg polymorphism in *XRCC1*. Patients with non-small cell lung cancer polymorphism may potentially be resistant to platinum [50, 124]. In a study involving 195 patients with epithelial ovarian cancer, it was observed that 45% of patients with *XRCC1*-positive tumours were resistant to platinum drugs. In contrast, only 17% of patients with *XRCC1*-negative tumours were resistant to platinum drugs. These findings suggest that *XRCC1* has clinical significance as a predictor of resistance to platinum therapy in patients with ovarian cancer [125]. Xu et al. reported that the methylation level of H3K4 is significantly reduced in drug-resistant cells. JIB-04, a chemical inhibitor of H3K4 demethylase, restores the methylation of H3K4, blocks the co-localisation of *XRCC1* and phosphorylation of H2AX (γ H2AX), and ultimately improves drug sensitivity. They also found that the expression level of KDM5B was significantly elevated in drug-resistant cells. Knockdown of KDM5B elevates the methylation level of H3K4, which hinders the localisation of *XRCC1* at the DNA damage site, resulting in heightened sensitivity [126]. Furthermore, in the context of gastric cancer, it has been reported that *XRCC1* expression was significantly elevated in cisplatin-resistant cells, and it independently promoted cisplatin resistance. Irinotecan, another chemotherapeutic agent that induces DNA damage, was used to treat patients with advanced gastric cancer who experienced progression on cisplatin therapy. Notably, irinotecan effectively inhibited *XRCC1* expression, resulting in increased sensitivity of resistant cells to cisplatin [127].

XRCC2 is indispensable for DNA repair following radiation damage. Radiation induces an abnormal increase in the expression level of *XRCC2* in lung cancer cells, which causes them to resist the damaging effects of radiation on tumour cell DNA. This results in the development of tumour resistance to radiotherapy [128, 129]. In glioblastoma, inhibition of *XRCC2* expression increases the radiosensitivity of tumour cells to radiation [130]. X-ray irradiation induces *XRCC2* expression in colorectal cancer cells and exhibits a dose- and time-dependent relationship between *XRCC2* expression and radiation exposure. Downregulation of *XRCC2* expression inhibits the proliferation of colorectal cancer cells and increases their sensitivity to radiation. In addition, gene silencing of *XRCC2* induces a decrease in the repair of radiation-induced cell damage, resulting in cellular arrest in the G2/M phase and increased apoptosis [131]. Expression abnormalities in *XRCC3* are associated with tumour resistance to DNA damage-inducing antitumour agents. *XRCC3* induces cisplatin resistance in tumour cells by activating Rad51-related recombination repair and S-phase monitor activation and by reducing apoptosis [132, 133]. *XRCC3* has been reported to protect glioma cells from temozolomide (TMZ)-induced cell death and cell cycle inhibition. In addition, *XRCC3* knockdown significantly reduces DSB repair after TMZ treatment, leading to increased drug sensitivity. This study confirms the importance of homologous recombination in conferring resistance to the methylating drug TMZ of glioma cells [18]. High *XRCC3* expression is positively associated with resistance to radiotherapy in oesophageal squamous cell carcinoma (ESCC) and is an independent predictor of short disease-specific survival in patients with ESCC. Knockdown of *XRCC3* in ESCC cells significantly improved the efficacy of radiotherapy in both in vitro and in vivo analyses. *XRCC3* overexpression significantly enhanced the resistance of ESCC cells to radiotherapy. Furthermore, the radiation resistance of *XRCC3* was mainly dependent on enhanced homologous recombination, telomere stabilisation, and ESCC cell death reduction mediated by radiation-induced apoptosis and mitotic mutations [134]. Overexpression of ubiquitin-like with PHD and RING finger domains 1 (UHRF1) increases *XRCC4* expression. Conversely, the downregulation of *XRCC4* renders retinoblastoma cells sensitive to etoposide treatment, indicating that *XRCC4* is a key mediator of drug sensitivity following UHRF1 consumption in retinoblastoma cells. Moreover, in retinoblastoma cells depleted of UHRF1, it was observed that the chromatin association of DNA ligase IV in response to acute DNA damage was significantly reduced. Functional complementation of *XRCC4* in cells depleted of UHRF1 weakens drug sensitivity, indicating that the downregulation

of XRCC4 in UHRF1-depleted cells impairs DNA repair, leading to a significant induction of apoptosis during treatment with genotoxic drugs [135]. Hori et al. investigated the relationship between NHEJ-related protein expression and the outcome of radiotherapy in oesophageal cancer. They employed immunohistochemical analysis of NHEJ-related proteins, including XRCC4, which holds promise as a potential predictive marker for assessing tumour radiosensitivity [136]. XRCC5 knockdown significantly enhanced the sensitivity of glioma cells to TMZ, whereas XRCC5 overexpression led to TMZ resistance in cancer cells. Both in vitro and in vivo experiments have shown that TMZ treatment induces XRCC5 expression in TMZ-resistant cells [137]. Chen et al. reported that the quercetin-targeted radiation-induced ARv7-mediated circNHS/miR-512-5p/XRCC5 signalling pathway increases radiosensitivity in prostate cancer [138].

Conclusions

Tumour development is the result of a complex interplay of various factors, and DNA damage is a significant contributor to this process. The XRCC gene family is a crucial group of genes involved in DNA damage repair responsible for maintaining the stability of genetic material and cellular function through their role in repairing DNA double-strand breaks and cross-link damage. Additionally, these genes play a significant role in ensuring the proper segregation of chromosomes during cell division. The XRCC family constitutes a group of susceptibility genes, and their polymorphisms are prevalent in the general population, exerting a substantial effect on tumorigenesis. An in-depth investigation of the correlation between XRCC gene polymorphisms and tumour development can help explore the interactions between related genes, as well as the interactions between genes and the environment. These investigations will substantially help in effectively formulating tumour prevention and treatment strategies, protecting the susceptible population to a larger extent, effectively reducing the incidence of tumours, and improving the cure rate of tumours. Although significant progress has been made, inconsistencies persist in the findings of several studies. Therefore, it is essential to increase the sample size and conduct a comprehensive population cohort study employing multivariate analysis of crucial prognostic factors, such as gender, age, smoking status, histopathological types, clinical stages, and treatment strategies. This approach enables the investigation of the correlation between gene polymorphisms and prognosis, as well as the interplay between genetic polymorphisms and environmental factors.

The mechanism of resistance to tumour radiotherapy and chemotherapy has been a popular research topic in

the field of oncology. DNA oxygenation and alkylation damage caused by numerous DNA-damaging anticancer drugs can be repaired via the XRCC gene family-mediated pathways. Research on the XRCC gene family and chemotherapeutic drug sensitivity is of particular interest. Inhibition of the XRCC gene family expression can sensitise various anticancer drugs, suggesting the XRCC gene family has the potential to influence the efficacy of tumor therapy by affecting chemotherapy sensitisation. However, the functions of these genes are not fully understood, and their relationship with anticancer drug sensitisation requires further exploration.

Abbreviations

XRCC	X-ray repair cross-complementary
IR	Ionizing radiation
UV	Ultraviolet
DDR	DNA damage repair
BER	Base excision repair
HR	Homologous recombination
MMR	Mismatch repair
NER	Nucleotide excision repair
NHEJ	Non-homologous end joining
DSB	DNA double-strand breaks
EMS	Ethyl methanesulfonate
SCE	Sister chromatid exchange
PARP	Poly(ADP-ribose) polymerase
SNP	Single nucleotide polymorphisms
ccRCC	Clear cell renal cell carcinoma
CTLA-4	Cytotoxic T-lymphocyte antigen 4
PD-1	Programmed death 1
PD-L1	Programmed death ligand 1
IELs	Intraepithelial lymphocytes
TLR4	Toll-like receptor 4
DEN	Diethylnitrosamine
ROS	Reactive oxygen species
ALDH2	Aldehyde dehydrogenase 2
MTR	Methionine synthase
NGS	Next-generation sequencing
ESCC	Esophageal squamous cell carcinoma
TMZ	Temozolomide

Acknowledgements

Not applicable.

Author contributions

QL, QP, BZ and YT collected the related paper and drafted the manuscript. BZ and YT revised and finalized the manuscript. All authors read and approved the final manuscript.

Funding

This work was supported in part by grants from the following sources: the National Natural Science Foundation of China (82203233), the Natural Science Foundation of Hunan Province (2022JJ70101, 2023JJ40413), the Research Project of Health Commission of Hunan Province (202302067467).

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Received: 27 March 2023 Accepted: 17 August 2023

Published online: 07 September 2023

References

- Huang RX, Zhou PK. DNA damage response signaling pathways and targets for radiotherapy sensitization in cancer. *Signal Transduct Target Ther.* 2020;5:60.
- Jackson SP, Bartek J. The DNA-damage response in human biology and disease. *Nature.* 2009;461:1071–8.
- Silas Y, Singer E, Das K, Lehming N, Pines O. A combination of Class-I fumarases and metabolites (alpha-ketoglutarate and fumarate) signal the DNA damage response in *Escherichia coli*. *Proc Natl Acad Sci USA.* 2021;118:e2026595118.
- Lemay JF, St-Hilaire E, Ronato DA, Gao Y, Belanger F, Gezzar-Dandashi S, Kimenyi Ishimwe AB, Sawchyn C, Levesque D, McQuaid M, et al. A genome-wide screen identifies SCA1 as a modulator of the UV-induced replicative stress response. *PLoS Biol.* 2022;20:e3001543.
- Khozooei S, Lettau K, Barletta F, Jost T, Rebholz S, Veerappan S, Franz-Wachtel M, Macek B, Iliakis G, Distel LV, et al. Fisetin induces DNA double-strand break and interferes with the repair of radiation-induced damage to radiosensitize triple negative breast cancer cells. *J Exp Clin Cancer Res.* 2022;41:256.
- Tarish FL, Schultz N, Tanoglidis A, Hamberg H, Letocha H, Karasz K, Hamdy FC, Granfors T, Helleday T. Castration radiosensitizes prostate cancer tissue by impairing DNA double-strand break repair. *Sci Transl Med.* 2015;7:312re311.
- Sunada S, Nakanishi A, Miki Y. Crosstalk of DNA double-strand break repair pathways in poly(ADP-ribose) polymerase inhibitor treatment of breast cancer susceptibility gene 1/2-mutated cancer. *Cancer Sci.* 2018;109:893–9.
- Maksoud S. The DNA double-strand break repair in glioma: molecular players and therapeutic strategies. *Mol Neurobiol.* 2022;59:5326–65.
- Karakashev S, Fukumoto T, Zhao B, Lin J, Wu S, Fatkhutdinov N, Park PH, Semenova G, Jean S, Cadungog MG, et al. EZH2 inhibition sensitizes CARM1-high, homologous recombination proficient ovarian cancers to PARP inhibition. *Cancer Cell.* 2020;37(157–167): e156.
- Hughes CD, Simons M, Mackenzie CE, Van Houten B, Kad NM. Single molecule techniques in DNA repair: a primer. *DNA Repair (Amst).* 2014;20:2–13.
- Ciccia A, Elledge SJ. The DNA damage response: making it safe to play with knives. *Mol Cell.* 2010;40:179–204.
- Chapman JR, Taylor MR, Boulton SJ. Playing the end game: DNA double-strand break repair pathway choice. *Mol Cell.* 2012;47:497–510.
- Chang HHY, Pannunzio NR, Adachi N, Lieber MR. Non-homologous DNA end joining and alternative pathways to double-strand break repair. *Nat Rev Mol Cell Biol.* 2017;18:495–506.
- Parry EM, Gable DL, Stanley SE, Khalil SE, Antonescu V, Florea L, Armanios M. Germline mutations in DNA repair genes in lung adenocarcinoma. *J Thorac Oncol.* 2017;12:1673–8.
- Helleday T, Petermann E, Lundin C, Hodgson B, Sharma RA. DNA repair pathways as targets for cancer therapy. *Nat Rev Cancer.* 2008;8:193–204.
- Zhao Z, He K, Zhang Y, Hua X, Feng M, Zhao Z, Sun Y, Jiang Y, Xia Q. XRCC2 repairs mitochondrial DNA damage and fuels malignant behavior in hepatocellular carcinoma. *Cancer Lett.* 2021;512:1–14.
- Wang Z, Lin H, Hua F, Hu ZW. Repairing DNA damage by XRCC6/KU70 reverses TLR4-deficiency-worsened HCC development via restoring senescence and autophagic flux. *Autophagy.* 2013;9:925–7.
- Roos WP, Frohnapfel L, Quiros S, Ringel F, Kaina B. XRCC3 contributes to temozolomide resistance of glioblastoma cells by promoting DNA double-strand break repair. *Cancer Lett.* 2018;424:119–26.
- Guo G, Gao M, Gao X, Zhu B, Huang J, Tu X, Kim W, Zhao F, Zhou Q, Zhu S, et al. Reciprocal regulation of RIG-I and XRCC4 connects DNA repair with RIG-I immune signaling. *Nat Commun.* 2021;12:2187.
- Gu Z, Li Y, Yang X, Yu M, Chen Z, Zhao C, Chen L, Wang L. Overexpression of CLC-3 is regulated by XRCC5 and is a poor prognostic biomarker for gastric cancer. *J Hematol Oncol.* 2018;11:115.
- Alblihi A, Ali R, Algethami M, Shoaqfi A, Toss MS, Brownlie J, Tatum NJ, Hickson I, Moran PO, Grabowska A, et al. Targeting Mre11 overcomes platinum resistance and induces synthetic lethality in XRCC1 deficient epithelial ovarian cancers. *NPJ Precis Oncol.* 2022;6:51.
- Fan Y, Gao Z, Li X, Wei S, Yuan K. Gene expression and prognosis of x-ray repair cross-complementing family members in non-small cell lung cancer. *Bioengineered.* 2021;12:6210–28.
- Caldecott KW. XRCC1 and DNA strand break repair. *DNA Repair (Amst).* 2003;2:955–69.
- Li D, Liu H, Jiao L, Chang DZ, Beinart G, Wolff RA, Evans DB, Hassan MM, Abbruzzese JL. Significant effect of homologous recombination DNA repair gene polymorphisms on pancreatic cancer survival. *Cancer Res.* 2006;66:3323–30.
- Siciliano MJ, Carrano AV, Thompson LH. Assignment of a human DNA-repair gene associated with sister-chromatid exchange to chromosome 19. *Mutat Res.* 1986;174:303–8.
- Thompson LH, Brookman KW, Jones NJ, Allen SA, Carrano AV. Molecular cloning of the human XRCC1 gene, which corrects defective DNA strand break repair and sister chromatid exchange. *Mol Cell Biol.* 1990;10:6160–71.
- Romanowicz H, Smolarz B, Baszczynski J, Zadrozny M, Kulig A. Genetics polymorphism in DNA repair genes by base excision repair pathway (XRCC1) and homologous recombination (XRCC2 and RAD51) and the risk of breast carcinoma in the Polish population. *Pol J Pathol.* 2010;61:206–12.
- O'Regan P, Wilson C, Townsend S, Thacker J. XRCC2 is a nuclear RAD51-like protein required for damage-dependent RAD51 focus formation without the need for ATP binding. *J Biol Chem.* 2001;276:22148–53.
- Tambini CE, Spink KG, Ross CJ, Hill MA, Thacker J. The importance of XRCC2 in RAD51-related DNA damage repair. *DNA Repair (Amst).* 2010;9:517–25.
- Park SW, Yoo NJ, Lee SH. Mutational analysis of mononucleotide repeats in XRCC2 and XRCC6 in cancers with microsatellite instability. *Pathology.* 2011;43:78–9.
- Fuller LF, Painter RB. A Chinese hamster ovary cell line hypersensitive to ionizing radiation and deficient in repair replication. *Mutat Res.* 1988;193:109–21.
- Tebbs RS, Zhao Y, Tucker JD, Scheerer JB, Siciliano MJ, Hwang M, Liu N, Legerski RJ, Thompson LH. Correction of chromosomal instability and sensitivity to diverse mutagens by a cloned cDNA of the XRCC3 DNA repair gene. *Proc Natl Acad Sci USA.* 1995;92:6354–8.
- Junop MS, Modesti M, Guarne A, Ghirlando R, Gellert M, Yang W. Crystal structure of the Xrcc4 DNA repair protein and implications for end joining. *EMBO J.* 2000;19:5962–70.
- Wu PY, Frit P, Meesala S, Dauvillier S, Modesti M, Andres SN, Huang Y, Sekiguchi J, Calsou P, Salles B, Junop MS. Structural and functional interaction between the human DNA repair proteins DNA ligase IV and XRCC4. *Mol Cell Biol.* 2009;29:3163–72.
- Pastwa E, Blasiak J. Non-homologous DNA end joining. *Acta Biochim Pol.* 2003;50:891–908.
- Ramsden DA, Gellert M. Ku protein stimulates DNA end joining by mammalian DNA ligases: a direct role for Ku in repair of DNA double-strand breaks. *EMBO J.* 1998;17:609–14.
- Gottlieb TM, Jackson SP. The DNA-dependent protein kinase: requirement for DNA ends and association with Ku antigen. *Cell.* 1993;72:131–42.
- Rathmell WK, Chu G. Involvement of the Ku autoantigen in the cellular response to DNA double-strand breaks. *Proc Natl Acad Sci USA.* 1994;91:7623–7.
- Lees-Miller SP, Godbout R, Chan DW, Weinfeld M, Day RS 3rd, Barron GM, Allalunis-Turner J. Absence of p350 subunit of DNA-activated protein kinase from a radiosensitive human cell line. *Science.* 1995;267:1183–5.

40. Li Y, Zhang F, Yang D. Comprehensive assessment and meta-analysis of the association between CTNNB1 polymorphisms and cancer risk. *Biosci Rep*. 2017. <https://doi.org/10.1042/BSR20171121>.
41. International HapMap Consortium, Frazer KA, Ballinger DG, Cox DR, Hinds DA, Stuve LL, Gibbs RA, Belmont JW, Boudreau A, Hardenbol P, et al. A second generation human haplotype map of over 3.1 million SNPs. *Nature*. 2007;449:851–61.
42. Dickinson AM, Norden J. Non-HLA genomics: does it have a role in predicting haematopoietic stem cell transplantation outcome? *Int J Immunogenet*. 2015;42:229–38.
43. Bogunia-Kubik K, Lacinia P. From genetic single candidate gene studies to complex genomics of GvHD. *Br J Haematol*. 2017;178:661–75.
44. Yige L, Dandan Z. Progress on functional mechanisms of colorectal cancer causal SNPs in post-GWAS. *Yi Chuan*. 2021;43:203–14.
45. Pittman AM, Naranjo S, Jalava SE, Twiss P, Ma Y, Olver B, Lloyd A, Vijayakrishnan J, Qureshi M, Broderick P, et al. Allelic variation at the 8q23.3 colorectal cancer risk locus functions as a cis-acting regulator of EIF3H. *PLoS Genet*. 2010;6:e1001126.
46. Dayeh TA, Olsson AH, Volkov P, Almgren P, Ronn T, Ling C. Identification of CpG-SNPs associated with type 2 diabetes and differential DNA methylation in human pancreatic islets. *Diabetologia*. 2013;56:1036–46.
47. Civelek M, Lusis AJ. Systems genetics approaches to understand complex traits. *Nat Rev Genet*. 2014;15:34–48.
48. Ruzzo A, Graziano F, Kawakami K, Watanabe G, Santini D, Catalano V, Bissonni R, Canestrari E, Ficarelli R, Menichetti ET, et al. Pharmacogenetic profiling and clinical outcome of patients with advanced gastric cancer treated with palliative chemotherapy. *J Clin Oncol*. 2006;24:1883–91.
49. Liu B, Wei J, Zou Z, Qian X, Nakamura T, Zhang W, Ding Y, Feng J, Yu L. Polymorphism of XRCC1 predicts overall survival of gastric cancer patients receiving oxaliplatin-based chemotherapy in Chinese population. *Eur J Hum Genet*. 2007;15:1049–53.
50. Gurubhagavatula S, Liu G, Park S, Zhou W, Su L, Wain JC, Lynch TJ, Neuberg DS, Christiani DC. XPD and XRCC1 genetic polymorphisms are prognostic factors in advanced non-small-cell lung cancer patients treated with platinum chemotherapy. *J Clin Oncol*. 2004;22:2594–601.
51. Chen B, Zhou Y, Yang P, Wu XT. Polymorphisms of XRCC1 and gastric cancer susceptibility: a meta-analysis. *Mol Biol Rep*. 2012;39:1305–13.
52. Yin J, Vogel U, Ma Y, Qi R, Sun Z, Wang H. The DNA repair gene XRCC1 and genetic susceptibility of lung cancer in a northeastern Chinese population. *Lung Cancer*. 2007;56:153–60.
53. Wojcik KA, Synowiec E, Polakowski P, Blasiak J, Szaflik J, Szaflik JP. Variation in DNA base excision repair genes in fuchs endothelial corneal dystrophy. *Med Sci Monit*. 2015;21:2809–27.
54. Sirisena ND, Samaranyake N, Dissanayake VHW. Electrophoretic mobility shift assays implicate XRCC2:rs3218550C>T as a potential low-penetrant susceptibility allele for sporadic breast cancer. *BMC Res Notes*. 2019;12:476.
55. Kluzniak W, Wokolorczyk D, Rusak B, Huzarski T, Gronwald J, Stempa K, Rudnicka H, Kashyap A, Debniak T, Jakubowska A, et al. Inherited variants in XRCC2 and the risk of breast cancer. *Breast Cancer Res Treat*. 2019;178:657–63.
56. Gok I, Baday M, Cetinkunar S, Kilic K, Bilgin BC. Polymorphisms in DNA repair genes XRCC2 and XRCC3 risk of gastric cancer in Turkey. *Bosn J Basic Med Sci*. 2014;14:214–8.
57. Balkan E, Bilici M, Gundogdu B, Aksungur N, Kara A, Yasar E, Dogan H, Ozturk G. ERCC2 Lys751Gln rs13181 and XRCC2 Arg188His rs3218536 gene polymorphisms contribute to susceptibility of colon, gastric, HCC, lung and prostate cancer. *J BUON*. 2020;25:574–81.
58. Daboussi F, Dumay A, Delacote F, Lopez BS. DNA double-strand break repair signalling: the case of RAD51 post-translational regulation. *Cell Signal*. 2002;14:969–75.
59. Sun H, Qiao Y, Zhang X, Xu L, Jia X, Sun D, Shen C, Liu A, Zhao Y, Jin Y, et al. XRCC3 Thr241Met polymorphism with lung cancer and bladder cancer: a meta-analysis. *Cancer Sci*. 2010;101:1777–82.
60. Santos EM, Santos HBP, de Matos FR, Machado RA, Coletta RD, Galvao HC, Freitas RA. Clinicopathological significance of SNPs in RAD51 and XRCC3 in oral and oropharyngeal carcinomas. *Oral Dis*. 2019;25:54–63.
61. Samara M, Papatthanassiou M, Mitrakas L, Koukoulis G, Vlachostergios PJ, Tzortzis V. DNA repair gene polymorphisms and susceptibility to urothelial carcinoma in a southeastern European population. *Curr Oncol*. 2021;28:1879–85.
62. Rajagopal T, Seshachalam A, Rathnam KK, Talluri S, Venkatabalasubramanian S, Dunna NR. Homologous recombination DNA repair gene RAD51, XRCC2 & XRCC3 polymorphisms and breast cancer risk in South Indian women. *PLoS ONE*. 2022;17:e0259761.
63. Mutlu P, Mutlu M, Yalcin S, Yaylaci A, Unsoy G, Saylam G, Akin I, Gunduz U, Korkmaz H. Association between XRCC3 Thr241Met polymorphism and laryngeal cancer susceptibility in Turkish population. *Eur Arch Otorhinolaryngol*. 2015;272:3779–84.
64. Jin D, Zhang M, Hua H. Impact of polymorphisms in DNA repair genes XPD, hOGG1 and XRCC4 on colorectal cancer risk in a Chinese Han population. *Biosci Rep*. 2019. <https://doi.org/10.1042/BSR20181074>.
65. Garcia JA, Kalacas NA, Sy Ortin T, Ramos MC, Albano PM. XRCC4 c.1394G>T single nucleotide polymorphisms and breast cancer risk among Filipinos. *Asian Pac J Cancer Prev*. 2019;20:1097–101.
66. Liu Y, Zhang H, Zhou K, Chen L, Xu Z, Zhong Y, Liu H, Li R, Shugart YY, Wei Q, et al. Tagging SNPs in non-homologous end-joining pathway genes and risk of glioma. *Carcinogenesis*. 2007;28:1906–13.
67. Hayden PJ, Tewari P, Morris DW, Staines A, Crowley D, Nieters A, Becker N, de Sanjose S, Foretova L, Maynadie M, et al. Variation in DNA repair genes XRCC3, XRCC4, XRCC5 and susceptibility to myeloma. *Hum Mol Genet*. 2007;16:3117–27.
68. Corral R, Lewinger JP, Van Den Berg D, Joshi AD, Yuan JM, Gago-Dominguez M, Cortes VK, Pike MC, Conti DV, Thomas DC, et al. Comprehensive analyses of DNA repair pathways, smoking and bladder cancer risk in Los Angeles and Shanghai. *Int J Cancer*. 2014;135:335–47.
69. Christmann M, Tomicic MT, Roos WP, Kaina B. Mechanisms of human DNA repair: an update. *Toxicology*. 2003;193:3–34.
70. Blankenburg S, Konig IR, Moessner R, Laspe P, Thoms KM, Krueger U, Khan SG, Westphal G, Berking C, Volkenandt M, et al. Assessment of 3 xeroderma pigmentosum group C gene polymorphisms and risk of cutaneous melanoma: a case-control study. *Carcinogenesis*. 2005;26:1085–90.
71. Aref S, El Menshawly N, Abou Zeid T, Gouda E, Abdel Aziz N. DNA repair genes polymorphisms: impact on acute myeloid leukemia patients outcome. *Asian Pac J Cancer Prev*. 2022;23:3577–85.
72. Jamshidi M, Farnoosh G, Mohammadi Pour S, Rafiee F, Saeedi Boroujeni A, Mahmoudian-Sani MR. Genetic variants and risk of thyroid cancer among Iranian patients. *Horm Mol Biol Clin Investig*. 2021;42:223–34.
73. Suhail Y, Cain MP, Vanaja K, Kurywchak PA, Levchenko A, Kalluri R. Kshitz: systems biology of cancer metastasis. *Cell Syst*. 2019;9:109–27.
74. Liu QH, Wang Y, Yong HM, Hou PF, Pan J, Bai J, Zheng JN. XRCC1 serves as a potential prognostic indicator for clear cell renal cell carcinoma and inhibits its invasion and metastasis through suppressing MMP-2 and MMP-9. *Oncotarget*. 2017;8:109382–92.
75. Bhandaru M, Martinka M, Li G, Rotte A. Loss of XRCC1 confers a metastatic phenotype to melanoma cells and is associated with poor survival in patients with melanoma. *Pigment Cell Melanoma Res*. 2014;27:366–75.
76. Bashir N, Sana S, Mahjabeen I, Kayani MA. Association of reduced XRCC2 expression with lymph node metastasis in breast cancer tissues. *Fam Cancer*. 2014;13:611–7.
77. He Y, Penney ME, Negandhi AA, Parfrey PS, Savas S, Yilmaz YE. XRCC3 Thr241Met and TYMS variable number tandem repeat polymorphisms are associated with time-to-metastasis in colorectal cancer. *PLoS ONE*. 2018;13:e0192316.
78. Yang Y, Li X, Hao L, Jiang D, Wu B, He T, Tang Y. The diagnostic value of DNA repair gene in breast cancer metastasis. *Sci Rep*. 2020;10:19626.
79. Leng N, Zhou W, Jiang L, Zhao Y, Zhou M, Sun S, Nie W. Biological functions of miRNA-188-5p/XRCC5 in the metastasis of glioma. *J BUON*. 2021;26:359–65.
80. Liu ZH, Wang N, Wang FQ, Dong Q, Ding J. High expression of XRCC5 is associated with metastasis through Wnt signaling pathway and predicts poor prognosis in patients with hepatocellular carcinoma. *Eur Rev Med Pharmacol Sci*. 2019;23:7835–47.
81. Luo S, Wang W, Feng J, Li R. TEX10 promotes the tumorigenesis and radiotherapy resistance of urinary bladder carcinoma by stabilizing XRCC6. *J Immunol Res*. 2021;2021:5975893.
82. Zhang Y, Zhang Z. The history and advances in cancer immunotherapy: understanding the characteristics of tumor-infiltrating immune cells and their therapeutic implications. *Cell Mol Immunol*. 2020;17:807–21.

83. Pulendran B, Davis MM. The science and medicine of human immunology. *Science*. 2020. <https://doi.org/10.1126/science.aay4014>.
84. Palacios LM, Peyret V, Viano ME, Geysels RC, Chocobar YA, Volpini X, Pellizas CG, Nicola JP, Motran CC, Rodriguez-Galan MC, Fozzatti L. TIM3 expression in anaplastic-thyroid-cancer-infiltrating macrophages: an emerging immunotherapeutic target. *Biology*. 2022;11:1609.
85. Mittal N, Singh S, Mittal R, Kaushal J, Kaushal V. Immune checkpoint inhibitors as neoadjuvant therapy in early triple-negative breast cancer: a systematic review and meta-analysis. *J Cancer Res Ther*. 2022;18:1754–65.
86. Michelson DA, Benoist C, Mathis D. CTLA-4 on thymic epithelial cells complements Aire for T cell central tolerance. *Proc Natl Acad Sci USA*. 2022;119: e2215474119.
87. Maruhashi T, Sugiura D, Okazaki IM, Okazaki T. LAG-3: from molecular functions to clinical applications. *J Immunother Cancer*. 2020. <https://doi.org/10.1136/jitc-2020-001014>.
88. Jiang M, Jia K, Wang L, Li W, Chen B, Liu Y, Wang H, Zhao S, He Y, Zhou C. Alterations of DNA damage response pathway: biomarker and therapeutic strategy for cancer immunotherapy. *Acta Pharm Sin B*. 2021;11:2983–94.
89. Green AR, Aleskandarany MA, Ali R, Hodgson EG, Atabani S, De Souza K, Rakha EA, Ellis IO, Madhusudan S. Clinical impact of tumor DNA repair expression and T-cell infiltration in breast cancers. *Cancer Immunol Res*. 2017;5:292–9.
90. Zhang Y, Zhang X, Jin Z, Chen H, Zhang C, Wang W, Jing J, Pan W. Clinical impact of X-ray repair cross-complementary 1 (XRCC1) and the immune environment in colorectal adenoma-carcinoma pathway progression. *J Inflamm Res*. 2021;14:5403–17.
91. Li F, Zhang Y, Shi Y, Liu S. Comprehensive analysis of prognostic and immune infiltrates for RAD51 in human breast cancer. *Crit Rev Eukaryot Gene Expr*. 2021;31:71–9.
92. Rieke DT, Ochsenreither S, Klinghammer K, Seiwert TY, Klauschen F, Tinhofer I, Keilholz U. Methylation of RAD51B, XRCC3 and other homologous recombination genes is associated with expression of immune checkpoints and an inflammatory signature in squamous cell carcinoma of the head and neck, lung and cervix. *Oncotarget*. 2016;7:75379–93.
93. Barbie DA, Tamayo P, Boehm JS, Kim SY, Moody SE, Dunn IF, Schinzel AC, Sandy P, Meylan E, Scholl C, et al. Systematic RNA interference reveals that oncogenic KRAS-driven cancers require TBK1. *Nature*. 2009;462:108–12.
94. Ahn J, Xia T, Konno H, Konno K, Ruiz P, Barber GN. Inflammation-driven carcinogenesis is mediated through STING. *Nat Commun*. 2014;5:5166.
95. Qi Z, Yan F, Chen D, Xing W, Li Q, Zeng W, Bi B, Xie J. Identification of prognostic biomarkers and correlations with immune infiltrates among cGAS-STING in hepatocellular carcinoma. *Biosci Rep*. 2020. <https://doi.org/10.1042/BSR20202603>.
96. Wang Z, Yan J, Lin H, Hua F, Wang X, Liu H, Lv X, Yu J, Mi S, Wang J, Hu ZW. Toll-like receptor 4 activity protects against hepatocellular tumorigenesis and progression by regulating expression of DNA repair protein Ku70 in mice. *Hepatology*. 2013;57:1869–81.
97. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144:646–74.
98. Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. *Cell Metab*. 2016;23:27–47.
99. Anurag M, Jaehnig EJ, Krug K, Lei JT, Bergstrom EJ, Kim BJ, Vashist TD, Huynh AMT, Dou Y, Gou X, et al. Proteogenomic markers of chemotherapy resistance and response in triple-negative breast cancer. *Cancer Discov*. 2022;12:2586–605.
100. Chen X, Legrand AJ, Cunniffe S, Hume S, Poletto M, Vaz B, Ramadan RS. Prognostic significance of folate metabolism polymorphisms for lung cancer. *Br J Cancer*. 2007;97:247–52.
101. Matakidou A, El Galta R, Rudd MF, Webb EL, Bridle H, Eisen T, Houlston RS. Prognostic significance of folate metabolism polymorphisms for lung cancer. *Br J Cancer*. 2007;97:247–52.
102. Adjei AA, Salavaggione OE, Mandrekar SJ, Dy GK, Ziegler KL, Endo C, Molina JR, Schild SE, Adjei AA. Correlation between polymorphisms of the reduced folate carrier gene (SLC19A1) and survival after pemetrexed-based therapy in non-small cell lung cancer: a North Central cancer treatment group-based exploratory study. *J Thorac Oncol*. 2010;5:1346–53.
103. Perez-Ramirez C, Canadas-Garre M, Alnatsa A, Villar E, Valdivia-Bautista J, Faus-Dader MJ, Calleja-Hernandez MA. Pharmacogenetics of platinum-based chemotherapy: impact of DNA repair and folate metabolism gene polymorphisms on prognosis of non-small cell lung cancer patients. *Pharmacogenom J*. 2019;19:164–77.
104. Damaraju S, Murray D, Dufour J, Carandang D, Myrehaug S, Fallone G, Field C, Greiner R, Hanson J, Cass CE, Parliament M. Association of DNA repair and steroid metabolism gene polymorphisms with clinical late toxicity in patients treated with conformal radiotherapy for prostate cancer. *Clin Cancer Res*. 2006;12:2545–54.
105. Kimmelman AC, White E. Autophagy and tumor metabolism. *Cell Metab*. 2017;25:1037–43.
106. Ma Y, Li W, Chen S, Lin S, Ding S, Zhou X, Liu T, Wang R, Wang W. Characteristics and response to next-generation sequencing-guided therapy in locally advanced or metastatic esophageal cancer. *Int J Cancer*. 2022;152(3):436–46.
107. Demirbag-Sarikaya S, Akkoc Y, Turgut S, Erbil-Bilir S, Kocaturk NM, Dengjel J, Gozuacik D. A novel ATG5 interaction with Ku70 potentiates DNA repair upon genotoxic stress. *Sci Rep*. 2022;12:8134.
108. Shirvani H, Ghanavi J, Aliabadi A, Mousavinasab F, Talebi M, Majidpoor J, Najafi S, Miryounesi SM, Aghaei Zarch SM. miR-211 plays a dual role in cancer development: from tumor suppressor to tumor enhancer. *Cell Signal*. 2023;101: 110504.
109. Faramin Lashkarian M, Hashemipour N, Niaraki N, Soghala S, Moradi A, Sarhangi S, Hatami M, Aghaei-Zarch F, Khosravifar M, Mohamadzadeh A, et al. MicroRNA-122 in human cancers: from mechanistic to clinical perspectives. *Cancer Cell Int*. 2023;23:29.
110. Bahari Khasraghi L, Nouri M, Vazirzadeh M, Hashemipour N, Talebi M, Aghaei Zarch F, Majidpoor J, Kalhor K, Farnia P, Najafi S, Aghaei Zarch SM. MicroRNA-206 in human cancer: mechanistic and clinical perspectives. *Cell Signal*. 2023;101: 110525.
111. Goodall GJ, Wickramasinghe VO. RNA in cancer. *Nat Rev Cancer*. 2021;21:22–36.
112. Anastasiadou E, Jacob LS, Slack FJ. Non-coding RNA networks in cancer. *Nat Rev Cancer*. 2018;18:5–18.
113. Pan J, Huang G, Yin Z, Cai X, Gong E, Li Y, Xu C, Ye Z, Cao Z, Cheng W. Circular RNA FLNA acts as a sponge of miR-486-3p in promoting lung cancer progression via regulating XRCC1 and CYP11A1. *Cancer Gene Ther*. 2022;29:101–21.
114. Li Y, Hu G, Li P, Tang S, Zhang J, Jia G. miR-3940-5p enhances homologous recombination after DSB in Cr(VI) exposed 16HBE cell. *Toxicology*. 2016;344–346:1–6.
115. Zhen N, Yang Q, Zheng K, Han Z, Sun F, Mei W, Yu Y. MiroRNA-127-3p targets XRCC3 to enhance the chemosensitivity of esophageal cancer cells to a novel phenanthroline-dione derivative. *Int J Biochem Cell Biol*. 2016;79:158–67.
116. Zhang Z, Yin J, Lu C, Wei Y, Zeng A, You Y. Exosomal transfer of long non-coding RNA SBF2-AS1 enhances chemoresistance to temozolomide in glioblastoma. *J Exp Clin Cancer Res*. 2019;38:166.
117. Unfried JP, Marin-Baquero M, Rivera-Calzada A, Razquin N, Martin-Cuevas EM, de Braganca S, Aicart-Ramos C, McCoy C, Prats-Mari L, Arribas-Bosacoma R, et al. Long noncoding RNA NIHCOLE promotes ligation efficiency of DNA double-strand breaks in hepatocellular carcinoma. *Cancer Res*. 2021;81:4910–25.
118. Chen P, Nie ZY, Liu XF, Zhou M, Liu XX, Wang B. CircXRCC5, as a potential novel biomarker, promotes glioma progression via the miR-490-3p/XRCC5/CLC3 competing endogenous RNA network. *Neuroscience*. 2022;494:104–18.
119. Li Q, Liu J, Jia Y, Li T, Zhang M. miR-623 suppresses cell proliferation, migration and invasion through direct inhibition of XRCC5 in breast cancer. *Aging*. 2020;12:10246–58.
120. Dang Y, Wang X, Hao Y, Zhang X, Zhao S, Ma J, Qin Y, Chen ZJ. MicroRNA-379-5p is associate with biochemical premature ovarian insufficiency through PARP1 and XRCC6. *Cell Death Dis*. 2018;9:106.
121. Hildebrandt MA, Gu J, Wu X. Pharmacogenomics of platinum-based chemotherapy in NSCLC. *Expert Opin Drug Metab Toxicol*. 2009;5:745–55.

122. Kiyohara C, Takayama K, Nakanishi Y. Association of genetic polymorphisms in the base excision repair pathway with lung cancer risk: a meta-analysis. *Lung Cancer*. 2006;54:267–83.
123. Jorgensen TJ. Enhancing radiosensitivity: targeting the DNA repair pathways. *Cancer Biol Ther*. 2009;8:665–70.
124. Rosell R, Taron M, Barnadas A, Scagliotti G, Sarries C, Roig B. Nucleotide excision repair pathways involved in Cisplatin resistance in non-small-cell lung cancer. *Cancer Control*. 2003;10:297–305.
125. Abdel-Fatah T, Sultana R, Abbotts R, Hawkes C, Seedhouse C, Chan S, Madhusudan S. Clinicopathological and functional significance of XRCC1 expression in ovarian cancer. *Int J Cancer*. 2013;132:2778–86.
126. Xu W, Zhou B, Zhao X, Zhu L, Xu J, Jiang Z, Chen D, Wei Q, Han M, Feng L, et al. KDM5B demethylates H3K4 to recruit XRCC1 and promote chemoresistance. *Int J Biol Sci*. 2018;14:1122–32.
127. Xu W, Wang S, Chen Q, Zhang Y, Ni P, Wu X, Zhang J, Qiang F, Li A, Roe OD, et al. TXNL1-XRCC1 pathway regulates cisplatin-induced cell death and contributes to resistance in human gastric cancer. *Cell Death Dis*. 2014;5:e1055.
128. Shan J, Wang X, Zhao J. XRCC2 reduced the sensitivity of NSCLC to radiochemotherapy by arresting the cell cycle. *Am J Transl Res*. 2022;14:3783–95.
129. He Y, Xue B, Xiong X, Wu W, Li X, Zhao H. Correlation analysis between XRCC2 polymorphism and radiosensitivity of non-small cell lung cancer. *Panminerva Med*. 2021. <https://doi.org/10.23736/S0031-0808.21.04472-4>.
130. Zheng Z, Ng WL, Zhang X, Olson JJ, Hao C, Curran WJ, Wang Y. RNAi-mediated targeting of noncoding and coding sequences in DNA repair gene messages efficiently radiosensitizes human tumor cells. *Cancer Res*. 2012;72:1221–8.
131. Wang Q, Wang Y, Du L, Xu C, Sun Y, Yang B, Sun Z, Fu Y, Cai L, Fan S, et al. shRNA-mediated XRCC2 gene knockdown efficiently sensitizes colon tumor cells to X-ray irradiation in vitro and in vivo. *Int J Mol Sci*. 2014;15:2157–71.
132. Xu ZY, Loignon M, Han FY, Panasci L, Alojz R. Xrcc3 induces cisplatin resistance by stimulation of Rad51-related recombinational repair, S-phase checkpoint activation, and reduced apoptosis. *J Pharmacol Exp Ther*. 2005;314:495–505.
133. Xu Z, Chen ZP, Malapetsa A, Alaoui-Jamali M, Bergeron J, Monks A, Myers TG, Mohr G, Sausville EA, Scudiero DA, et al. DNA repair protein levels vis-a-vis anticancer drug resistance in the human tumor cell lines of the National Cancer Institute drug screening program. *Anticancer Drugs*. 2002;13:511–9.
134. Cheng J, Liu W, Zeng X, Zhang B, Guo Y, Qiu M, Jiang C, Wang H, Wu Z, Meng M, et al. XRCC3 is a promising target to improve the radiotherapy effect of esophageal squamous cell carcinoma. *Cancer Sci*. 2015;106:1678–86.
135. He H, Lee C, Kim JK. UHRF1 depletion sensitizes retinoblastoma cells to chemotherapeutic drugs via downregulation of XRCC4. *Cell Death Dis*. 2018;9:164.
136. Hori M, Someya M, Matsumoto Y, Nakata K, Kitagawa M, Hasegawa T, Tsuchiya T, Fukushima Y, Gocho T, Sato Y, et al. Influence of XRCC4 expression in esophageal cancer cells on the response to radiotherapy. *Med Mol Morphol*. 2017;50:25–33.
137. Lee IN, Yang JT, Huang C, Huang HC, Wu YP, Chen JC. Elevated XRCC5 expression level can promote temozolomide resistance and predict poor prognosis in glioblastoma. *Oncol Lett*. 2021;21:443.
138. Chen D, Chou FJ, Chen Y, Huang CP, Tian H, Wang Y, Niu Y, You B, Yeh S, Xing N, Chang C. Targeting the radiation-induced ARV7-mediated circNHS/miR-512-5p/XRCC5 signaling with Quercetin increases prostate cancer radiosensitivity. *J Exp Clin Cancer Res*. 2022;41:235.
139. Mei PJ, Bai J, Miao FA, Li ZL, Chen C, Zheng JN, Fan YC. Relationship between expression of XRCC1 and tumor proliferation, migration, invasion, and angiogenesis in glioma. *Invest New Drugs*. 2019;37:646–57.
140. Zheng Y, Zhang H, Guo Y, Chen Y, Chen H, Liu Y. X-ray repair cross-complementing protein 1 (XRCC1) loss promotes beta-lapachone-induced apoptosis in pancreatic cancer cells. *BMC Cancer*. 2021;21:1234.
141. Xu K, Song X, Chen Z, Qin C, He Y, Zhan W. XRCC2 promotes colorectal cancer cell growth, regulates cell cycle progression, and apoptosis. *Medicine*. 2014;93:e294.
142. Yan CT, Kaushal D, Murphy M, Zhang Y, Datta A, Chen C, Monroe B, Mostoslavsky G, Coakley K, Gao Y, et al. XRCC4 suppresses medulloblastomas with recurrent translocations in p53-deficient mice. *Proc Natl Acad Sci USA*. 2006;103:7378–83.
143. Zhang Z, Zheng F, Yu Z, Hao J, Chen M, Yu W, Guo W, Chen Y, Huang W, Duan Z, Deng W. XRCC5 cooperates with p300 to promote cyclooxygenase-2 expression and tumor growth in colon cancers. *PLoS ONE*. 2017;12:e0186900.
144. Kim JH, Park SY, Jeon SE, Choi JH, Lee CJ, Jang TY, Yun HJ, Lee Y, Kim P, Cho SH, et al. DCLK1 promotes colorectal cancer stemness and aggressiveness via the XRCC5/COX2 axis. *Theranostics*. 2022;12:5258–71.
145. Zhu B, Cheng D, Li S, Zhou S, Yang Q. High expression of XRCC6 promotes human osteosarcoma cell proliferation through the beta-Catenin/Wnt signaling pathway and is associated with poor prognosis. *Int J Mol Sci*. 2016;17:1188.
146. Tang B, Zhang Y, Wang W, Qi G, Shimamoto F. PARP6 suppresses the proliferation and metastasis of hepatocellular carcinoma by degrading XRCC6 to regulate the Wnt/beta-catenin pathway. *Am J Cancer Res*. 2020;10:2100–13.
147. Naguib M, Helwa MM, Soliman MM, Abdel-Samiee M, Eljaky AM, Hammam O, Zaghla H, Abdelsameea E. XRCC1 gene polymorphism increases the risk of hepatocellular carcinoma in Egyptian population. *Asian Pac J Cancer Prev*. 2020;21:1031–7.
148. Sobiahe A, Hijazi E, Al-Ameer HJ, Almasri Y, Jarrar Y, Zihlif M, Shomaf M, Al-Rawashdeh B. Arg399Gln XRCC1 polymorphism and risk of squamous cell carcinoma of the head and neck in Jordanian patients. *Asian Pac J Cancer Prev*. 2020;21:663–5.
149. Wang F, Zhao Q, He HR, Zhai YJ, Lu J, Hu HB, Zhou JS, Yang YH, Li YJ. The association between XRCC1 Arg399Gln polymorphism and risk of leukemia in different populations: a meta-analysis of case-control studies. *Oncotargets Ther*. 2015;8:3277–87.
150. Zhang L, Zhao J, Yu B, Song X, Sun G, Han L, Wang L, Dong S. Correlations between microsatellite instability, ERCC1/XRCC1 polymorphism and clinical characteristics, and FOLFOX adjuvant chemotherapy effect of colorectal cancer patients. *Cancer Genet*. 2017;218–219:51–7.
151. Lin WY, Camp NJ, Cannon-Albright LA, Allen-Brady K, Balasubramanian S, Reed MW, Hopper JL, Apicella C, Giles GG, Southey MC, et al. A role for XRCC2 gene polymorphisms in breast cancer risk and survival. *J Med Genet*. 2011;48:477–84.
152. Michalska MM, Samulak D, Bienkiewicz J, Romanowicz H, Smolarz B. Association between -41657C/T single nucleotide polymorphism of DNA repair gene XRCC2 and endometrial cancer risk in Polish women. *Pol J Pathol*. 2015;66:67–71.
153. Pasqualetti F, Gonnelli A, Orlandi P, Palladino E, Giannini N, Gadducci G, Mattioni R, Montrone S, Calistri E, Mazzanti CM, et al. Association of XRCC3 rs1799794 polymorphism with survival of glioblastoma multiforme patients treated with combined radio-chemotherapy. *Invest New Drugs*. 2021;39:1159–65.
154. Nowacka-Zawisza M, Raszkievicz A, Kwasiborski T, Forma E, Brys M, Rozanski W, Krajewska WM. RAD51 and XRCC3 polymorphisms are associated with increased risk of prostate cancer. *J Oncol*. 2019;2019:2976373.
155. Gresner P, Gromadzinska J, Polanska K, Twardowska E, Jurewicz J, Wasowicz W. Genetic variability of Xrcc3 and Rad51 modulates the risk of head and neck cancer. *Gene*. 2012;504:166–74.
156. Singh PK, Mistry KN, Chiramana H, Rank DN, Joshi CG. Exploring the deleterious SNPs in XRCC4 gene using computational approach and studying their association with breast cancer in the population of West India. *Gene*. 2018;655:13–9.
157. Makkoch J, Praianantathavorn K, Sopipong W, Chuaypen N, Tangkijvanich P, Payungporn S. Genetic variations in XRCC4 (rs1805377) and ATF6 (rs2070150) are not associated with hepatocellular carcinoma in Thai patients with hepatitis B virus infection. *Asian Pac J Cancer Prev*. 2016;17:591–5.
158. Hasan SK, Buttari F, Ottone T, Voso MT, Hohaus S, Marasco E, Mantovani V, Garagnani P, Sanz MA, Cicconi L, et al. Risk of acute promyelocytic leukemia in multiple sclerosis: coding variants of DNA repair genes. *Neurology*. 2011;76:1059–65.
159. Willems P, De Ruyck K, Van den Broecke R, Makar A, Perletti G, Thierens H, Vral A. A polymorphism in the promoter region of Ku70/XRCC6, associated with breast cancer risk and oestrogen exposure. *J Cancer Res Clin Oncol*. 2009;135:1159–68.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.