



## Clinical image: chronic skin ulcers in a patient with rheumatoid arthritis on immunosuppressant therapy

Takaaki Kobayashi<sup>1</sup> • Brian L. Swick<sup>2</sup> • Christine Cho<sup>1,3</sup>

Received: 29 May 2020 / Revised: 12 June 2020 / Accepted: 18 June 2020 / Published online: 26 June 2020  
© International League of Associations for Rheumatology (ILAR) 2020

### Presentation

A 50-year-old woman with a history of rheumatoid arthritis (RA) and interstitial lung disease (ILD) on prednisone, methotrexate, and hydroxychloroquine presented with a 3-month history of chronic left leg ulcers. Lesions were initially red patches that later blistered and ulcerated. She did not have any associated systemic symptoms. She had never received any biologic agents such as tumor necrosis factor inhibitors for RA and ILD. Empiric treatment with topical gentamicin and clobetasol did not resolve skin lesions. Physical examination demonstrated multiple ulcerated lesions with surrounding erythema in various stages of healing along the left leg (Fig. 1a). Basic metabolic panel, complete blood count, and inflammatory markers were within normal limits. Human immunodeficiency virus screen was negative. An interferon-gamma release assay was negative. Chest radiograph showed known increased interstitial lung markings due to ILD, which were stable without new infiltration. Punch biopsy of proximal lesion demonstrated numerous suppurative granulomas with long beaded acid-fast bacilli (AFB, × 1000 magnification) (Fig. 1b). AFB culture was negative, but 16s ribosomal RNA gene sequencing detected *Mycobacterium haemophilum*. She was treated with ciprofloxacin, clarithromycin, rifampin,

and intravenous amikacin for 2 months followed by ciprofloxacin, clarithromycin, and rifampin for 4 months. Due to drug-drug interaction, hydroxychloroquine was held, but the rest of immunosuppressive therapy was continued. At the end of therapy, her lesions significantly improved (Fig. 1c).

### Discussion

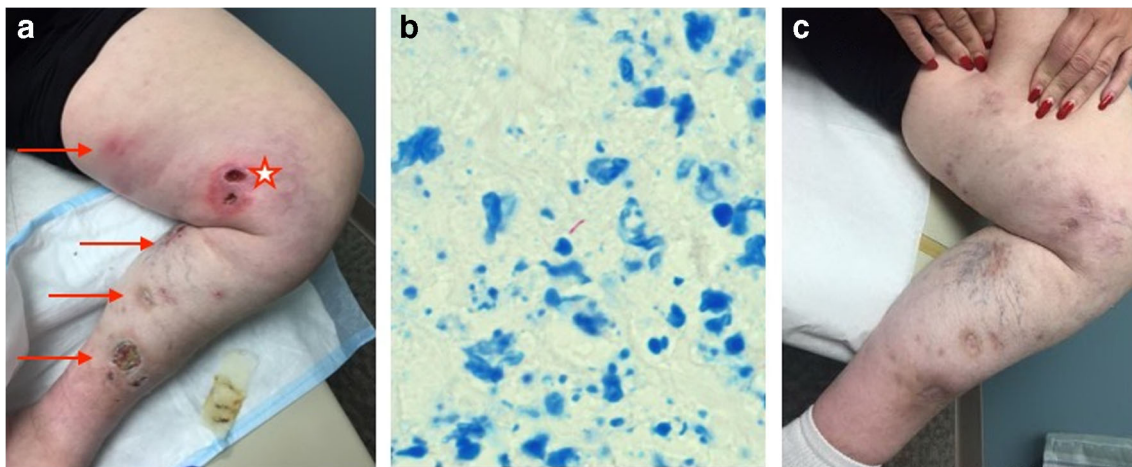
*M. haemophilum* is a slow-growing, aerobic, fastidious mycobacterium that requires heme-supplemented medium and lower incubation temperature (30–32 °C) [1]. *M. haemophilum* is presumed to be ubiquitous, but its exact habitat has not been defined [2]. Therefore, we could not identify the source of infection in our patient. *M. haemophilum* can cause a wide range of infections (cutaneous, pyomyositis, lymphadenitis, pulmonary, or disseminated infection), especially in immunocompromised patients [3]. Cutaneous disease is the most common clinical manifestation, as seen in our patient. Most strains of *M. haemophilum* demonstrate in vitro susceptibility to ciprofloxacin, clarithromycin, rifamycins, and clofazimine; variable susceptibility to doxycycline, minocycline, para-aminosalicylic acid, and amikacin; and resistance to isoniazid, ethambutol, and pyrazinamide [4]. Therapy includes combination of two to three active drugs [5]. Isolated cutaneous disease with *M. haemophilum* typically responds well to shorter duration of therapy (3–6 months), while central nervous system, musculoskeletal, or disseminated infection requires longer therapy, 12 months [4]. Relapse rate has been reported 4–14% [6].

✉ Takaaki Kobayashi  
Takaaki-kobayashi@uiowa.edu

<sup>1</sup> Department of Internal Medicine, University of Iowa Hospitals & Clinics, 200 Hawkins Drive, SW34 GH, Iowa, IA, USA

<sup>2</sup> Department of Pathology, University of Iowa Hospitals & Clinics, Iowa, IA, USA

<sup>3</sup> Iowa Inflammation Program, Roy J. and Lucille A. Carver College of Medicine, University of Iowa, and Veterans Administration Medical Center, Iowa, IA, USA



**Fig. 1** Skin manifestation of *Mycobacterium haemophilum* infection. **a** Left leg with multiple flat lesions and ulcers (arrows) with surrounding erythema in various stages of healing (prior to initiating antimicrobial therapy). Star indicates punch biopsy sites. **b** Punch biopsy of the lesion

with Ziehl-Neelsen stain positive with long beaded acid-fast bacilli ( $\times 1000$  magnification). **c** Significantly improved left leg ulcers after 6 months of antimicrobial therapy

### Compliance with ethical standards

**Disclosures** None.

**Patient consent** Obtained.

### References

- Dawson DJ, Jennis F (1980) Mycobacteria with a growth requirement for ferric ammonium citrate, identified as *Mycobacterium haemophilum*. *J Clin Microbiol* 11(2):190–192
- Saubolle MA, Kiehn TE, White MH, Rudinsky MF, Armstrong D (1996) *Mycobacterium haemophilum*: microbiology and expanding clinical and geographic spectra of disease in humans. *Clin Microbiol Rev* 9(4):435–447
- Kelley CF, Armstrong WS, Eaton ME (2011) Disseminated *Mycobacterium haemophilum* infection. *Lancet Infect Dis* 11(7):571–578
- Lindeboom JA, Buijnesteijn van Coppenraet LE, van Soolingen D, Prins JM, Kuijper EJ (2011) Clinical manifestations, diagnosis, and treatment of *Mycobacterium haemophilum* infections. *Clin Microbiol Rev* 24(4):701–717
- Griffith DE, Aksamit T, Brown-Elliott BA, Catanzaro A, Daley C, Gordin F, Holland SM, Horsburgh R, Huitt G, Iademarco MF, Iseman M, Olivier K, Ruoss S, von Reyn C, Wallace RJ Jr, Winthrop K, ATS Mycobacterial Diseases Subcommittee, American Thoracic Society, Infectious Disease Society of America (2007) An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. *Am J Respir Crit Care Med* 175(4):367–416
- Nookeu P, Angkasekwinai N, Foongladda S, Phoompong P (2019) Clinical characteristics and treatment outcomes for patients infected with *Mycobacterium haemophilum*. *Emerg Infect Dis* 25(9):1648–1652

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.