



The interactions between habitat, sex, biomass and leaf traits of different willow (*Salix*) genotypes

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Abstract

Knowledge of the impacts of sex on plant mortality and biomass production has scientific and practical importance. In the case of willows, we know relatively little about such effects. The main objective of this study was to evaluate whether the sex of individuals of different willow species determines their biomass and mortality. An additional goal was to determine whether the secondary sex characteristics, such as leaf traits, depend on sex. The experiment was conducted from 2011 to 2014 with 8100 plants comprising 150 willow genotypes, including 8 species, 16 interspecies hybrids, cultivars, and specimens differentiated by sex. Statistical analysis of the leaf traits revealed their relationship to sex. On average, male specimens have longer and wider leaves. They also have longer petioles. Males of the studied *Salix* genotypes were characterized by higher biomass and showed a greater survival rate than females but only under better site conditions; when the site conditions were poorer, males had higher mortality than females.

Article Highlights

- Effective rock phosphate (RP) solubilization by *Bacillus thuringensis* P₀B₁₁, *Lysinobaccillus fusiformis* P₀B₂₈, and *Aspergillus aculeatus* P₀F₃ isolated from Mahanadi estuary of Odisha, India.
- First report of RP solubilization by *L. fusiformis* P₀B₂₈. First report of in vitro stearic acid production during RP solubilization.
- Organic acids were the major mechanism for RP solubilization and responsible for morphological and mineralogical changes confirmed by SEM, XRD and FTIR, respectively.

Keywords Biomass · Mortality · *Salix* · Sexual dimorphism · Willows

Introduction

Willows (genus *Salix*) have a very significant utility and play an important environmental role. Their importance in medicine (e.g., Uehleke et al. 2013; Desborough and Keeling 2017), as a source of renewable energy (Lipiński and Żejmo 2012; Bakšienė and Titova 2018), in the basketry and furniture industries (Bhat et al. 2017), phytoremediation (Goliński et al. 2015; Mleczek et al. 2017, 2018) and many other fields is well known. *Salix* species are components of such important Natura 2000 habitats as 2170 (dunes with *Salix repens* ssp. *argentea*), 3210 (Fennoscandian natural rivers), 4080 (subarctic *Salix* spp. scrub), 6450 (northern boreal alluvial meadows), 91E0 (alluvial

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forests), 92A0 (*Salix alba* and *Populus alba* galleries) and 92B0 (riparian formations on intermittent Mediterranean water courses) (EC 2013). Willows are also a source of pollen and nectar for pollinating insects (Ostaff et al. 2015). However, despite such a wide field of interest in numerous studies devoted to willows, the fact that willows are dioecious and that the results of investigations may depend on the sex of the tested specimens is often ignored (Hytönen 1995; Larsson 1998; Labrecque and Teodorescu 2003; Nordh and Verwijst 2004; Smart et al. 2005).

It is well-known—almost a truism—that differences between sexes are common in nature and determine primary and secondary sex characters (Barrett and Hough 2013). Sexual dimorphism (SD) is observed in many organisms, such as humans, cervids (Geist and Bayer 2009), birds (Owens and Hartley 1998), spiders (Foellmer and Moya-Laraño 2007; Inkpen and Foellmer 2010) and other animals, while SD in plants is much less widely appreciated (Geber et al. 1999). Meanwhile, focusing attention on the sex of plant specimens and the effects of the differences between the sexes may have significant ecological and practical significance. This has been demonstrated, among others, on an example of *Cannabis sativa* and *Spinacia oleracea* (Lloyd and Webb 1977), *Nyssa aquatica* (Shea et al. 1993), *Phoenix dactylifera*, *Silene latifolia*, *Zea mays* and *Ceratopteris richardii* (Juarez and Banks 1998), *Corema album* (Álvarez-Cansino et al. 2010), *Mercurialis annua* (Hesse and Pannell, 2011), *Phoenix dactylifera* (Intha and Chairprasart, 2018) and *Ambrosia artemisiifolia* (Nakahara et al. 2018), but there are few such works on willows (Mirski 2014; Yang et al. 2020).

Current knowledge allows us to conclude that males tend to be larger than females in relatively large species, while females tend to be the larger in relatively small species (Abouheif and Fairbairn, 1997; Fairbairn 1997; Colwell 2000; Obeso 2002), this is known as Rensch's rule (Rensch, 1960). It is also considered that dioecious male individuals usually produce higher biomass than those that are not dioecious (Galambosi et al. 2009), but biomass production may depend on site conditions. As Tognetti (2012) summarized, according to Dawson and Bliss (1989), females are more common in high-resource and males in low-resource microsites. Similar observations were noted by Bierzychudek and Eckhart (1988), Mercer and Eppley (2010) and Barrett and Hough (2013).

Science has attempted to explain why females and males display different advantages under different site conditions but there remain many unanswered questions. Therefore, the main objective of this study was to test whether the sex of specimens of different willow species affected their biomass and mortality. An additional goal

was to determine whether secondary sex characteristics, such as leaf traits, depend on the sex of an individual.

Materials and methods

The described project is based on the Poznań University of Life Sciences Willow Collection, which is located in different parts of Poland. The collection dates back to the 1960s and now comprises 150 genotypes. Their taxonomy was verified by Professor Jerzy Zieliński of the Institute of Dendrology, Polish Academy of Sciences.

Study area

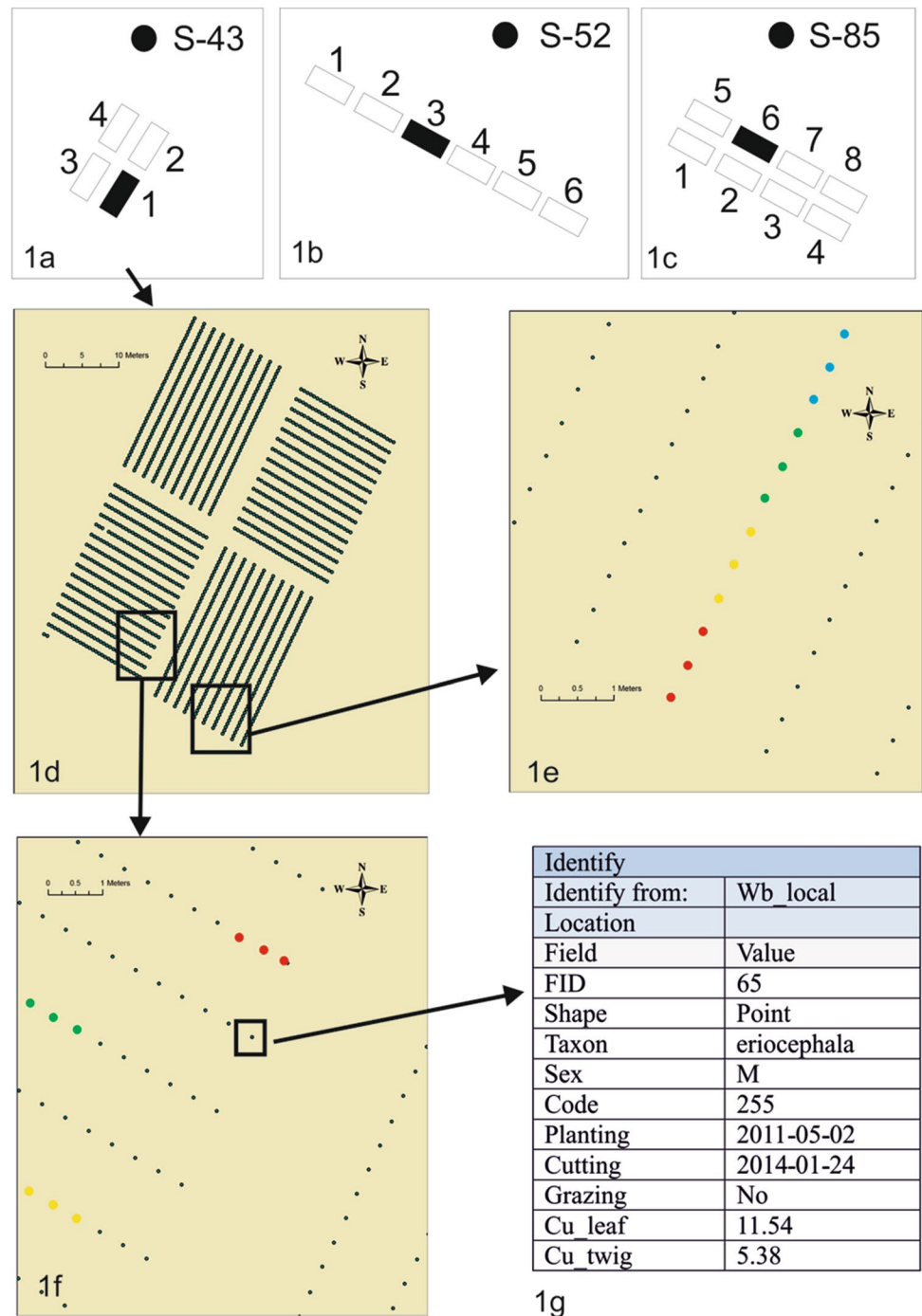
All genotypes of willows included in the Willow Collection were randomly planted in 2011, with replication across 34 experimental plots at 6 locations. The research areas described in this paper comprise three locations named: S-43, S-52 and S-85. The coordinates of the centers of these areas are S-43—51° 13' 4" N; 18°02' 19" E (Fig. 1a), S-52—51°13' 9" N; 18°00'12" E (Fig. 1b) and S-85—51°11' 33" N; 18°04' 47" E (Fig. 1c).

Research area S-43 comprises four experimental plots (Fig. 1a), S-52—six plots (Fig. 1b), and S-85—eight plots (Fig. 1c). Figure 1d shows an enlargement of the S-43 research area. In plots No. 1 and No. 4, the rows were oriented to the NE–SW direction. At plots No. 2 and No. 3, they were oriented to the NW–SE direction. Figure 1e, f show the enlargement of the rows at plots Nos. 1 and 3, where all plants were planted in rows at a distance of 0.5 m between cuttings and 1.5 m between rows (as in all the other plots). The distribution of genotypes in each plot is random. In each plot, each taxon is represented by three plants (Fig. 1e, f; red, yellow, green and blue dots). Each plant is mapped and the location and features of each plant were noted in the ArcGIS program (Fig. 1g).

The maximal distance between these three areas is 6.2 km, hence macroclimatic conditions are similar [for Wieluń meteorological station (51° 12' 39" N; 18° 33' 25" E; altitude: 201 m a.s.l.), for the period 1987–2014 the average annual temperature was 8.8 °C and the average annual precipitation was 564 mm (public data)]. Research areas S-43, S-52 and S-85 are flat so the relief of all locations is identical. Therefore, climatic and relief interactions were excluded from the sex—biomass and sex—mortality analyses.

In 2010, the year before willow planting, a basic soil description was performed which included soil type, its granulometric composition [determined using sedimentation methods (PN-R-04032 1998; PN-R-04033 1998), pH (PN-ISO 10390. 1997) and the water regime. The soil features, such as soil type, type of sediment and water availability were described on the basis of the pit soil,

Fig. 1 a–c Experimental design of plots at the research areas: S-43 (a), S-52 (b), S-85 (c); the numbers on Figs. 1a–c mean the number of replicated experimental plots in given location; **d** the enlargement of the S-43 research area with visible rows of planted willows; **e** and **f** the enlargement of the rows at plots; red, yellow, green and blue dots show the distribution of genotypes in each row (3 plants by each taxon); **g** an example of a description of each plant made in the ArcGIS program



dug by hand in the middle of each research area to a depth of 1.5 m. At two of the three locations -at depths of 0.95 and 1.50 m—the groundwater table was found (S-85 and S-43, respectively). Additionally, in 2011 soil samples were collected with a soil sampling tube from a depth of 0.0–0.25 m. The samples were then placed in polypropylene containers and transported to the laboratory, where the total content of organic carbon according to the Polish

standard (PN-ISO 14235 2003) and nitrogen according to the Kjeldahl method (PN-ISO 11261 2002) were analyzed.

Characteristics of plant material and experimental design

The Poznań University of Life Sciences Willow Collection comprises 150 genotypes, including species, natural

Table 1 Summary of shoot mass in all tested genotypes and the results of Tukey's test

No	Name of taxon	Sex code	Location			Average	Tukey's test
			S 43	S 52	S 85		
Mass (kg)							
1	<i>Salix purpurea</i> L. × <i>triandra</i> L. × <i>viminalis</i> L. '15'	F	14.5	12	8	11.5	a
2	<i>S. purpurea</i> × <i>triandra</i> . × <i>viminalis</i> '46'	F	5	9.5	12.5	9	ab
3	<i>S. × smithiana</i> Willd. '5'	M	10.5	3	11.5	8.33	abc
4	<i>S. × smithiana</i> '1'	F	8	8	8	8	abcd
5	<i>S. × smithiana</i> 'Semipalatinskiensis '89'	M	5.5	6	12	7.83	abcd
6	<i>S. × blanda</i> Andersson '247'	F	3.3	8.5	11	7.6	abcde
7	<i>S. viminalis</i> × <i>S. sp.</i> '8'	M	2.7	7.9	11.5	7.37	abcde
8	<i>S. × sepulcralis</i> Simonk. '24'	B	3	3.8	13.5	6.77	abcde
9	<i>S. × sepulcralis</i> '4'	B	4.2	5	9.5	6.23	abcde
10	<i>S. purpurea</i> '70'	M	3.5	3.2	12	6.23	abcde
11	<i>S. purpurea</i> 'Angustifolia 68'	M	5	7	6.5	6.17	abcde
12	<i>S. purpurea</i> 'Angustifolia 72'	M	4.5	3.5	10	6	abcde
13	<i>S. × sepulcralis</i> '17'	M	2.4	4.3	10.5	5.73	abcde
14	<i>S. cinerea</i> L. × <i>S. sp.</i> '205/185'	F	4.9	3.9	8	5.6	abcde
15	<i>S. eriocephala</i> Michx. '51'	F	5.5	3.8	7.5	5.6	abcde
16	<i>S. purpurea</i> '168/155'	F	3.5	2.1	11	5.53	abcde
17	<i>S. purpurea</i> '7'	F	3.5	4	8.8	5.43	abcde
18	<i>S. fragilis</i> '22'	0	4	2.9	9	5.3	abcde
19	<i>S. purpurea</i> '87'	F	3.8	3.2	8.5	5.17	abcde
20	<i>S. purpurea</i> '151/138'	M	5	3.8	6.5	5.1	abcde
21	<i>S. purpurea</i> '126'	F	2.5	4	8.5	5	abcde
22	<i>S. × pontederana</i> Schleich. '196/187'	F	2.5	3.5	8.5	4.83	abcde
23	<i>S. × rubra</i> Huds. '101'	F	3.4	2.5	8.5	4.8	abcde
24	<i>S. triandra</i> '122'	F	3.5	7.1	3.5	4.7	abcde
25	<i>S. purpurea</i> '69'	F	1.2	2.9	10	4.7	abcde
26	<i>S. eriocephala</i> '237'	F	9	2.5	2.5	4.67	abcde
27	<i>S. purpurea</i> '74'	M	4	3.2	6.8	4.67	abcde
28	<i>S. purpurea</i> '63'	F	1	3.1	9.5	4.53	bcde
29	<i>S. purpurea</i> '66'	M	2.5	3.4	7.6	4.5	bcde
30	<i>S. purpurea</i> 'Helix Pyramidalis 160.147'	M	3	3.4	7	4.47	bcde
31	<i>S. viminalis</i> '65/51'	M	4.2	4	4.8	4.33	bcde
32	<i>S. alba</i> L. × <i>S. triandra</i> '206'	F	1.5	2.5	9	4.33	bcde
33	<i>S. × rubra</i> Huds. '49'	F	2.2	3.2	7.5	4.3	bcde
34	<i>S. purpurea</i> '75'	F	1.7	3.2	8	4.3	bcde
35	<i>S. purpurea</i> '79'	F	1.5	4.5	6.4	4.13	bcde
36	<i>S. fragilis</i> '9'	F	2.3	2	8	4.1	bcde
37	<i>S. purpurea</i> '136/123'	M	2.4	3.9	5.8	4.03	bcde
38	<i>S. × rubra</i> '102'	F	2.2	2.2	7.4	3.93	bcde
39	<i>S. triandra</i> × <i>alba</i> '36'	F	5.5	4.8	1.5	3.93	bcde
40	<i>S. × rubra</i> '94'	F	3.8	3	5	3.93	bcde
41	<i>S. purpurea</i> '84'	M	3.8	3.5	4.5	3.93	bcde
42	<i>S. eriocephala</i> '253'	F	3.5	3.4	4.8	3.9	bcde
43	<i>S. myrsinifolia</i> Salisb. '52'	F	2	4.8	4.5	3.77	bcde
44	<i>S. purpurea</i> '67'	M	3	2.2	6	3.73	bcde
45	<i>S. × rubra</i>	F	0.5	4.1	6.5	3.7	bcde
46	<i>S. fragilis</i> '95'	M	6	1.5	3.5	3.67	bcde
47	<i>S. triandra</i> '33'	F	5	0.9	5	3.63	bcde

Table 1 (continued)

No	Name of taxon	Sex code	Location			Average	Tukey's test
			S 43	S 52	S 85		
Mass (kg)							
48	<i>S. × rubra</i> '26/18'	F	2	4.4	4.5	3.63	bcde
49	<i>S. purpurea</i> '61'	F	1.6	2.9	6.4	3.63	bcde
50	<i>S. purpurea</i> '81'	F	2.8	2.4	5.7	3.63	bcde
51	<i>S. cinerea</i> × <i>S.</i> sp. '121'	0	5.5	1.8	3.5	3.6	bcde
52	<i>S. × rubra</i> '28'	M	2	2	6.8	3.6	bcde
53	<i>S. purpurea</i> '78'	M	0.7	2.6	7.5	3.6	bcde
54	<i>S. fragilis</i> '214'	F	4	1	5.4	3.47	bcde
55	<i>S. fragilis</i> '199'	0	3.7	1.4	5.3	3.47	bcde
56	<i>S. viminalis</i> '99'	M	0.9	1.5	7.8	3.4	bcde
57	<i>S. × rubra</i> '47'	F	4.2	2.7	3.2	3.37	bcde
58	<i>S. × sepulcralis</i> '11'	B	0.5	4	5.5	3.33	bcde
59	<i>S. × rubra</i> '30'	M	1	2.2	6.8	3.33	bcde
60	<i>S. × rubra</i> '64'	F	1.2	1.1	7.5	3.27	bcde
61	<i>S. purpurea</i> '82'	F	3.5	1.8	4.5	3.27	bcde
62	<i>S. × smithiana</i> '207/191'	M	3.4	0.8	5.3	3.17	bcde
63	<i>S. triandra</i> '205'	F	1	1	7.4	3.13	bcde
64	<i>S. fragilis</i> × <i>triandra</i> '20'	M	1.5	2.4	5.5	3.13	bcde
65	<i>S. × pontederana</i> '56'	F	2.4	3.7	3.2	3.1	bcde
66	<i>S. eriocephala</i> '209'	F	3.2	2.5	3.5	3.07	bcde
67	<i>S. purpurea</i> '149/136'	F	2	2	5	3	bcde
68	<i>S. eriocephala</i> '255'	M	2	4	3	3	bcde
69	<i>S. fragilis</i> '239'	M	3.8	2.4	2.8	3	bcde
70	<i>S. purpurea</i> 'Scharfenbergiensis 76'	F	1	2.7	5	2.9	bcde
71	<i>S. purpurea</i> '57'	F	1.5	3.5	3.7	2.9	bcde
72	<i>S. purpurea</i> '156/143'	F	1	1	6.5	2.83	bcde
73	<i>S. × smithiana</i> '38'	M	1.4	2.5	4.5	2.8	bcde
74	<i>S. purpurea</i> '23'	F	1.9	2	4.2	2.7	bcde
75	<i>S. purpurea</i> '153/140'	F	1.5	3.2	3	2.57	bcde
76	<i>S. fragilis</i> '257'	0	2.4	1.8	3.5	2.57	bcde
77	<i>S. × rubra</i> '93'	F	0.5	0.1	7	2.53	bcde
78	<i>S. × rubra</i> '43'	M	0.5	1	6	2.5	bcde
79	<i>S. eriocephala</i> '241'	F	3	0.3	4	2.43	bcde
80	<i>S. purpurea</i> '73'	F	0.5	0.5	6	2.33	bcde
81	<i>S. purpurea</i> '125'	M	0.5	2	3.5	2	cde
82	<i>S. purpurea</i> '16'	F	1	1.4	3.5	1.97	cde
83	<i>S. × laurina</i> Sm. '220/205'	0	1	2.2	2.5	1.9	cde
84	<i>S. eriocephala</i> '202'	F	2	0.9	2.5	1.8	cde
85	<i>S. triandra</i> '235'	F	1	0.3	4	1.77	cde
86	<i>S. purpurea</i> '58'	F	1.3	2	2	1.77	cde
87	<i>S. myrsinifolia</i> 'Prunifolia 53'	F	3	1.8	0.2	1.67	cde
88	<i>S. purpurea</i> '155/142'	0	2	0.7	2.2	1.63	cde
89	<i>S. purpurea</i> '39/26'	F	0.5	2.1	2.2	1.6	cde
90	<i>S. triandra</i> '34'	F	1.4	1	2	1.47	de
91	<i>S. purpurea</i> '77'	F	0.2	2.5	1.5	1.4	de
92	<i>S. eriocephala</i> 'Nicholsoni Purpurescens 204'	F	2	0.4	1.6	1.33	de
93	<i>S. eriocephala</i> × <i>S. cordata</i> Michx. '210'	F	1.2	0.2	2.5	1.3	de
94	<i>S. alba</i> 'Chermesina 301/301'	M	0.5	0.9	1.5	0.97	e

Table 1 (continued)

No	Name of taxon	Sex code	Location			Average	Tukey's test
			S 43	S 52	S 85		
Mass (kg)							
95	<i>S. purpurea</i> '19/15'	M	0	2.5	13	5.17	–
96	<i>S. × smithiana</i> '44'	M	0	7.2	8	5.07	–
97	<i>S. × sepulcralis</i> '162/143'	B	6.5	6.8	0	4.43	–
98	<i>S. purpurea</i> '71'	F	0	3.9	9	4.3	–
99	<i>S. fragilis</i> '25'	0	0	3	8	3.67	–
100	<i>S. purpurea</i> '26'	M	1	0	9	3.33	–
101	<i>S. × rubra</i> '66/52'	F	2.8	0	6.5	3.1	–
102	<i>S. purpurea</i> 'Angustifolia 159/146'	M	4.5	0	3	2.5	–
103	<i>S. × sepulcralis</i> '40'	B	7	0	0	2.33	–
104	<i>S. purpurea</i> '65'	F	0	1.5	5.5	2.33	–
105	<i>S. × rubra</i> '223'	F	0	0	6.8	2.27	–
106	<i>S. × smithiana</i> '221/206'	M	6	0	0.2	2.07	–
107	<i>S. × laurina</i> '216'	F	2.5	3.5	0	2	–
108	<i>S. purpurea</i> '158/145'	F	0.5	0	5.5	2	–
109	<i>S. purpurea</i> '167/154'	F	0	1.2	4.5	1.9	–
110	<i>S. fragilis</i> '45'	B	0	1.8	3.7	1.83	–
111	<i>S. purpurea</i> '21'	F	0.8	0	4.5	1.77	–
112	<i>S. × rubra</i> '96'	F	0	2.1	3	1.7	–
113	<i>S. purpurea</i> '83'	B	2.2	2.9	0	1.7	–
114	<i>S. cinerea</i> × <i>S. triandra</i> '186/171'	F	5	0	0	1.67	–
115	<i>S. fragilis</i> '197/182'	0	1.2	0	3.8	1.67	–
116	<i>S. purpurea</i> '86'	M	0	0	5	1.67	–
117	<i>S. alba</i> '225'	0	0	0	4.8	1.6	–
118	<i>S. fragilis</i> 'Monspeliensis '46/32'	0	3	1.2	0	1.4	–
119	<i>S. × rubra</i> '29'	M	0	0.9	2.8	1.23	–
120	<i>S. eriocephala</i> 'Nicholsoni Purpurescens 150/137'	F	2.5	1.2	0	1.23	–
121	<i>S. reichardtii</i> Kern. '50'	F	1	2.1	0	1.03	–
122	<i>S. fragilis</i> '249'	0	1	2	0	1	–
123	<i>S. acutifolia</i> Willd. '124'	F	0	1.9	1	0.97	–
124	<i>S. alba</i> × <i>triandra</i> '234'	0	2	0	0	0.67	–
125	<i>S. purpurea</i> '60'	F	2	0	0	0.67	–
126	<i>S. fragilis</i> '231'	0	0.8	0	0	0.27	–
127	<i>S. × rubens</i> Schrank. '178/164'	F	0.6	0	0	0.2	–

A value of "0" in the "Mass" columns means that the plant did not survive after 3 years of cultivation. In the "Sex code" column, 0 denotes genotypes for which the sex was not determined, F indicates female, M indicates male, and B indicates bisexual; the values in columns are the averages for replications from each location

hybrids and cultivars differentiated by sex and varieties. The list of genotypes is shown in Table 1. In accordance with the design (Fig. 1) the research areas described in this paper included a total of 18 plots (4, 6 and 8, respectively) in which 8100 plants were planted (3 cuttings of each genotype × 150 genotypes × 18 plots). All cuttings were of a similar length (29–30 cm) and thickness (0.7–10 mm). Before planting the soil was ploughed and harrowed. Next, soaked cuttings were planted at a depth of about 25 cm (5 cm being left above the ground). The planting was carried out by hand.

All plants growing on the indicated plots (Fig. 1a–c—black rectangles) were cut in January 2014 and further analyzed.

Sex determination

Sex was determined in the third vegetation season (the year 2013) directly at experimental plots. Willows are generally dioecious, but among the tested genotypes there were willows with male and female catkins on the same shrub or willows with male and female features in one catkin (mainly

Salix × *sepulcralis* and single genotypes of *S. fragilis* and *S. purpurea*). There were also some plants without any catkins (mainly *S. fragilis*), and in such cases it was impossible to determine the sex. Therefore, all genotypes were divided into four categories: 0—with indeterminate sex, 1—females, 2—males, and 3—willows with separate male and female catkins on the same shrub or willows with male and female features in one catkin.

Biomass measurements

The measurements of biomass were performed in January 2014 after three full vegetation seasons (years: 2011, 2012, 2013), after growth cessation and leaf fall, when the shoots and root age was 3 years. All living shoots of each plant were cut at 5 cm above the shoot base, and the plant fresh weight was determined by weighing all shoots of each taxon in the field using a field hook balance with an accuracy of 10 g. Fresh weight instead of dry weight was measured as the cut shoots were to be used for the next experiment and had to be live.

Leaf feature measurements

Leaf features were measured with the digiShape computer program. For each taxon in July 2013, 100 leaves were picked from the outer part of the crown, at half the height of the shrub, 25 from the N, S, W and E directions. The leaves were then scanned and prepared for measurement. 50 leaves from each taxon were scanned from the upper side of the leaf blade and 50 from the lower side (Figs. 2, 3). Thus, length, width, elongation, area of leaf blade and length of petiole were measured.

Statistical analysis

The following correlations were evaluated: habitat and shoot mass; sex and shoot mass; leaf traits and sex. To perform the statistical calculations, genotypes were divided—in accordance with their relationships—into 24 main groups numbered from 1 to 24 (Table 2), representing basic species and hybrids between species. The basic groups were divided into subdivisions according to differences in sex or other features and sequentially numbered as N.1, N.2, etc. The

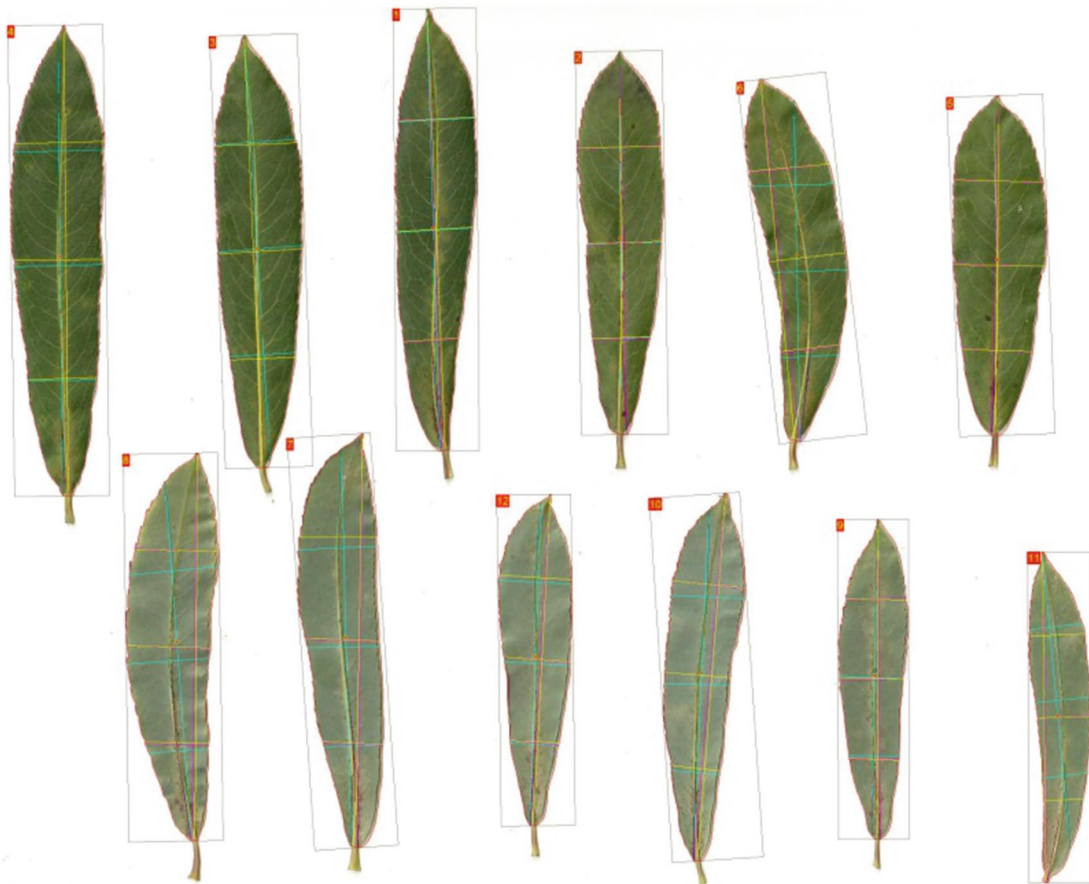


Fig. 2 Scanned and measured leaves of *Salix purpurea* '65', ♀

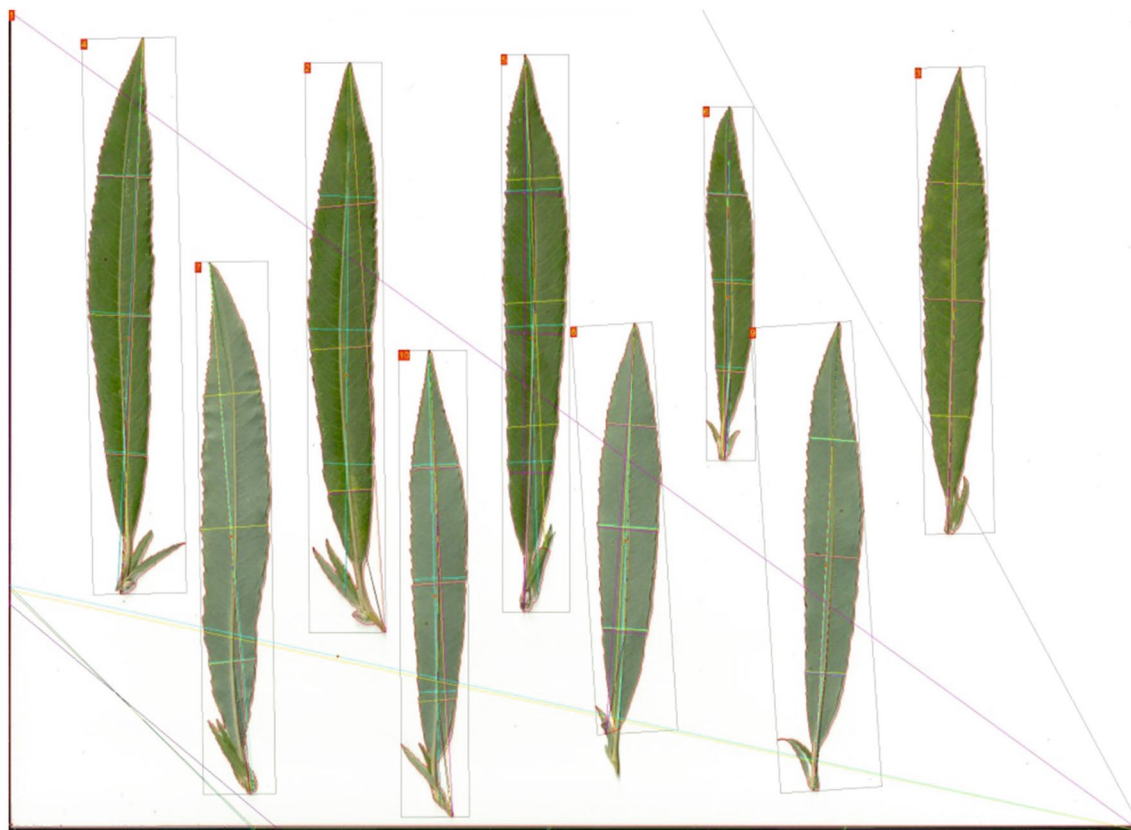


Fig. 3 Scanned and measured leaves of *Salix purpurea* × *triandra* × *viminalis* '15', ♀

greatest number of units (46) was distinguished in *S. purpurea* (17.11–17.56). Groups 1, 4, 8, 11, 13, 20 and 22–24 were represented by a single taxon only.

Shoot mass analysis of variance was used according to the model

$$y_{ijkl} = \mu + \alpha_i + \beta_j + \gamma_k + (\alpha_i\beta_j) + e_{ijkl}, \quad (1)$$

where, μ is the overall average, α_i is the the effect of sex, $i = 1, 2, \dots, 4$, β_j is the the effect of location, $j = 1, 2, 3$, γ_k is the the effect of taxon, $k = 1, 2, \dots, 91$, $\alpha_i\beta_j$ is the the effect of the interaction between i -sex and j -location, e_{ijkl} is the random error.

For comparisons of the average shoot mass, multiple comparisons on the basis of Tukey's method according to Ott (1984) were performed.

Results

Mortality

150 genotypes of willows were planted on each plot; however, after 3 years of cultivation a total of 94 genotypes

survived in all tested plots (Fig. 1a–c black rectangles). Some of the willow genotypes died at one or two of the three given plots: S-43—13 genotypes, S-52—16 and S-85—13 genotypes (Fig. 4) and 23 genotypes did not survive in any of the studied areas.

From the data presented in Fig. 4, it appears that the female genotypes showed the highest mortality, although it should be noted that among all tested willows female genotypes (category F) were definitely dominant (56.7% of all genotypes). The other sex categories showed the following shares: genotypes without flowers for which sex was not determined (category 0)—10.2%, males (category M)—27.6% and bisexual (category B)—5.5%. Therefore, the variation in the willows of the different sex categories in terms of the percent mortality, as shown in Fig. 5, differs from that shown by the willows when considering survival according to number.

Figure 5 also shows that bisexual specimens (generally hybrids) and specimens of unspecified sex had the highest mortality, which in turn may be a sign of their generally worse condition in comparison to that shown by the other sex categories, and this is emphasized by the lack of flowering. Additional conclusions can also be drawn from Fig. 5. It can be seen that for male specimens the S-85 area proved

Table 2 List of *Salix* genotypes and the average mass of shoots (kg); abbreviations: 0—indeterminate sex, F—female, M—male, B—bisexual, T—total

No	Name of taxon	Number of tested genotypes					Average mass (kg) of single shrub of tested genotypes				
		0	F	M	B	T	0	F	M	B	T
1	<i>Salix</i> × <i>blanda</i>		1			1	7.6				7.6
2	<i>S.</i> × <i>laurina</i>	1	1			2	1.9	2.0			2.0
3	<i>S.</i> × <i>pontederana</i>		2			2	4.0				4.0
4	<i>S.</i> × <i>rubens</i>		1			1	0.2				0.2
5	<i>S.</i> × <i>rubra</i>		12	4		16	3.4	2.7			3.2
6	<i>S.</i> × <i>sepulcralis</i>			1	5	6		5.7	4.6		4.8
7	<i>S.</i> × <i>smithiana</i>		1	6		7	8.0	4.9			5.3
8	<i>S. acutifolia</i>		1			1	1.0				1.0
9	<i>S. alba</i>	1		1		2	1.6		1.0		1.3
10	<i>S. alba</i> × <i>S. triandra</i>	1	1			2	0.7	4.3			2.5
11	<i>S. cinerea</i> × <i>S. triandra</i>		1			1	1.7				1.7
12	<i>S. cinerea</i> × <i>S. sp.</i>	1	1			2	3.6	5.6			4.6
13	<i>S. eriocephala</i> × <i>S. cordata</i>		1			1	1.3				1.3
14	<i>S. eriocephala</i>		8	1		9	3.0	3.0			3.0
15	<i>S. fragilis</i>	8	2	2	1	13	2.4	3.8	3.4	1.8	2.7
16	<i>S. myrsinifolia</i>		2			2	2.8				2.8
17	<i>S. purpurea</i>	1	28	16	1	46	1.6	3.2	4.2	1.7	3.4
18	<i>S. purpurea</i> × <i>triandra</i> × <i>viminalis</i>		2			2	10.3				10.3
19	<i>S. triandra</i>		5			5	2.9				2.9
20	<i>S. triandra</i> × <i>alba</i>		1			1	3.9				3.9
21	<i>S. viminalis</i>			2		2		3.9			3.9
22	<i>S. viminalis</i> × <i>S. sp.</i>			1		1		7.4			7.4
23	<i>S. fragilis</i> × <i>triandra</i>			1		1		3.1			3.1
24	<i>S. reichardtii</i>		1			1	1.0				1.0
Total		13	72	35	7	127	2.2	3.4	4.0	3.8	3.5

Fig. 4 Total number of genotypes that did not survive the 3 years of cultivation at locations S-43, S-52 and S-85 according to sex category. Legend: 0—genotypes without flowers for which sex was not determined, F—female, M—male, B—bisexual

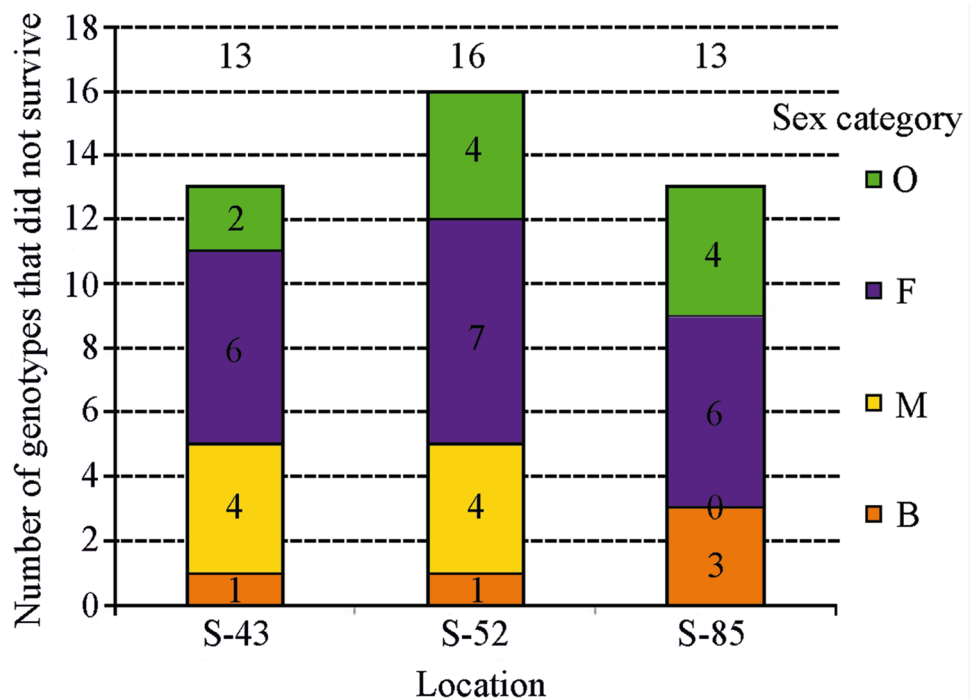
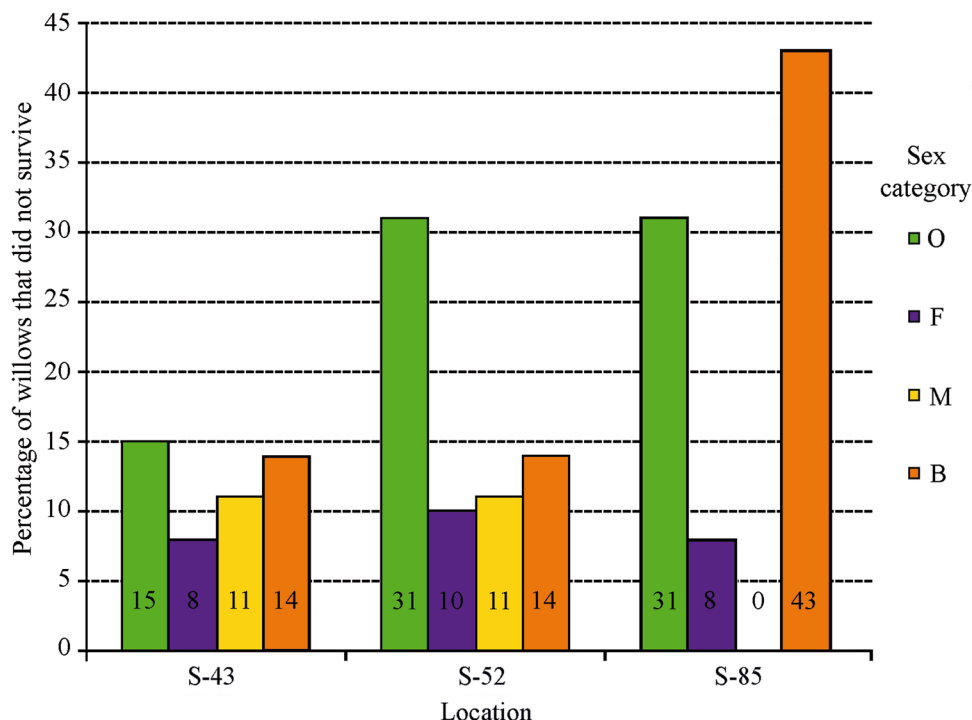


Fig. 5 Mortality (in %) of different genotypes following 3 years of cultivation according to sex category. Legend: O—genotypes for which sex was not determined, F—female, M—male, B—bisexual



to be the most beneficial, as all male genotypes survived the 3 years of cultivation (mortality = 0), while female specimens showed similar mortality across all three locations. It is worth noting, however, that the survival of female specimens was higher at the S-43 and S-52 locations than that of male specimens. Considering the soil conditions prevailing across all three research areas, among which the most favorable prevailed at the S-85 location, it can be argued that in better habitat conditions, male specimens have lower mortality, but when the conditions worsen, females survive better.

Finally, considering the mortality across all 150 genotypes, 127 were further examined, including 13 genotypes with indeterminate sex, 72 females, 35 males and 7 genotypes with male and female catkins on the same shrub (Tables 1 and 2) that survived in at least one of the three study areas (S-43, S-52, S-85).

Biomass: site interactions

The biomass—site interaction data are given in Fig. 6.

The weight of the shoots of willows growing at the S-85 location was twice as high as that at the other locations (S-43 and S-52). Due to the close proximity of the research areas, it could be assumed that of the three factors included in the term “habitat” (climate, soil and location), the key role in the differentiation of the tested plants was played by soil, especially the water regime and water balance. At location S-52, the soil is composed of clay. It is fertile ground but the supply of water depends on rain and snowfall, which

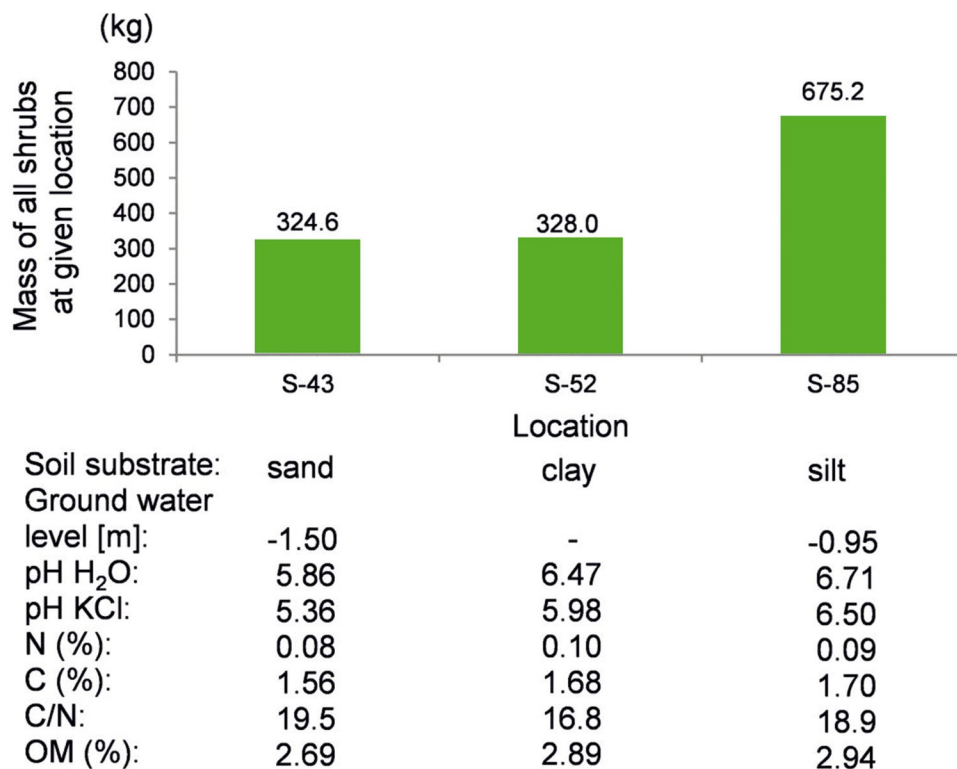
are variable across years in Poland, e.g., from 311.6 mm in 1989 to 723.6 mm per year in 1993 (Rutkowski et al. 2007). Old shrubs, which have deep root systems, can survive unfavorable years but for young plants, such conditions may have critical effects.

At site S-43 the soil is sandy, with the groundwater level at a depth of 1.50 m. For older shrubs such a situation is favorable but for young plants the first years of cultivation are vital because their root systems must be sufficiently long to reach the water. Optimal site conditions were noted at location S-85, where the soil is composed of silt and the groundwater level is at a depth of 0.95 m. These conditions were reflected in the high yield from this site. The biomass values given in Fig. 5 refer to the total mass of shoots of all 127 genotypes tested separately at each plot. At a spacing of 0.5×1.5 m, each plant occupied 0.75 m^2 , while all plants occupied a total area of 285.75 m^2 per plot. Thus, in the particular plots, all the evaluated willows had a fresh shoot mass of 1.14 kg/m^2 (S-43), 1.15 kg/m^2 (S-52), and 2.36 kg/m^2 (S-85).

Biomass: taxon interactions

The most important factor affecting the tested plants was taxonomic identity. Among the 127 tested genotypes the largest shoot mass occurred in 2 genotypes of *S. purpurea* \times *triandra* \times *viminalis* (Table 1). After 3 years of cultivation hybrid *S. purpurea* \times *triandra* \times *viminalis* ‘15’ produced an average, calculated from all three localities together, of 11.5 kg per shrub of fresh shoot mass, (S-43—14.5 kg, S-53—12.0 kg,

Fig. 6 Comparison of the total biomass at each studied plot [S-43 (Fig. 1a); S-52 (Fig. 1b) and S-85 (Fig. 1c)] and the soil characteristics of each area



S-85—8.0 kg), while *S. purpurea* × *triandra* × *viminalis* ‘46’ produced 9.0 kg (5.0, 9.5, and 12.5 kg, respectively). A summary of the biomass production of the tested genotypes is shown in Fig. 7.

For the willows that survived the 3-year study period at each of the three locations described above (94 of 127 genotypes), an analysis of variance was performed. The results of the analysis showed significant correlations between the

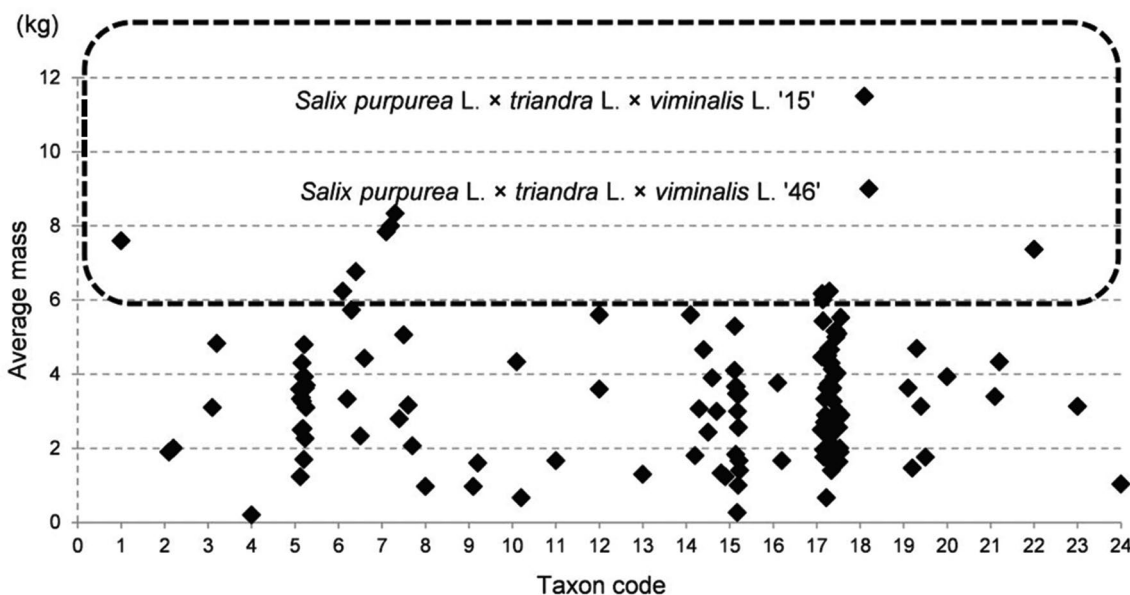


Fig. 7 Differentiation of the average fresh biomass of shoots of 3-year-old shrubs among all (127) tested genotypes. Species and hybrids were coded according to natural numbers from 1 to 24

(Table 1). The codes of all genotypes are given in Table 4. Each black point denotes one genotype. The dashed line indicates the best-growing willow genotypes (all top genotypes are hybrids)

mass of shoots and location, the mass of shoots and taxon, and the mass of shoots and sex (Table 4).

For the genotypes that survived at all three locations (Table 1, positions 1–94), Tukey’s test was also performed. Table 1 also includes genotypes for which Tukey’s test, due to an insufficient number of replicates (absence of the taxon at one or two of the three locations), was not carried out. These genotypes were, however, included in Table 1 (positions 95–127) to show that under suitable site conditions, the biomass of shrubs could be relatively high even if the survivability of a given taxon is generally low.

The results of Tukey’s test, presented in a Venn diagram (Fig. 8), clearly confirm the separation of the genotypes with the highest (*S. purpurea* × *triandra* × *viminalis*—letter “a”) and lowest (*S. alba* ‘Chermesina’—letter “e”) biomass.

Biomass: sex interactions

Tukey’s test performed for 94 genotypes revealed 3 different groups of willows:

- group “a”—bisexual specimens (Table 3: C3),
- group “ab”—male (Table 3: C2) and female (Table 3: C1) specimens,
- group “b”—specimens of indeterminate sex (Table 3: C0).

The results are also shown in Fig. 9 and Table 1.

The results of the 3-factor analysis of variance for the 94 genotypes of willows in regard to the interactions of sex—biomass, location—biomass, taxon—biomass and sex—location are shown in Table 4. Of course, the evaluation of the sex—location interaction does not make sense, but the “interaction insignificant” result was kept in Table 4 because

it shows the correctness of the test and strengthens the significance of the other demonstrated interactions.

Biomass: leaf feature and sex—leaf feature interactions

For *S. purpurea*, which represents the largest number of genotypes among all the tested genotypes (46 of a total of 127), an analysis of the relationships between location, biomass and leaf traits was also performed to evaluate:

1. the influence of leaf characteristics on biomass,
2. the relationships between leaf traits and sex.

For this purpose, 35 genotypes of *S. purpurea* that survived at all 3 locations (S-43, S-52 and S-85) were analyzed. Among these 35 genotypes, 3 sex categories were distinguished: indeterminate sex (1 genotype), female (23 genotypes) and male (11 genotypes) (Table 5). Their masses are illustrated by a heat map in which two-dimensional variables are represented by different colors (Fig. 10). Cluster

Table 3 The results of Tukey’s test performed for 94 genotypes divided into 4 sex groups (C0—indeterminate sex, C1—female, C2—male, C3—bisexual) tested in 3 locations; R number of genotypes in given sex category × 3 plots

Group	Average (±SD)	R	Min	Max	Tukey’s test
C3	5.44 (3.85)	9	0.5	13.5	a
C2	4.31 (2.84)	84	0.5	12.0	ab
C1	3.83 (2.82)	171	0.1	14.5	ab
C0	3.08 (1.99)	18	0.7	9.0	b

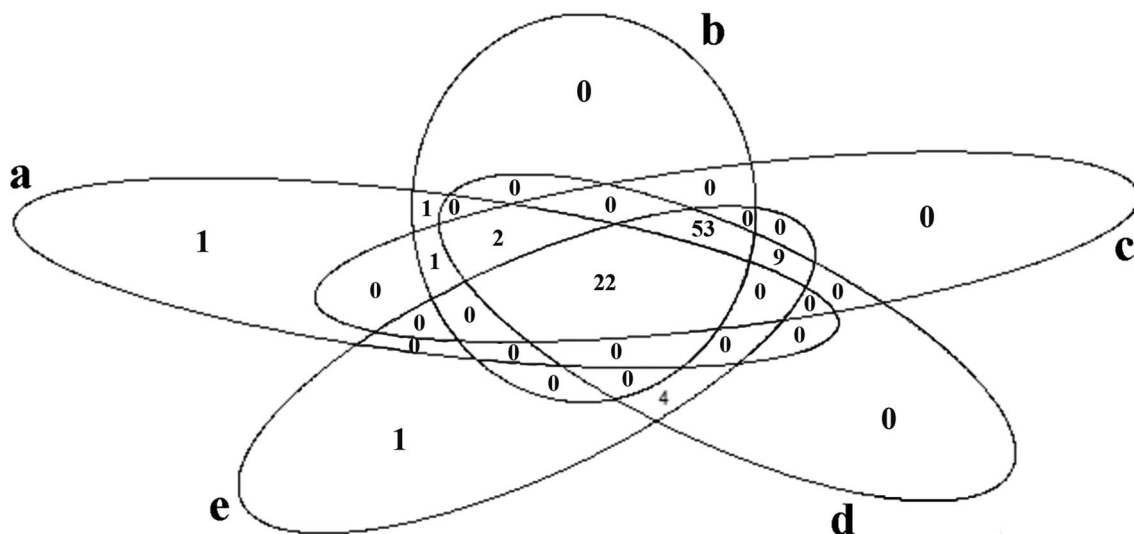


Fig. 8 Venn diagram for 94 genotypes evaluated with Tukey’s test. Letters “a” to “e” denote relations between the genotypes of willows with the highest (a) to lowest biomass (e)

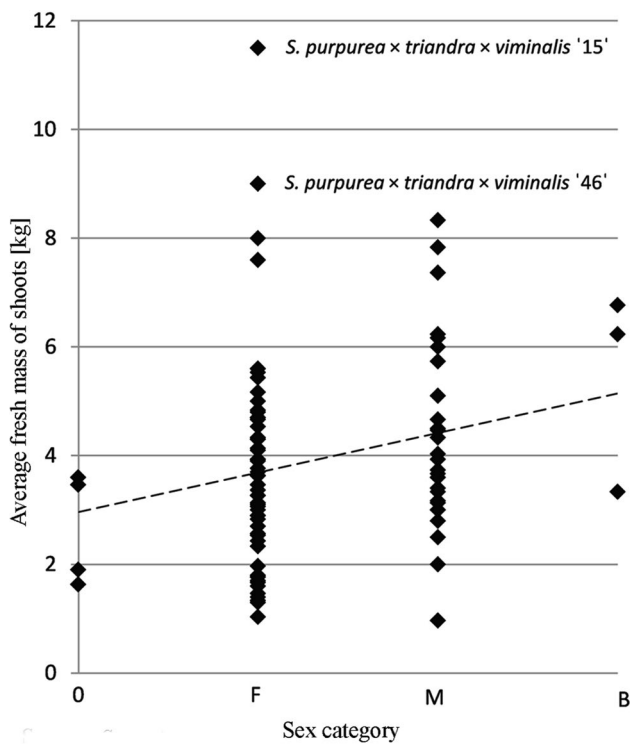


Fig. 9 Average fresh mass of shoots (in kg) for 94 genotypes grouped into 4 sex categories. Sex category 0—genotypes for which sex was not determined, F—female, M—male, B—bisexual

analysis enabled the grouping of the analyzed genotypes by location in such a way that the degree of combinations within one group was the largest, while that between groups was the smallest. Using Ward’s hierarchical clustering and the measure of Euclidean distance a tree diagram with three clusters (Fig. 10) was obtained.

In the heat map shown in Fig. 10, the differences between location S-85 and the others are clearly visible. Sex differences are less prominent, although the analysis of genotypes belonging to particular groups clearly shows these differences. Female specimens dominate in group 1, which accounts for 39% of all plants in this sex category (26% in group 2, 35% in group 3). Male specimens were strongly concentrated in group 3 (73%). Specimens for which sex was not determined occurred only in group 1.

Table 4 The results of the three-factor analysis of variance for 94 genotypes of willows

Interactions	df	Sum of squares	Mean square	F	p	Level of significance
Sex-biomass	3	46.6	15.55	4.219	0.00653	**
Location-biomass	2	599.5	299.76	81.332	<2e−16	***
Taxon-biomass	90	903.2	10.04	2.723	6.02e−09	***
Sex-location	6	41.1	6.85	1.858	0.09033	Interaction insignificant
Residuals	180	663.4	3.69			

Levels of significance: ** $p < 0.01$, *** $p < 0.001$, interaction insignificant ($0.05 \leq p \leq 1$)

In an attempt to identify common features of the genotypes in individual clusters, an analysis of leaf traits of *S. purpurea*, comprising the leaf area, leaf blade length and width, leaf slenderness (length–width ratio), and petiole length, was conducted. All data were given in the Catalogue of Willows published by Rutkowski et al. (2013). The genotypes grouped into cluster 1 had, on average, the shortest petioles (4.8 mm), the shortest length of the leaf blade (70.5 mm) and the smallest leaf width (14.6). The petioles of the genotypes grouped into cluster 2 were the longest (average 6.5 mm). The genotypes grouped into cluster 2 also had the longest (75.6 mm) and the widest (16.8 mm) leaf blades on average.

Statistical analysis of the leaf traits showed their relationship to sex (Table 6). On average, male specimens have longer and wider leaves and thus a larger leaf area than specimens in the other two sex categories. They also have the longest petioles by far. The smallest leaves and the shortest petioles were found in willows of indeterminate sex (one genotype). Longer petioles may help the leaves move more efficiently towards the sun. In addition to a longer and wider leaf blade, this trait may affect biomass production through the better use of solar energy in photosynthesis.

Discussion

Approximately 18 willow species show exceptions from dioecy. Thirteen of them occur naturally, three arose in experiments and two are found under both natural and experimental conditions (Mirski 2014). The main objective of this study was to test whether the sex of different willow species affects their biomass and mortality, but the results show that we can add 2 more to the list of 18 species mentioned by Mirski: *S. fragilis* and *S. purpurea*. The conducted studies also included bisexual *S. x sepulcralis*, which is a hybrid of *S. babylonica* L. × *S. alba* L., but bisexual catkins are not rare in hybrids (Stace 1999).

Yang et al. (2020) have shown that the mortality of male individuals of *S. suchowensis* is higher than females. Our results confirmed the impact of site conditions on the mortality of males and females but indicated that male specimens show higher survival when they grow under better

Table 5 List of willows analyzed in the heat map; abbreviations: O—indeterminate sex, F—female, M—male

No	<i>Salix purpurea</i> genotype name	Sex category	Average					Group
			Length of leaf blade (mm)	Width of leaf blade (mm)	Area of leaf blade (mm ²)	Leaf elongation	Length of petiole (mm)	
1	'155/142'	O	34	9	220	3.7	3.6	1
2	'7'	F	71	18	840	4.0	7.0	2
3	'16'	F	84	18	1051	4.6	6.6	1
4	'23'	F	76	15	717	5.2	3.4	1
5	'57'	F	66	21	997	3.2	5.2	1
6	'58'	F	85	15	878	5.7	5.6	1
7	'61'	F	61	14	553	4.3	5.3	3
8	'63'	F	71	17	785	4.3	6.5	2
9	'69'	F	85	18	1083	4.8	5.9	2
10	'73'	F	69	14	713	4.8	6.1	3
11	'75'	F	70	16	840	4.4	5.0	3
12	'77'	F	64	10	437	6.2	3.8	1
13	'79'	F	69	15	721	4.5	6.6	3
14	'81'	F	68	13	625	5.1	6.6	3
15	'82'	F	68	15	672	4.6	5.9	3
16	'87'	F	74	17	820	4.3	6.8	2
17	'126'	F	74	19	956	3.9	4.3	2
18	'39/26'	F	83	18	1120	4.6	5.2	1
19	'149/136'	F	39	9	228	4.4	3.1	1
20	'153/140'	F	72	14	685	5.0	2.9	1
21	'156/143'	F	58	14	538	4.1	6	3
22	'168/155'	F	72	16	798	4.4	6.5	2
23	Scharfenbergensis '76'	F	91	19	1207	4.9	7.1	1
24	Helix Pyramidalis '160/147'	F	77	13	664	5.8	6.5	3
25	'Angustifolia 68'	M	82	15	818	5.6	7.5	3
26	'Angustifolia 72'	M	88	14	839	6.1	7.3	2
27	'66'	M	67	16	738	4.2	6.6	3
28	'67'	M	94	16	908	6.0	8.1	3
29	'70'	M	70	17	820	4.2	7.5	2
30	'74'	M	77	18	937	4.4	6.5	3
31	'78'	M	64	16	725	4.1	5.7	3
32	'84'	M	65	15	666	4.4	5.5	3
33	'125'	M	82	14	733	5.7	6.4	1
34	'136/123'	M	70	16	813	4.4	6	3
35	'151/138'	M	92	21	1215	4.4	6.8	3

conditions. Under worse conditions female individuals have higher survivability. This observation is similar to Tognetti (2012) conclusion that females are more common in high-resource and males in low-resource microsites, but finally survivability is not the same as being more common.

Sex can indirectly affect plant growth, e.g., through the different preferences of herbivorous insects (Kabir et al. 2014) or roe deer (Moritz et al. 2018) for female and male specimens of willows. Therefore, drawing attention to willow sex seems to be essential, especially for owners of willow plantations. This was clearly indicated

in this paper for *S. purpurea*, for which the shoot mass of male specimens was 31% greater than that of female specimens. This result, however, did not unambiguously confirm Harris and Pannell's (2008) thesis that females tend to be smaller than males in woody dioecious plant species because in the current investigation, the female specimens of other genotypes (*Salix* × *rubra*, *S.* × *smithiana*, *S. fragilis*) had a greater shoot mass than male specimens. This clearly indicates the need to correlate sex with the taxon. Especially, as in many scientific papers devoted to willow plantations this topic is often overlooked (Weih

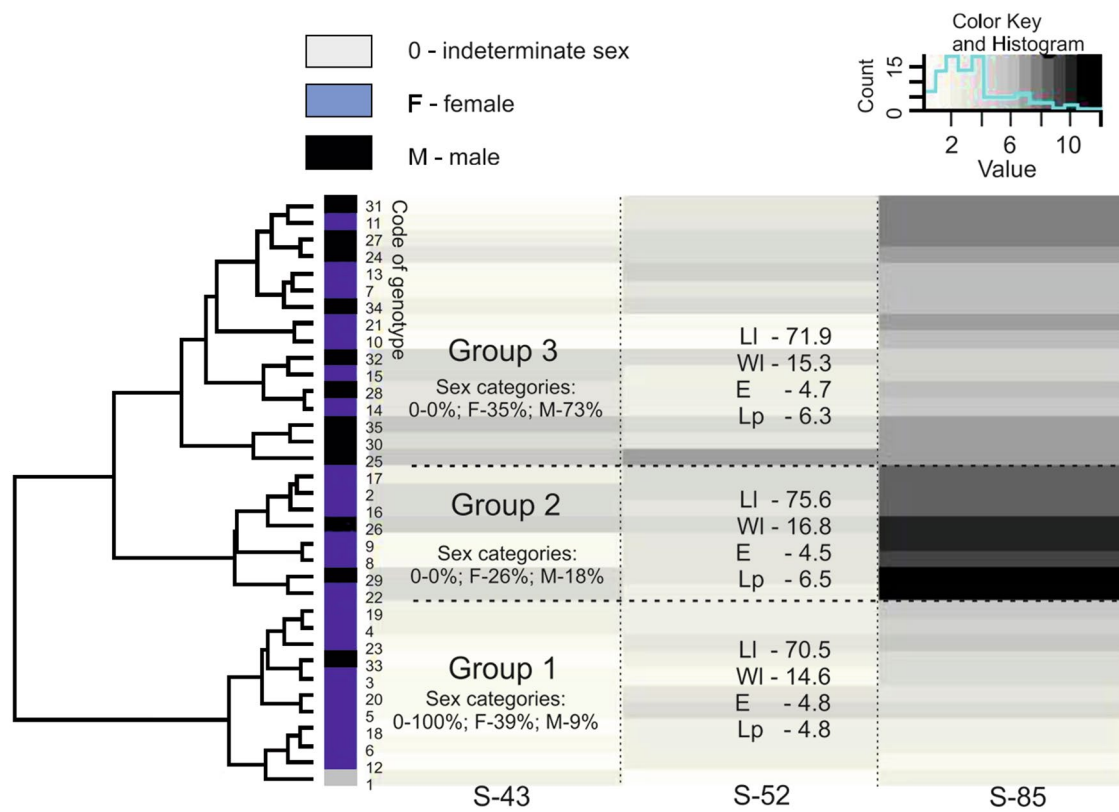


Fig. 10 Division into clusters in relation to the length of the leaf blade (LI), leaf blade width (WI), leaf slenderness (E) and petiole length (Lp)

Table 6 Differentiation of average length of leaf blade (LI), width of leaf blade (WI), area of leaf blade (AI), leaf elongation (E) and length of petiole (Lp) in three sex categories of *Salix purpurea*

Sex category	No. of genotypes	Average/SD				
		LI (mm)	WI (mm)	AI (mm ²)	E	Lp (mm)
0—sex unknown	1	34.0/–	9	220	3.7	3.6
1—females	23	71.6/10.8	15	785	4.6	5.5
2—males	11	77.4/10.9	16	823	4.9	6.7

and Nordh 2005; Mitsui et al. 2010; Sevel et al. 2012; Aust et al. 2014; Han et al. 2020).

Leaf characteristics also reflected relationships with sex, as demonstrated on *Hippophae rhamnoides* (Li et al. 2007). The obtained results allow us to conclude that male specimens of *S. purpurea* have, on average, longer and wider leaves and thus a larger leaf area than female specimens. They also have much longer petioles. The smaller area of leaves in females can be associated with the energy that females have to expend in connection with the production of seeds. The correlation between this energy expenditure and leaf area is also indicated by the results obtained in this work showing that the smallest leaves and the shortest petioles were associated with willows of indeterminate sex, which may be a reflection of the relatively poor condition of these plants.

Differences between treatments in the biomass production of *Corema album* female individuals found by Álvarez-Cansino et al. (2010) confirmed the results of shoot elongation and supported the existence of a balance in resource investment between reproduction and maintenance of vegetative growth. Our study reveals a similar effect in the case of willows. There are still many questions associated with sexual dimorphism but we hope that the results described in this paper can help answer some of them.

Conclusions

Based on 8100 described and measured plants representing 150 genotypes of willows, including 8 species, 16 interspecies hybrids, cultivars and sex-differentiated specimens, we

showed that: (1) males of the tested *Salix* genotypes had a higher biomass than females but the highest biomass yields were produced by bisexual specimens; (2) males of the tested willows showed a greater survival rate than females but only under better site conditions; when the site conditions were poorer, males had higher mortality rates than females; (3) hybrids had higher shoot biomass than the tested pure species; (4) for *S. purpurea*, which represented the largest number of genotypes in the study (46), male specimens had longer and wider leaves and thus a larger surface area on average than female specimens. They also had clearly longer petioles. The smallest leaves and the shortest petioles were associated with willows without catkins.

The results of this study indicate that in all works devoted to willows, both practical and scientific, it is necessary to refer not only to the willow species and environmental conditions in which they grow but also to the sex of the examined or cultivated individuals.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethics approval and consent to participate This article does not contain any studies involving human participants or animals performed by any of the authors.

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