



# Tumor Type-Agnostic Treatment and the Future of Cancer Therapy

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Published online: 10 October 2018  
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It is fascinating to see how the science of cancer therapy has evolved. We first classified tumors as “solid” or “liquid” and created the specialties of oncology and hematology to later discover that the shape of the tumors has nothing to do with their etiology, so we ended up combining both specialties. Next, we proceeded to classify cancers according to the organ they grow in, thinking that the origin of the tumors is what causes their biological behaviors, and could guide us in understanding and fighting them properly. After so many years taking this approach, with both tremendous successes and deep disappointments, we are now beginning to appreciate that there is much more complexity to cancer biology than simply the tissue that tumors arise from.

Molecular mechanisms (DNA mutations, translocations, deletions, fusions, etc.) are responsible for the origin and behavior of most tumors. We are starting to change the focus of cancer therapy from organ-specific treatments to molecular marker-specific approaches. This has very recently become a reality when in May 2017 the US Food and Drug Administration (FDA) granted accelerated approval for the anti-PD-1 immune checkpoint inhibitor pembrolizumab in adult and pediatric patients with locally advanced or metastatic solid tumors that are mismatch-repair deficient (dMMR) or microsatellite instability-high (MSI-H), who have progressed after prior treatment and who have no satisfactory alternative treatment options [1]. This “tissue-agnostic” approval has created a new paradigm in oncology.

In the current issue, Kummar and Lassen [2] present a very comprehensive review of NTRK gene aberrations as one example of success in using this tumor site-agnostic approach. The authors review the diagnostic and treatment strategies that are being implemented to deal with NTRK-fusion genes and the diseases that they cause.

NTRK genes encode for the Trk-family of tyrosine kinases: TrkA, TrkB, and TrkC (encoded by NTRK1, NTRK2, and NTRK3). Normally, these proteins are involved in the development of the nervous system [3]. However, Trks are also present in solid tumors as fusion proteins responsible for the growth of cancer cells, and these oncogenic fusions are associated with poor survival in lung cancers and other tumor types [4]. As seen with several other oncogenes (e.g., ALK, BRAF, ROS1, and others.), NTRK-fusions are present in several different tumors (so far, they are actionable in 17 tumor types). These genomic alterations are becoming a prime example for why the tumor site-agnostic approach might be the new paradigm in fighting cancer [5].

Despite this promise, there are still many challenges that lie ahead. For the molecular diagnosis of these genetic aberrations, fluorescence in situ hybridization (FISH), immunohistochemistry (IHC), reverse-transcriptase polymerase chain reaction (RT-PCR), and next generation sequencing (NGS) of DNA or RNA (or cfDNA), are all possible options. Each of these approaches has strengths and weaknesses, but we also have to play this in the context of the workup for other genetic abnormalities, and keep in mind that in many instances tumor tissue specimens are limited. As an example, in lung cancer we currently perform FISH for ALK and ROS1 translocations, and adding three more FISH tests for each of the NTRKs (and maybe one more for RET fusions) will markedly increase the cost of workup for these patients. Furthermore, the gene fusion partner, which might become relevant in the future, will not be identified by this technique. RT-PCR is a very sensitive assay, but we would need a lot of primers to cover all known NTRK genetic abnormalities (there are more than 60 NTRK fusions documented to date). Is the solution to develop improved IHC methods, like we are doing for ALK, and hopefully will one day have for ROS1 and NTRK

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oncogenes? Or should we try to establish NGS as the standard of care, to detect a large panel of possible genetic alterations, followed by a more limited follow up, for example when we look for TKI-resistant ALK variants? Nonetheless, for those who are not fans of NGS, we can say that at the moment, several NTRK 1–3 introns are not well covered by currently available NGS panels. Should we do whole genome NGS to find all possible alterations while significantly increasing the cost of the diagnostic workup? These are some of the questions that remain while we increase the accuracy of available diagnostic techniques and make them more cost effective.

For the treatment of NTRK-rearranged tumors, we are fortunate to already have clinical data available from phase I studies with two agents, entrectenib and larotrectenib, with acceptable toxicities regardless of tumor type, patient age, and fusion type [5, 6], and more data is coming from other agents in development, including CEP-701, ARRRY-470, DS-6051b, and TPX-0005 [7–10]. While NTRK-fusions are not very frequent in most cancers (less than 1%), in common tumors like lung cancer, where this year we can expect more than 235,000 new patients in the US alone, this 1% becomes an important group of patients who will benefit from such novel targeted agents.

Other tumor site-agnostic genetic abnormalities that can be targeted include BRAF mutations, and dMMR or MSI-H. To date, immunotherapy has become a panacea for almost all tumor types, especially after the US FDA granted a tumor type-agnostic approval to pembrolizumab, as mentioned before [1]. This indication covers patients with solid tumors that are dMMR or MSI-H, have progressed following prior treatment, and who have no satisfactory alternative treatment options, as well as patients with colorectal cancer who have progressed following treatment with certain chemotherapy drugs.

MSI involves the gain or loss of nucleotides from DNA elements composed of repeating motifs, which occur as alleles of variable lengths, called microsatellite tracts [11]. MSI can result from inherited mutations or originate somatically. Tumors are classified as dMMR if they have somatic or germ line mutations in MMR genes (Lynch syndrome results from inherited mutations of known MMR genes). MSI can also be the result of certain epigenetic changes or altered microRNA pathways affecting MMR proteins [12]. Although MSI is most commonly found in colon and endometrial cancers, it has also been detected in as many as 24 cancer types, suggesting that MSI is a generalized cancer phenotype [13, 14]. Noteworthy, the genomes of dMMR tumors contain a high number of somatic mutations, and hence make them more susceptible to immune checkpoint blockade regardless of their tissue of origin.

In many tumors, immune checkpoint ligands on tumor cells and immune cells interact with their cognate receptors on effector T-cells, inhibiting an adaptive immune response to the cancer cells. Once this interaction is blocked by checkpoint

inhibitors, it uncovers a subset of tumors which are highly responsive to an endogenous immune response [15]. This immunologic attack can translate into a potent and durable anti-tumor effect, but can also exert severe autoimmune adverse events [16]. How to identify and enrich tumors that respond to checkpoint inhibition is an area under intense investigation. Predictive biomarkers include PD-L1 expression, tumor mutation burden (TMB), lymphocytic infiltrates, RNA expression signature, and mutation-associated neoantigens (MANA). The latter is a biomarker that we may use across tumor types [17]. dMMR cancers are predicted to contain a large number of MANAs, and hence be recognized by the immune system [18].

In their prospective study, Le et al. used pembrolizumab in dMMR cancers across different tumor types. All patients ( $n = 86$ ) had received at least one prior therapy, had evidence of progressive disease prior to enrollment, and had dMMR assessed by either PCR or IHC [19]. Twelve different cancer types were enrolled in the study. Objective radiographic responses were noted in 53% of patients (52% of patients with colorectal cancer and 54% of patients with cancers originating in other organs), with 21% achieving a complete response (CR). Disease control was attained in 77% of the 86 patients. At the time of publication, neither median progression-free survival (PFS) nor overall survival (OS) had been reached and the study was ongoing. Eleven patients achieved a CR and were taken off therapy after two years of treatment. No evidence of cancer recurrence has been observed in these patients with a median time off therapy of 8.3 months. In terms of toxicity, treatment-related adverse events were manageable, and resembled those reported in other studies using pembrolizumab.

Other tumors are being driven by BRAF genetic abnormalities, which present another tumor site-agnostic target. Mutations in the *BRAF* gene were first identified and implicated in human cancers in 2002 [20]. Constitutively activating *BRAF* mutations have been reported in 7–15% of all human cancers, with melanoma having one of the highest incidences (40–70%) [21]. Other tumor types with a high prevalence of *BRAF*<sup>V600</sup> mutations include hairy cell leukemia, multiple myeloma, papillary thyroid cancer, histiocytic conditions (e.g., Erdheim-Chester disease and Langerhans cell histiocytosis), serous ovarian cancer, non-small cell lung cancer, and colorectal cancer [22–28]. The oncogenic *BRAF*<sup>V600</sup> driver mutation is often associated with an aggressive phenotype, and shorter disease-free survival (DFS) and OS than wild type *BRAF* [29]. We now have highly active therapies targeting this mutation in melanoma patients, such as the BRAF TKIs dabrafenib, vemurafenib, and encorafenib, and combinations of BRAF inhibitors and MEK inhibitors [30–32]. Thus, there is an increased interest in the evaluation of these treatments in solid tumors with *BRAF*<sup>V600</sup> mutations other than melanoma. The combination of dabrafenib and the

MEK inhibitor trametinib was approved for BRAF mutated NSCLC in June 2017. This approval was based on results from a multicenter, open-label trial, which sequentially enrolled 93 patients who had received previous systemic treatment for advanced NSCLC (Cohort B,  $n = 57$ ) or were treatment-naïve (Cohort C,  $n = 36$ ) [33]. The response rate was 63% for cohort B and 61% for cohort C. Interestingly, monotherapy with dabrafenib in 78 patients with previously treated BRAF mutant NSCLC, yielded a response rate of 27%. Although this is a great activity for previously treated NSCLC, the addition of a MEK inhibitor (trametinib) was necessary to achieve a significant higher response rate. Cohn et al. in 2017 published their results of a multicenter, national screening study for BRAF<sup>V600</sup> mutations, which confirmed previously reported incidences of this driver oncogene, and will allow the identification and possible enrollment of patients into the VE-BASKET study [34].

In conclusion, we can say that several actionable somatic cancer gene aberrations (NTRK, dMMR, MSI-H, BRAF) are present in several different tumor types, validating the concept of site-agnostic tumor therapy. The “organ-based” or “tumor site-based” clinical models that we have used to guide therapy before have not been adequate to predict tumor response or resistance and need to change. The new generation of oncology trials of tumor type-agnostic drugs bring hope to find new pathways of drug discovery and development.

## Compliance with Ethical Standards

**Funding** No external funding was used in the preparation of this manuscript.

**Conflict of Interest** Luis E. Raez has research support from LOXO oncology. Edgardo S. Santos declares that he has no conflicts of interest that might be relevant to the contents of this manuscript.

## References

1. <https://www.fda.gov/Drugs/InformationOnDrugs/ApprovedDrugs/ucm560040.htm> (accessed 8/26/2018).
2. Kummar S, Lassen U. TRK inhibition - a new tumor-agnostic treatment strategy. *Targ Oncol*. 2018; <https://doi.org/10.1007/s11523-018-0590-1>.
3. Brodeur GM, Minturn JE, Ho R, et al. Trk receptor expression and inhibition in neuroblastomas. *Clin Cancer Res*. 2009;15:3244–50.
4. Okamura K, Harada T, Wang S, et al. Expression of TrkB and BDNF is associated with poor prognosis in non-small cell lung cancer. *Lung Cancer*. 2012;78:100–6.
5. Drilon A, Laetsch TW, Kummar S, DuBois SG, Lassen UN, Demetri GD, et al. Efficacy of Larotrectinib in TRK fusion-positive cancers in adults and children. *N Engl J Med*. 2018;378(8):731–9.
6. Ardini E, Bosotti R, Borgia AL, De Ponti C, Somaschini A, Cammarota R, et al. The TPM3-NTRK1 rearrangement is a recurring event in colorectal carcinoma and is associated with tumor sensitivity to TRKA kinase inhibition. *Mol Oncol*. 2014;8:1495–507.
7. <https://www.clinicaltrials.gov/ct2/show/NCT02675491?cond=DS-6051b&rank=1> (accessed 8/26/2018).
8. <https://www.clinicaltrials.gov/ct2/show/NCT02048488?cond=TSR-011&rank=1> (accessed 8/26/2018).
9. <https://www.clinicaltrials.gov/ct2/show/NCT03093116?cond=TPX-0005&rank=1> (accessed 8/26/2018).
10. <https://www.clinicaltrials.gov/ct2/show/NCT03215511?cond=LOXO-195&rank=1> (accessed 8/26/2018).
11. de la Chapelle A, Hampel H. Clinical relevance of microsatellite instability in colorectal cancer. *J Clin Oncol*. 2010;28:3380–7.
12. Murphy KM, Zhang S, Geiger T, Hafez MJ, Bacher J, Berg KD, et al. Comparison of the microsatellite instability analysis system and the Bethesda panel for the determination of microsatellite instability in colorectal cancers. *J Mol Diagn*. 2006;8:305–11.
13. Zehir A, Benayed R, Shah RH, Syed A, Middha S, Kim HR, et al. Mutational landscape of metastatic cancer revealed from prospective clinical sequencing of 10,000 patients. *Nat Med*. 2017;23:703–13.
14. Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science*. 2017;357:409–13.
15. Topalian SL, Sznol M, McDermott DF, Kluger HM, Carvajal RD, Sharfman WH, et al. Survival, durable tumor remission, and long-term safety in patients with advanced melanoma receiving nivolumab. *J Clin Oncol*. 2014;32:1020–30.
16. Gettinger SN, Horn L, Gandhi L, Spigel DR, Antonia SJ, Rizvi NA, et al. Overall survival and long-term safety of nivolumab (anti-programmed death 1 antibody, BMS-936558, ONO-4538) in patients with previously treated advanced non-small-cell lung cancer. *J Clin Oncol*. 2015;33:2004–12.
17. Ward JP, Gubin MM, Schreiber RD. The role of neoantigens in naturally occurring and therapeutically induced immune responses to cancer. *Adv Immunol*. 2016;130:25–74.
18. Smyrk TC, Watson P, Kaul K, Lynch HT. Tumor-infiltrating lymphocytes are a marker for microsatellite instability in colorectal carcinoma. *Cancer*. 2001;91:2417–22.
19. Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, et al. Mismatch-repair deficiency predicts response of solid tumors to PD-1 blockade. *Science*. 2017;357(6349):409–13.
20. Davies H, Bignell GR, Cox C, et al. Mutations of the BRAF gene in human cancer. *Nature*. 2012;417(6892):949–54.
21. Dhomen N, Marais R. New insight into BRAF mutations in cancer. *Curr Opin Genet Dev*. 2007;17(1):31–9.
22. Tiacci E, Schiavoni G, Forconi F, et al. Simple genetic diagnosis of hairy cell leukemia by sensitive detection of the BRAF-V600E mutation. *Blood*. 2012;119(1):192–5.
23. Grisham RN, Iyer G, Garg K, et al. BRAF mutation is associated with early stage disease and improved outcome in patients with low-grade serous ovarian cancer. *Cancer*. 2013;119(3):548–54.
24. Chesi M, Bergsagel PL. Molecular pathogenesis of multiple myeloma: basic and clinical updates. *Int J Hematol*. 2013;97(3):313–23.
25. Xi L, Arons E, Navarro W, et al. Both variant and IGHV4-34-expressing hairy cell leukemia lack the BRAF V600E mutation. *Blood*. 2012;119(14):3330–2.
26. Marchetti A, Felicioni L, Malatesta S, et al. Clinical features and outcome of patients with non-small-cell lung cancer harboring BRAF mutations. *J Clin Oncol*. 2011;29(26):3574–9.
27. Xing M, Alzahrani AS, Carson KA, et al. Association between BRAF V600E mutation and mortality in patients with papillary thyroid cancer. *JAMA*. 2013;309(14):1493–501.
28. Heaney ML, Golde DW. Myelodysplasia. *N Engl J Med*. 1999;340(21):1649–60.
29. Roth AD, Tejpar S, Delorenzi M, et al. Prognostic role of KRAS and BRAF in stage II and III resected colon cancer: results of the

- translational study on the PETACC-3, EORTC 40993, and SAKK 60-00 trial. *J Clin Oncol.* 2010;28(3):466–74.
30. Hauschild A, Grob J-J, Demidov LV, et al. Dabrafenib in *BRAF*-mutated metastatic melanoma: a multicentre, open-label, phase 3 randomised controlled trial. *Lancet.* 2012;380(9839):358–65.
  31. Chapman PB, Hauschild A, Robert C, et al. Improved survival with vemurafenib in melanoma with *BRAF* V600E mutation. *N Engl J Med.* 2011;364(26):2507–16.
  32. Dummer R, Ascierto PA, Gogas HJ, Arance A, Mandala M, Liskay G, et al. Encorafenib plus binimetinib versus vemurafenib or encorafenib in patients with *BRAF*-mutant melanoma (COLUMBUS): a multicentre, open-label, randomised phase 3 trial. *Lancet Oncol.* 2018;19(5):603–15.
  33. Odogwu L, Mathieu L, Blumenthal G, Larkins E, Goldberg KB, Griffin N, et al. FDA approval summary: dabrafenib and trametinib for the treatment of metastatic non-small cell lung cancers harboring *BRAF* V600E mutations. *Oncologist.* 2018;23(6):740–5.
  34. Cohn AL, Day B-M, Abhyankar S, McKenna E, Riehl T, Puzanov I. *BRAF*<sup>V600</sup> mutations in solid tumors, other than metastatic melanoma and papillary thyroid cancer, or multiple myeloma: a screening study. *Onco Targets Ther.* 2017;10:965–71.