CASE REPORT



Primary pulmonary extranodal NK/T cell lymphoma of an elderly adult: a case report and literature review

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Abstract

Extranodal natural killer/T-cell lymphoma (ENKTCL), nasal type, is rare and aggressive and often involves the nose, nasopharynx, and upper aerodigestive tract. The non-nasal type can affect the skin, salivary glands, gut, testes, brain, salivary glands, and other sites. Primary ENKTCL of the lung is rare. Here, we report a 68-year-old non-smoking female who presented with fever, dry cough, and night sweats. The chest image showed lung consolidation in the right lower lung field. Pulmonary biopsy showed diffuse abnormal lymphocyte infiltrate in the necrotic exudate. Immunohistochemical data indicated that the tumour cells were positive for CD56, granzyme B, CD3, and TIA. Using in situ hybridization, Epstein–Barr virus-encoded ribonucleic acid (EBER) was detected. There was no evidence to indicate extrathoracic lymphoma involvement. Primary pulmonary ENKTCL was therefore diagnosed. The patient underwent chemotherapy using the P-GEMOX regimen (pegaspargase, gemcitabine, and oxaliplatin) and is still alive.

Keywords Extranodal NK/T-cell lymphoma · Epstein–Barr virus · Lung cancer · Lymphoma · P-GEMOX

Introduction

Extranodal NK/T-cell lymphoma (ENKTCL), nasal type, is a rare and invasive neoplasm with a poor prognosis that is closely associated with Epstein–Barr virus (EBV) infection. While lymphoma can occur in the nasal and upper aerodigestive regions, the skin, salivary gland, testis, or gastrointestinal tract, and causes lymphadenopathy and hepatosplenomegaly, the nasal and upper aerodigestive regions are the most common invasive sites [1, 2]. Primary pulmonary ENKTCL is

very rare, with no more than twenty cases reported in the English literature [3–15]. Here, we describe a case of primary pulmonary ENKTCL and review the related literature.

Clinical history

A 68-year-old non-smoking female presented with fever, dry cough, and night sweats in the clinic. The chest computed tomography (CT) scan revealed a soft tissue mass in the right

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lung (Fig. 1a). The T-SPOT, bone scan, and head CT did not show any abnormalities. Her temperature was 36.1 °C, pulse 78 bpm, breathing 18 bpm, and blood pressure 124/75 mmHg. No palpable lymph nodes were found on the neck, supraclavicular area, axillary, or inguinal lymph nodes. Laboratory examination results showed the following: white blood cell count, 6.7×10^9 /L; neutrophil percentage, 74.6%; haemoglobin, 111 g/L; platelet count, 296×10^9 /L; total

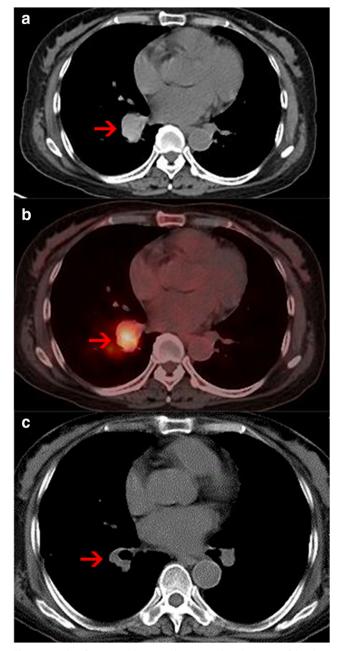


Fig. 1 Imaging features. **a** Computed tomography (CT) scan of the chest showed dense consolidation in the right lung. **b** Positron emission tomography—computed tomography (PET/CT) scan showed a mass in the right lung with high-18F-fluorodeoxyglucose uptake. **c** After 5 cycles of treatment, chest CT scan revealed a reduction in the area of consolidation



protein (TP), 66.3 g/L: blood urea nitrogen (BUN), 3.88 mmol/L; creatinine (Cr), 43.9 µmol/L; C-reactive protein (CRP), 46.92 mg/L; LDH, 194 U/L; procalcitonin (PCT), 0.03 ng/mL; and serum ferritin, 610.95 ng/mL. Tumour markers were all normal. She was then diagnosed with pneumonia and treated with cephalosporins. However, her clinical condition did not obviously improve. Bronchoscopic biopsy was not performed because she declined the procedure. Then, CT-guided needle biopsy was performed, and a final diagnosis of ENKTCL was made based on immunohistochemistry, Tcell receptor (TCR) gene rearrangement and EBER analysis results. A positron emission tomography-CT (PET-CT) scan showed significant fluorodeoxyglucose (FDG) uptake in the right lung (Fig. 1b). No other organ involvement was found by PET/CT. Based on the neoplastic morphology and the immunohistochemical examination, a diagnosis of primary pulmonary ENKTCL was made. There was no obvious abnormality in bone marrow biopsy. According to the National Comprehensive Cancer Network (NCCN) guidelines, patients are treated using the pegaspargase, gemcitabine, and oxaliplatin (P-GEMOX) regimen [16].

Materials and methods

Morphology and immunohistochemistry

Lung specimens were fixed in formalin (10%) and processed routinely. Paraffin-embedded blocks were sectioned (4 μ m thick) and stained with haematoxylin-eosin (H&E). Immunohistochemical analysis was carried out with a Ventana BenchMark GX system using the following primary antibodies: anti-CD56 (123C3.D5), anti-CD3 (SP7), anti-TIA (TIA-1), and anti-granzyme B (GZB01). Working solutions of the above antibodies were purchased from Fuzhou Maxim, China. Additionally, EBER in situ hybridization (EBER1 DNP Probe, Ventana) was performed.

Multiplex polymerase chain reaction for TCR gene rearrangement

Human peripheral blood mononuclear cells (PBMCs) from patients were isolated by density gradient centrifugation using Ficoll-Paque (BD, USA), and genomic DNA was extracted using the Blood Genomic DNA Kit (Fastagen, China). The multiplex polymerase chain reaction (PCR) protocols and the 56 primers used for TCR gene analysis were standardised BIOMED-2 protocols. The 56 primers used for TCR clonal rearrangement were selected from BIOMED-2 [17]. Nine groups of PCRs were conducted, including five groups for the VDJ domains of TCR γ , one group for the VDJ/DD/VD/DJ domains of TCR δ , and one group for the positive control. The PCR

cycling conditions were as follows: denaturation at 95 °C for 7 min; 45 cycles of denaturation at 95 °C for 45 s, annealing at 60 °C for 45 s, and extension at 72 °C for 90 s; and a final extension at 72 °C for 10 min. All PCR products were separated via 6% polyacrylamide gel electrophoresis in a DG-600C electrophoresis system (Dingguo, China) according to the manufacturer's instructions. The gels were collected and analysed using a ChemiDoc XRS+ imager (Bio-Rad, USA).

Results

Histopathologic sections revealed massive prominent necrosis with diffuse dense medium-sized lymphoma cells infiltrate. These cells were markedly atypical, and nuclei were irregular, exhibiting an angiocentric angiodestructive quality (Fig. 2a). Immunohistochemical analysis revealed that the cells had the following immunotypes: CD56(+), granzyme B(+), CD3(+), TIA(+), CD20(-), CD5(-), CD8(-), CD79a(-), CD10(-), BCL-6(-) (Fig. 2b-e). Furthermore, in situ hybridization showed that these cells were strongly positive for EBER (Fig. 2f), and EBV DNA tests showed 1.24 \times 10 5 copies/mL (normal, 5 \times 10 3 copies/mL). TCR gene rearrangement studies for TCR β , TCR γ , and TCR δ were negative.

Following 5-cycle P-GEMOX chemotherapy, the patient's symptoms were significantly alleviated, and a repeat CT scan showed decreasing consolidation in the right lung (Fig. 1c). The patient achieved partial remission (PR) and is still alive.

Discussion

ENKTCL, nasal type, is rare and accounts for 10% of all peripheral T-cell lymphomas (PTCLs) worldwide. Primary pulmonary NK/T-cell lymphoma represents 0.5 to 1% of all primary pulmonary lymphomas [12, 18]. ENKTCL is geographically predominant in East Asia and southern North America but rare in Europe and Africa. In the World Health Organization (WHO) records, most patients with ENKTCL have a median age of 44-54 years, and it affects more males than females. Primary tumour sites of ENKTCL are located in the upper airway region. Usually, in the nasal area, the lesion presents as an invasive mass with surrounding tissue hyperaemia and oedema. It erodes the hard palate, leading to extensive midfacial lesions, and then extends to other adjacent and nonadjacent facial tissues, but bone marrow involvement is uncommon. The presentation of non-nasal NK/Tcell lymphoma can vary, depending on the involved sites. Histologically, features of ENKTCL show no obvious difference irrespective of their anatomical location. Lymphomatous infiltrate often exhibits diffuse and permeative infiltration. Growth patterns present angiocentricity and angiodestruction, leading to coagulative necrosis. Cytologically, most neoplastic cells are medium-sized or comprise a mixture of small and large cells with irregularly folded nuclei. In other types of lymphoma, coagulation necrosis and vascular invasion are not obvious. Its typical immunophenotype is CD56(+), cytoplasmic CD3 ε (+), and CD2(+), and it is positive for cytotoxic molecules (TIA-1, granzyme B, and perforin) but negative for surface CD3 and

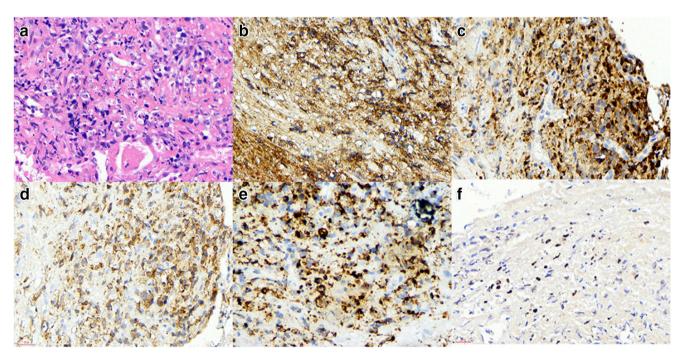


Fig. 2 a Lung biopsy showed diffuse infiltrative atypical medium-sized tumour cells had irregular nuclei, moderate in amount of cytoplasm and pale to clear in extensive coagulative necrosis (H&E, × 400 magnification). **b–e** Immunohistochemical analysis showed that these cells were

positive for CD56 (\times 400 magnification), granzyme B (\times 400 magnification), CD3 (\times 400 magnification), and TIA (magnification, \times 400). **f** Epstein–Barr virus-encoded RNA was proven by in situ hybridization



Table 1 Details of the clinicopathological features and outcomes of primary pulmonary NK/T-cell lymphoma

SOB ^a , cough, fever Dyspnoea, cough, fever SOB, cough, fever Cough, haemoptysis Fever Fever Dyspnoea, cough, fever Cough, SOB, fever Cough, sputum Fever Cough, sputum Weakness, cough Weakness, cough	Image studies Immunophenotype and EBER		Treatment and outcome
134/F Dyspnoea, cough, fever 31/M SOB, cough, fever 80/M Cough, haemoptysis 50/M Fever 73/F Fever 53/M Dyspnoea, cough, fever 39/M Cough, SOB, fever Cough, SOB, fever 46/M Cough, sputum 47/M Fever, cough 44/M Weakness, cough, fever 34/F Dyspnoea, fever	Consolidation diffuse nodules CD56(+) CD3(+)	Died	
1] 31/M SOB, cough, fever 80/M Cough, haemoptysis 50/M Fever 73/F Fever 53/M Dyspnoea, cough, fever 39/M Cough, sputum, fever 55/M Cough, sputum 47/M Fever, cough 44/M Weakness, cough, fever 34/F Dyspnoea, fever	Consolidation, pleural effusion		died
80/M Cough, haemoptysis 50/M Fever 73/F Fever 53/M Dyspnoea, cough, fever 39/M Cough, SOB, fever 46/M Cough, sputum, fever 55/M Cough, sputum 47/M Fever, cough 1 44/M Weakness, cough, fever	Consolidation		CT, HSCT', died
1] 50/M Fever 73/F Fever 53/M Dyspnoea, cough, fever 39/M Cough, SOB, fever 46/M Cough, sputum, fever 55/M Cough, sputum 47/M Fever, cough 1 44/M Weakness, cough, fever 34/F Dyspnoea, fever			lied
73/F Fever 53/M Dyspnoea, cough, fever 39/M Cough, SOB, fever 46/M Cough, sputum, fever 55/M Cough, sputum 47/M Fever, cough Meakness, cough, fever 34/F Dyspnoea, fever	Nodules $CD56(+), CD3\varepsilon(+), EBER(+)$	EBER(+) CT, died	lied
53/M Dyspnoea, cough, fever 39/M Cough, SOB, fever 46/M Cough, sputum, fever 55/M Cough, sputum 47/M Fever, cough Weakness, cough, fever 34/F Dyspnoea, fever	Space-occupying lesion CD56(+), CD3ε(+), TIA1(+), EBER(+)	TIA1(+), EBER(+) Died	
39/M Cough, SOB, fever 46/M Cough, sputum, fever 55/M Cough, sputum 47/M Fever, cough Meakness, cough, fever 34/F Dyspnoea, fever	Consolidation, pleural effusion	$CD56(+)$, granzyme B(+), $CD3\varepsilon(+)$, EBER(+) CT, Pl	CT, PBSCT ^d , died
46/M Cough, sputum, fever 55/M Cough, sputum 47/M Fever, cough Weakness, cough, fever 34/F Dyspnoea, fever	J	$CD56(+)$, granzyme B(+), $CD3\varepsilon(+)$, $TIA1(+)$ CT, CR^e	$^{2}\mathrm{R}^{\mathrm{e}}$
55/M Cough, sputum 47/M Fever, cough J 44/M Weakness, cough, fever 34/F Dyspnoea, fever		3 (+), EBER(+) Died	
47/M Fever, cough 44/M Weakness, cough, fever 34/F Dyspnoea, fever	GGOs ^f CD56(+), granzyme B(+), EBER(+),	B(+), $EBER(+)$, CT	
44/M Weakness, cough, fever 34/F Dyspnoea, fever	Multiple nodules CD56(+), CD3 (+), TIA1(+), EBER(+),	$TIA1(+)$, $EBER(+)$, CT , PR^g	oR.g
34/F Dyspnoea, fever	Multiple nodules (CD56(-), granzyme B(-), CD3(+), TIA1(+), EBER(+), Died	
	J	CD56(-), granzyme B(+), CD3 (+), TIA1(+) Died	
Our case 68/F Cough, fever Consolidation	Consolidation CD56(+), granzyme	CD56(+), granzyme B(+), CD3(+), TIA(+), EBER(+) CT, PR	N.

SOB shortness of breath, b CT chemotherapy, c HSCT haematopoietic stem cell transplantation, d PBSCT auto-peripheral blood stem cell transplantation, c CR complete remission, f GGOs ground glass opacities, gPR partial remission

TCR gene rearrangement. In most cases, EBER is detected by in situ hybridization, which indicates that EBV plays a highly important role in its pathogenesis [2, 19]. In our case, mediumsized atvoical lymphoid and coagulative necrosis were easily found. Immunohistochemical staining confirmed that these cells were reactive for CD56, granzyme B, CD3, and TIA but were negative for CD20, CD5, CD8, CD79a, CD10, BCL-6, and TCR rearrangement. EBER was positive, and EBV DNA levels were 1.24×10^5 copies/mL. The combination of histopathology and immunophenotyping along with EBV status led to a diagnosis of ENKTCL. Based on the PET-CT results, there were no other involvements except the lung; thus, we can confirm that NK/T-cell lymphoma originated from the lung. Because of the unique pathological characteristics of the disease, it is often misdiagnosed as other diseases, such as Wegener granulomatosis (GPA) and PTCL-not otherwise specified (NOS). GPA is a systemic autoimmune condition that often occurs in the upper and lower respiratory tract and kidney. Patients can experience recurrent sinusitis and exhibit lung nodules, inflammatory infiltration, glomerulonephritis, and mucocutaneous lesions. The histological characteristics of this disease are vasculitis and non-caseating granulomas. In the initial stage, the clinical features of ENKTCL and GPA are very similar. However, the sera of most patients with GPA contain antineutrophil cytoplasmic antibodies (ANCA) and lack EBER. Furthermore, the prognosis of GPA is much better than that of ENKTCL. PTCL-NOS represents a heterogeneous category of mature T-cell origin lymphomas affecting nodal and extranodal tissues. As the clinical manifestations are non-specific, we should rely on immunophenotyping for diagnosis. These neoplastic cells have the following markers: surface CD3(+), cytoplasmic CD3ε(-), CD56(-), and EBER(-), and TCR rearrangement (+). Thus, repeat biopsy and testing for CD56 or EBV-positive lymphocytes can increase the sensitivity and specificity for ENKTCL diagnoses.

Primary pulmonary ENKTCL is extremely rare. A list of retrospective clinical studies on similar English cases is provided in Table 1, encompassing studies from China, Japan, Korea, and the USA. Regarding the patients with these available cases, nine were male, and five were female, for a male-to-female ratio of approximately 1.8:1, and ranged in age from 31 to 80 years. Most patients presented with fever, cough, expectoration, and dyspnoea. The most common imaging characteristics included alveolar infiltration, pleural effusion, nodules, and ground glass opacities (GGOs). Immunologically, the patients were positive for CD56, cytoplasmic CD3ε, TIA-1, granzyme B, and perforin and negative for surface CD3, CD20, and CD79, and 64.3% were positive for EBV. The prognosis of these patients was generally poor. Three of the patients improved, but most died soon after diagnosis. Cyclophosphamide, doxorubicin, vincristine, and prednisone (CHOP)-based chemotherapy has been reported in some studies [8, 10, 14], but the patients' symptoms did not obviously improve. Our patient underwent 5 cycles of



chemotherapy using the P-GEMOX regimen according to the NCCN guidelines for the treatment of ENKTCL. Currently, the patient's symptoms have improved. She is still alive and will continue to receive further treatment.

Conclusion

We report a rare case of primary pulmonary ENKTCL and summarise previous reported articles. If a patient presents with persistent cough, sputum, and fever, the possibility of this disease should not be ignored. Because of the non-specific clinical symptoms and imaging findings, the correct diagnosis of primary pulmonary ENKTCL depends on histopathology and immunopathological data. Currently, therapeutic strategies are based on conventional ENKTCL treatment, but the prognosis of this disease originating from the lung is poor. Based on the NCCN guidelines, patients with extranasal involvement of NK/T-cell lymphoma should be treated with haematopoietic stem cell transplantation (HSCT) if they are in good health. However, because our patient was older and generally in poor condition (Eastern Cooperative Oncology Group (ECOG) score > 2), we did not recommend HSCT [16].

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflicts of interest.

Informed consent Informed consent was obtained from all participants included in the study.

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