

Allergic contact dermatitis: epidemiology, molecular mechanisms, in vitro methods and regulatory aspects

Current knowledge assembled at an international workshop at BfR, Germany

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Abstract Contact allergies are complex diseases, and one of the important challenges for public health and immunology. The German ‘Federal Institute for Risk Assessment’ hosted an ‘International Workshop on Contact Dermatitis’. The scope of the workshop was to discuss new discoveries and developments in the field of contact dermatitis. This included the epidemiology and molecular biology of contact allergy, as well as the development of new in vitro methods. Furthermore, it considered regulatory aspects aiming to reduce exposure to contact

sensitisers. An estimated 15–20% of the general population suffers from contact allergy. Workplace exposure, age, sex, use of consumer products and genetic predispositions were identified as the most important risk factors. Research highlights included: advances in understanding of immune responses to contact sensitisers, the importance of autoxidation or enzyme-mediated oxidation for the activation of chemicals, the mechanisms through which hapten-protein conjugates are formed and the development of novel in vitro strategies for the identification of skin-sensitising chemicals. Dendritic cell cultures and structure-activity

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relationships are being developed to identify potential contact allergens. However, the local lymph node assay (LLNA) presently remains the validated method of choice for hazard identification and characterisation. At the workshop the use of the LLNA for regulatory purposes and for quantitative risk assessment was also discussed.

Keywords Contact allergy · Dermatitis · Epidemiology · Molecular mechanisms · Regulatory aspects

Introduction

The prevalence of contact allergy is rising worldwide [1–3]. This results in high costs for health care systems and the economy as well as in an impairment of the quality of life for the patients. The German ‘Federal Institute for Risk Assessment’ (BfR) invited national and international experts from the field of contact dermatitis for a 2-day workshop in Berlin, Germany. The workshop was organised as a part of the ‘action plan against allergies’, initiated by the German ‘Federal Ministry of Food, Agriculture and Consumer Protection’ (BMELV). The aim of the workshop was to provide a better understanding of allergic skin reactions by summarising the current state of research and to elucidate prevention strategies, as well as the requirement for any further regulation. The first day emphasised the magnitude of the problem in the general population, highlighted possible prevention strategies, and summarised known cellular and molecular mechanisms of contact allergy. The second day focused on new *in vitro* methods for assessing allergen potency and on regulatory aspects of contact allergy.

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The epidemiology of contact dermatitis: prevalence, correlations and molecular markers

Allergy incidence and prevalence

In Europe about 20% of the general population suffers from contact allergy to at least one contact allergen. Most common are allergies to nickel, fragrances and preservatives. Allergic reactions to chromate and *p*-phenylenediamine (PPD) are generally less common but occur frequently in occupationally exposed subgroups of the population [4]. Contact dermatitis occurs twice as frequently in women as in men [4] and often starts at a young age, with a prevalence of 15% in 12–16 year olds (Fig. 1) [5].

Of major concern is occupational contact dermatitis (OCD), which ranks first among occupational diseases in many countries [6]. The reported annual incident rate for OCD is 0.5–1.9‰ [7]. However, incidences are likely to be underestimated because of underreporting and lacking registration for milder cases of skin disease [8]. Moreover, the notification systems differ amongst countries, as do the criteria for compensation. In a Bavarian study, about a third of patients registered with OCD were severely professionally affected, facing either retraining or unemployment. The most affected professions (~80%) were metal workers, hairdressers, health care workers, employees in the food industry, cleaners, construction workers and painters [9].

Nickel Nickel is one of the major contact allergens worldwide. Therefore, the European Union (EU) restricted its use in consumer products in 1994 [10]. As a result nickel allergy among young patients showed a decline in several countries such as Germany [11], Sweden [12] and Denmark [13]. In Denmark the frequency of nickel allergies dropped from 26.9% before the EU directive to 12.4% thereafter [13]. However, despite the initial decline nickel sensitisation is still frequent among young women in Germany, probably due to insufficient protection [14]. Further on a significant number of people are still exposed to nickel, mainly in their working environment [15] and new sources of nickel exposure, such as mobile phones, have been reported recently [16]. Nevertheless, in the absence of legislative regulation for the use of nickel (e.g. in the US) the prevalence of nickel allergy is still increasing, especially among women [1].

Fragrances Fragrance allergy is among the most frequently detected allergies in the general population and has a prevalence ranging from 1.0–4.2% [17]. About a third of all allergies against cosmetic products are caused by fragrance allergies. Fragrances are complex mixtures

comprising altogether more than 2,000 substances, many of which are contact allergens. A mix of eight substances, fragrance mix I, has routinely been used to detect fragrance-mediated contact dermatitis. This mix has now been supplemented with a second mix of five compounds, called fragrance mix II [18]. Recent years saw a decline in allergy prevalence against fragrance mix I. Nevertheless, there remain a high number of reported cases with acute eczemas caused by fragrances [19–21]. The frequency of fragrance allergies is increased further by the autoxidative formation of allergens from commonly used fragrances [22–24].

A fragrance allergen with increasing prevalence is hydroxyisohexyl 3-cyclohexene carboxaldehyde (HICC). It is extensively used in deodorants [25] and has been linked to strong allergic reactions in users [26]. Hence HICC is now one of 26 fragrance chemicals recognised as contact allergens that is required to be labelled when present in cosmetic products and detergents (household products) [27]. This list is currently being reviewed.

Chromium Contact dermatitis against chromium (Cr^{VI}) has been a recognised problem in the occupational setting, with a prevalence of e.g. 17% in cement workers during the construction of the channel tunnel connecting continental Europe with Britain [28]. Therefore, the EU regulated the content of chromium in cement in 2005 and sensitisation to chromate in construction workers has since declined [29, 30]. However, this regulation does not include leather products such as shoes, where an increasing incidence has been recognised [31]. In Germany, though, the sale of consumer products with skin contact is banned if they release detectable amounts of Cr^{VI} (ordinance of commodities).

***p*-Phenylenediamine (PPD)** Allergic reactions caused by oxidative hair dye ingredients (e.g. PPD) are of special concern because of their severity and the widespread use of hair dyes. An estimated 0.2–2.5% of the European population and up to 20% of hairdressers suffer from hair dye allergies [32]. One risk factor for sensitisation to PPD in the general population is black henna tattoos with illegally added PPD [33–35]. Of further concern are cross-reacting substances closely related to PPD, such as the hair dye toluene-2,5-diamine (PTD) or the antioxidant isopropyl-phenyl *p*-phenylenediamine (IPPD) [36].

Risk factors

Risk factors for allergic contact dermatitis (ACD) can be subdivided into acquired and inherent. Acquired risk factors are generally inflammatory skin diseases such as irritant contact dermatitis (ICD), stasis dermatitis and possibly atopic dermatitis, while inherent risk factors are

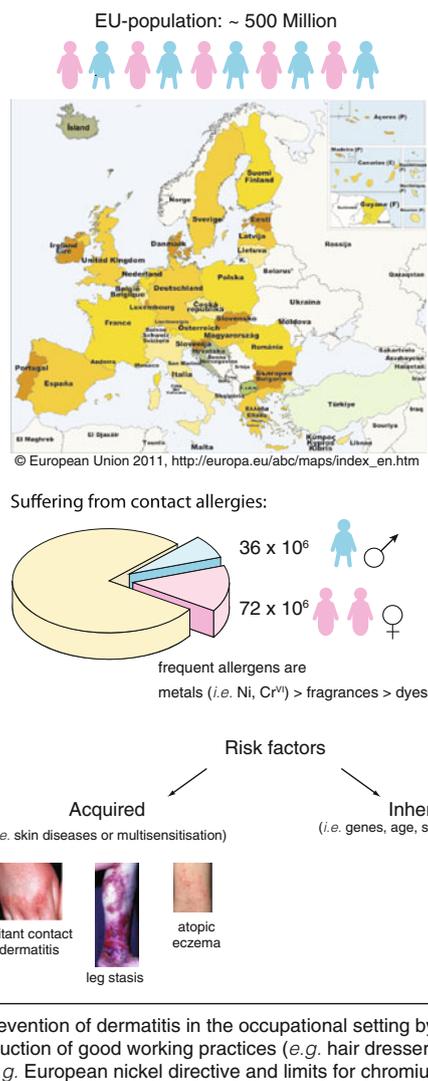


Fig. 1 Allergic contact dermatitis in the European Union, incidence and preventive measures

genetic variances resulting in a higher susceptibility (Fig. 1).

Acquired risk factors ICD often precedes ACD. Usually ICD results from a breakdown of the barrier function of the skin after exposure to skin irritants. Approximately 10% of the population suffer from ICD. The most common causes are wet work, hand washing and the wearing of occlusive rubber gloves. Atopy is a well recognised risk factor for ICD and thus possibly also for ACD [37]. However, recent studies on the gene polymorphisms of filaggrin published conflicting results regarding the question if ICD is a purely acquired risk factor [38, 39]. It seems likely that ICD has an inherent basis [40, 41].

Stasis (or leg) dermatitis is caused by venous insufficiency rather than allergens. Nevertheless, after adjusting for confounders such as sex, age and atopic dermatitis, leg

stasis could be identified to increase the risk for ACD against distinct allergens significantly [42]. Another independently acquired risk factor that was identified by regression analysis is multisensitisation. The risk of contact allergy to a specific allergen increases with the number of positive reactions in patch testing [43]. It was further shown that polysensitised individuals reveal stronger reactions to patch tests [44] and that there is an association between polysensitisation and sensitisation to weak allergens [45].

A further potential risk factor is atopic eczema, which often results in a reduced skin barrier function and therefore facilitates the penetration of toxins and allergens. However, differences in the immunological response in patients with atopic eczema frequently seems to mitigate some of the observed effects [46].

Inherent risk factors Genes, age, sex and ethnicity are the main inherent risk factors in regard to susceptibility for ACD.

Genetic risk factors are based on variations in genes (e.g. polymorphisms) involved in relevant steps for the development of contact dermatitis. Genetically influenced steps are the antigen uptake through the skin barrier, the antigen-specific response by immune cells or the metabolism of antigens by cutaneous enzymes [for a comprehensive review refer to ref. 47]. An example for the latter is the metabolism and possible activation of antigens by epidermal *N*-acetyltransferases (NATs). Studies found a relationship between the genetic polymorphism for these phase II enzymes and the risk for contact dermatitis. Patients with contact dermatitis tended to have NATs with a higher than average enzymatic activity [48, 49]. Other studies link the allele for a rapid acetylating NAT1 to a lower susceptibility for PPD sensitisation [50]. Similarly, a homogenous deletion of the glutathione *S*-transferases (GSTs) M1 and T1 showed an association with increased sensitisation against the preservative thimerosal [51]. The role of GSTs was confirmed in another study showing an elevated risk for chromate sensitisation in cement workers with a GST-T1 null phenotype [52]. Cytokine gene polymorphisms represent possible genetic risk factors at the level of an immunologic response [53]. Mutation of the promoter for tumour necrosis factor α at position 308 is associated with a higher susceptibility for chromate sensitisation in cement workers [52]. Likewise, the homozygous allele interleukin (IL) 16^{-295C} is found more frequently in polysensitised individuals with ACD [54]. Other gene polymorphisms increasing the risk for ACD have been observed in coding regions of enzymes, i.e. angiotensin-converting enzyme [55]. Induction and elicitation of contact dermatitis decline with increasing age [due reduced immune functions, see ref. 56], whereas the frequency of sensitisation increases [42].

The sex prevalence of ACD in the German population is reported to be 8% in men and 21% in women [57]. The more than two-fold higher prevalence in women is due to different exposures, such as nickel through piercing [13, 58]. However, even if nickel is not considered, women still have a higher prevalence for ACD. This higher susceptibility is probably caused by hormonal influences [59, 60]. Boys on the other hand show a higher prevalence for allergic skin reactions against fragrances [61].

Studies investigating the relation between ACD and ethnicity are inconclusive in regard to ethnicity as an inherent risk factor [e.g. ref. 62]. Some reports implicate darker skin to have a higher barrier function for some substances, thus lowering the respective risk for ACD [63, 64].

Risk factors in the occupational setting Atopic skin diathesis (ASD) was recognised as a major risk factor for OCD [65]. Data analysed from a registry of occupational skin diseases in Bavaria showed that 37% of patients with OCD also suffered from ASD [66]. Cohort studies subsequently identified several professional risk factors for the development of OCD. In the car industry the most important risk factors were ASD, a history of hand eczema and more than 3 h wet work per day [67]. Likewise hairdressers were significantly affected, with 59% developing hand eczema during their first year of apprenticeship. Again, ASD and wet work (4 h) were identified as the statistically most significant risk factors ($p < 0.001$, T.L.D., personal communication).

Similar results come from studies with nurses, where ASD together with hand washing and disinfection was a significant risk factor ($p < 0.001$) for the development of hand eczema (T.L.D., personal communication). This is in line with the findings of studies looking into the causes of a recent epidemic of ICD in health care workers in the UK's national health system (NHS). Health care workers in the UK wash their hands up to 60 times a day as a result of the NHS' 'Clean Hands Campaign', which was introduced to reduce microbial cross infections, especially with multi-drug resistant *Staphylococcus aureus*. As a result there has been a two-fold increase in the amount of hand cleansing soap purchased by the NHS in the past 4 years, while the amount of alcohol gel purchased is a tenth of what health care providers in continental Europe tend to use. In order to lower the risk for ICD, educational hand cleansing campaigns should thus emphasise the need to use alcohol gels rather than hand washing (J.E., personal communication).

Prevention

Surveillance is the basis of prevention [68], and therefore population-based investigations in consumer and

occupational settings are important. However, they cannot provide a tool for continuous monitoring. They have to be complemented by clinical epidemiological data, which indeed are collected in many countries. This has previously been the basis for successful interventions such as the nickel regulation [10] or the ban of the preservative methylidibromo glutaronitrile (MDGN) [69].

Prevention of OCD Occupational contact dermatitis frequently occurs because of lack of awareness of dermatitis hazards, complacency and poor working practices. There are various approaches to prevent OCD. Generally speaking ‘general primary prevention’, that is the elimination, replacement or reduction of allergenic substances, is by far the most effective approach [e.g. 36, 70]. Alternatively, exposure to harmful substances could be minimised by avoiding their release from the corresponding product, e.g. by encapsulation. Other preventive measures include the reorganisation of work, for example reducing the hours of wet work in order to minimise the risk for hand eczema (Fig. 1).

Efforts to minimise risk factors for OCD should be complemented by the use of personal protective equipment and pre-employment screening. In Germany, prevention measures are described in the “Approved Code of Practice” (TRGS) regulations. For hairdressers these include the replacement of harmful substances and instructions on the use of personal protective equipment (e.g. gloves). As a result the annual incidence of OCD in hairdressers dropped significantly [70, 71]. Skin care management should include preventive skin protection as well as skin care after hand cleansing [72]. Recommendations for the minimisation of work-related hand eczema include the washing of the hands with lukewarm water, the use of appropriate gloves for the shortest possible time, the removal of hand jewellery prior to work, the wearing of cotton liners when possible and the avoidance of disinfectant hand cleansers. Creams should be applied after work and at home [73].

Molecular mechanisms of chemically induced contact dermatitis

Innate immune mechanisms involved in contact dermatitis

Dendritic cells (DCs) and the local tissue microenvironment are crucial factors in the development of ACD. Within the immune system DCs are the cell type that primes naïve T cells and thus forms a crucial link between the innate and adaptive immune system.

The precise role of DCs in ACD is still under investigation; especially the contributions of the respective cellular pools are still disputed. In the current model Langerhans cells (LCs), as epidermal DCs, and dermal DCs

are centrally involved in the sensitisation and the elicitation phases of ACD (Fig. 2). During sensitisation, DCs react to potentially allergic chemicals by interaction with neighbouring keratinocytes, migration to the local draining lymph nodes and the priming of naïve T cells. These reactions are mediated by inflammatory cytokines, chemokines and adhesion molecules [74]. Allergen-specific effector T cells are then recruited into the skin upon contact with the same allergen (elicitation). Following their recruitment these T cells are activated by allergen-presenting skin cells, including LCs, dermal DCs and most likely keratinocytes [75]. Cytotoxic effector T cells in the epidermal-dermal border then deliver an inflammatory ‘lethal hit’ killing, amongst others, keratinocytes at the suprabasal layer [76]. The following activation of further skin-specific effector cells, i.e. cytotoxic T (CD8⁺ Tc1) cells and T helper (Th) cells 1 and 17, results from the interaction of DCs, keratinocytes and the loss of regulatory T (T_{reg}) cell-mediated inhibition [77, 78].

The priming of naïve T cells in skin is the result of a molecular signal cascade originating from skin DCs. The latter present antigenic peptides or allergens on their major histocompatibility complex molecules (MHC) for recognition by antigen specific T cell receptors (TCRs). Concomitantly there is co-stimulation of the T cell population, e.g. by the interaction of DC-expressed CD80/CD86 and CD28 on T cells. Cytokines excreted by the DCs (e.g. IL-12, IL-6) polarise the T cell subset differentiation. This leads to the loss of regulatory T cell inhibition, as well as the generation of Th cells 1, 2, 3 and 17, CD8⁺ Tc1 cells and most likely other subsets. Finally the expression of T cell homing receptors (e.g. E-selectin ligand, CCR4 and CCR10) is induced by migratory DCs in the draining lymph nodes. Primed T cells will subsequently home into the tissue of origin of the corresponding DCs, e.g. the skin in case of dermal DCs and LCs [79, 80]. Skin DCs acquire their potential to imprint tissue-specific homing receptors in CD8⁺ Tc1 cells by interaction with stromal and epithelial cells. In cocultures of DCs with dermal fibroblasts, DCs are prompted to imprint skin homing receptors on T cells [81].

In humans ACD has been associated with defective T_{reg} cells [82, 83] and indeed it has become clear that T_{reg} cells influence sensitisation as well as elicitation. Originally T_{reg} cells were defined as CD4⁺CD25⁺-T cells and were mainly associated with self-tolerance [84, 85]. We now know that this definition comprises a heterogeneous cell population that includes natural T_{reg} and inducible T_{reg} cells under the transcriptional control of Foxp3 as well as Tr1- and Th3-cells [reviewed in refs. 86, 87]. The skin contains predominantly inducible T_{reg} cells, which can be triggered by LCs as well as dermal DCs [88, 89]. However, the precise phenotypes of T_{reg} cells involved in ACD are

still not known [87]. Recently the induction of contact allergen-specific CD4⁺CD25⁺ICOS⁺ T_{reg} cells during sensitisation was shown to be an important regulator of CD8⁺ effector T cell responses in contact hypersensitivity [90]. Following exposure to a contact allergen T_{reg} cells can lower or even completely suppress the process of sensitisation [89, 91–93]. During subsequent elicitation they can further downregulate the immune response (i.e. by CD39) and influence the influx of leukocytes (mediated by IL-10) [94, 95]. Finally, T_{reg} cells are involved in the control and eventually termination of the inflammatory response [96].

The innate stress and immune response preceding the induction of skin homing T cells is triggered by several complex interactions of contact allergens with the skin and partly resembles the innate immune response to pathogens. This involves the triggering of Toll-like receptors, induction of reactive oxygen species and activation of the NLRP3 inflammasome [97]. As a cytosolic complex the latter consists of the innate immune receptor NLRP3, the adaptor protein ASC and pro-caspase-1. Its activation is the result of allergen-induced ATP-release from skin cells and triggering the ATP receptor P2X₇ [98–100]. Subsequently this leads to release of active caspase-1, which processes contact allergen-induced pro-IL-1 β and pro-IL-18 to the mature and secreted cytokines. These cytokines are involved in the inflammation of the skin and trigger migration of DCs, thus mediating a ‘danger’ signal function for contact sensitisers [101, 102]. Knock out studies in mice further confirm this signal function for contact sensitisers. Recent studies show that Toll-like receptor (TLR) deletion mutants (Δ TLR 2/ Δ TLR 4 or, alternatively, Δ TLR 2 or 4/ Δ IL-12R β 2) can not be sensitised to 2,4,6-trinitro-1-chlorobenzene (TNCB) and other contact allergens. In the absence of TLR2- or TLR4-mediated signalling DCs are only partially activated by contact allergens, upregulating co-stimulatory molecules but no pro-inflammatory cytokines [103]. While TNCB activates TLR2 and TLR4 indirectly through the induction of endogenous TLR2/4 ligands, nickel can trigger TLR4 signalling directly. Nickel ions can interact with histidine residues in human TLR4. Interestingly, these histidine residues are missing in the mouse TLR4. This explains why mice do not develop contact hypersensitivity to nickel unless LPS is co-injected [104]. These new findings show a physiological role for TLRs in the induction of contact hypersensitivity.

Furthermore, contact allergens can induce the cytoprotective phase II response. The phase II response is activated by the binding of electrophilic contact allergens to the cysteine-rich sensor Kelch-like ECH-associated protein (Keap1). This results in the release and nuclear translocation of the transcription factor Nrf2, thereby leading to the

activation of genes that contain antioxidant response elements [105]. Thus the Keap/Nrf2 pathway modulates inflammation and other responses of the cell. Contact sensitisers like 1-chloro-2,4-dinitrobenzene, *p*-phenylenediamine and NiSO₄ are potent inducers of the Keap/Nrf2 pathway in DCs, suggesting Nrf2 may be a new biomarker for the sensitisation potential of chemicals [106].

The detailed understanding of the induction of innate stress and immune responses by contact allergens will help to develop new therapeutic strategies to treat ACD, to identify potential contact allergens and to discriminate them from irritants. Another goal is to develop in vitro assays for hazard identification and risk assessment in order to replace animal testing.

The skin immune system and contact dermatitis: role of keratinocytes and NKT cells

Until recently the classical paradigm stated LCs to be the primary antigen presenting cells for T cell responses in skin. The importance of LCs being undisputed, this statement had to be revised to include the aforementioned DCs and, most likely, keratinocytes. Meanwhile there has been increasing evidence that LCs might also control the T cell response by secretion of anti-inflammatory cytokines (IL-10 and TGF- β) and the induction of regulatory T cells [88, 107, 108]. Accordingly, following treatment with oxazolone or 2,4-dinitro-1-fluorobenzene (DNFB) mice with ablated LCs show increased ear swelling [109].

In this context keratinocytes may as well play an important role in downregulating the skin’s immune responsiveness in steady state and may interfere with the induction of contact hypersensitivity. Only low levels of the inflammation-stimulating ligand B7 are expressed by keratinocytes during steady state. However, elevated levels of B7-1 (CD80) were found in keratinocytes transfected with B7-1 reporter constructs after exposure to allergens like oxazolone, *Myroxylon perireae* (Balsam of Peru) or nickel chloride [110]. Keratinocytes from transgenic mice (B7-1 and B7-2) delivered costimulating signals for skin inflammation during ACD. After treatment with TNCB or DNFB, B7-1 transgenic mice showed pronounced ear swelling and elevated levels of the cytokines IL-6, TNF- α and LT- β . Furthermore, the expression of IL-10 and the hapten specific IgE is seen in B7-1 transgenic mice but absent in B7-2 (CD86) or wild-type (*wt*) mice. This inflammatory response is typical for Th1 reactions in chronic dermatitis [111]. The elicitation of ear swelling by the hapten-specific IgE indicates a deviated Th2-mediated immune response, even in the presence of Th1 specific cytokines [111, 112]. In *wt* mice keratinocytes thus seem to modulate T cell-mediated inflammation and induce immune tolerance by T cell anergy.

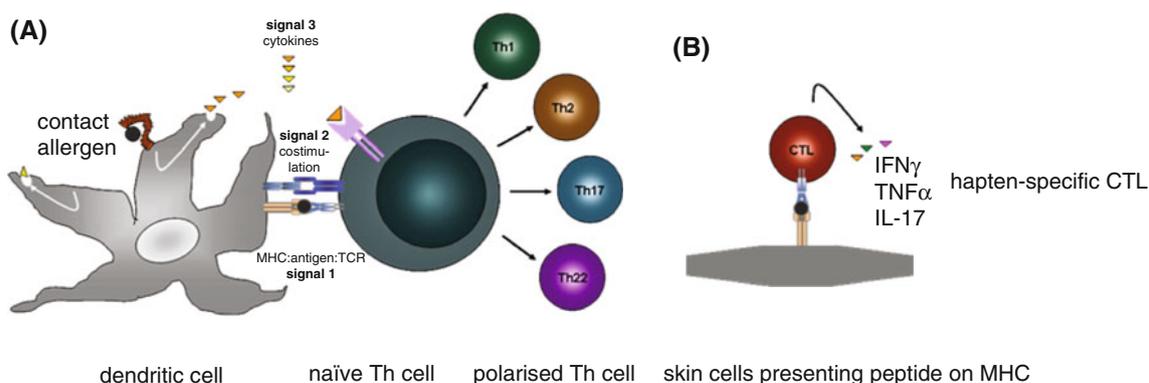


Fig. 2 Lymphocyte-mediated immune mechanisms in contact allergy. Sensitisation phase (a). The contact allergen activates dendritic cells in the skin via ‘pattern recognition receptors’ such as TLRs. Subsequently naïve T helper (Th) cells are polarised upon specific recognition of the haptened allergen by the major

histocompatibility complex (MHC), costimulatory signals and cytokines such as IL-12, IL-4, IL-1 β and IL-6. Elicitation phase (b). Haptenspecific cytotoxic CD8⁺ T lymphocytes (CTLs) release inflammatory cytokines and induce disease-specific local skin lesions following re-exposure of the skin to the same contact allergen

Further, keratinocytes are most likely the main source of IL-33 in the epidermis. This proinflammatory member of the IL-1 family is found in high concentrations in barrier tissues [113–115]. Located in the nuclear compartment of keratinocytes IL-33 is discussed to be an alarmin released by necrotic cells and was found to exacerbate contact sensitivity [113, 116]. The precise function of IL-33 is unknown. However, IL-33 is known to activate the ST2-1RAcP receptor complex, thus triggering activation of NF- κ B [117, 118]. Further on IL-33 was found to enhance IL-5 production by Th2 cells and to induce the proliferation of B1 lymphocytes independent of IL-5. The respective studies indicate both effects to be ST2-mediated [116, 119]. The IgM secreted by B1 cells is not only reactive to a broad range of antigens, but also is required for the initiation of antigen specific T-cell migration [74, 116]. B1 B cells have been shown to participate in the early initiation phase of contact hypersensitivity by IgM-mediated facilitation of effector T cell recruitment for elicitation. Sensitisation with a contact allergen leads to rapid activation of natural killer T lymphocytes (NKT cells) in the liver. These then activate peritoneal B1 B cells via IL4, recruiting them to lymphoid organs [120–122].

Natural killer T lymphocytes are known to be crucial for autoimmune diseases, allograft rejection, anti-tumor immune responses and anti-microbial immunity. It remains to be determined if they contribute to the molecular mechanisms of ACD as well. In humans these lymphocytes recognise glycolipid antigens using a TCR composed of V β 24-J α 15 and V α 11. Recognised antigens can be either self-antigens or microbial antigens and are presented by CD1d1. It was previously shown that terminally differentiated keratinocytes from human skin increase surface expression of CD1d [123]. A recent study examined skin biopsies from positively patch-tested patients using

quantitative RT-PCR and immune histochemistry. Expression of CD1d and CD161 (NKR-P1A) was found to be upregulated in epidermal cells. The NKT frequency was determined by immunological detection of the V α 24 TCR chain. Following exposure to PPD, NiSO₄ and epoxy resin NKT frequencies in ACD lesions were 2, 4 and 33% respectively. In addition, the expression of the NKT cytokines IFN- γ and IL-4 was upregulated. The occurrence of NKT in blood was constantly below 0.1%, confirming the observed effects to be specific for the ACD lesions [124]. These data support the notion that keratinocytes actively influence T cell mediated allergic immune responses in the skin.

Cytochrome P450 in keratinocytes and antigen presenting cells

Numerous isoforms of cytochrome P450-dependent monooxygenases (CYPs) as well as various transport systems are expressed within the skin. The respective cells comprise skin cells and include keratinocytes as the major compartment of the epidermis as well as antigen-presenting cells such as monocytes or dendritic cells [125]. CYPs are heme-containing enzymes catalysing the oxidative conversion of a range of predominantly lipophilic molecules into species that are generally more reactive and/or hydrophilic (water soluble), thus facilitating phase II metabolism and subsequent excretion from the body. Keratinocytes express multiple enzymes belonging to CYP families 1, 2 and 3, which are well known to metabolise xenobiotics. CYPs 1A1, 1B1, 2B6, 2E1, 3A5 and 4B1 were found to be constitutively expressed in skin-derived keratinocytes. In addition, expression of CYP3A4 was induced by dexamethasone and levels of CYP1A1 were elevated following induction with benzantracene [126].

Exon arrays show different expression patterns for CYPs between skin and liver cells as well as for keratinocytes and monocyte-derived DCs. Especially CYP1B1 and CYP27A1 were primarily expressed in DCs [125].

Several studies looked at the transport systems of keratinocytes and antigen-presenting cells because of the inherent linkage between CYP catalysed metabolism and cell transport. Keratinocytes and liver cells show similar expression patterns for their efflux transport systems while differing in the expression of influx systems [127]. However, keratinocytes and antigen-presenting cells exhibit a similar pattern of expression for their influx proteins and differ only slightly regarding their efflux proteins. Both cell types express organic anion-transporting polypeptides B, D and E, the corresponding ATP-binding cassette C transporters and the multi-drug resistance-associated proteins 1, 3, 4, 5 and 6 [128]. The inhibition of transport proteins influences the phenotype of DCs and possibly the differentiation of keratinocytes [129]. Furthermore transport efficiency can influence allergen exposure and thus sensitisation. The delayed efflux of eugenol metabolites leads to an increased IL-8 expression because of high internal eugenol concentrations [130]. Other substances require CYP-dependent activation in order to become allergens and thus provide conflicting data *in vivo* and *in silico*. For carboxime in particular uptake and subsequent CYP-dependent metabolism are regarded as prerequisite for skin sensitisation [131].

Common chemicals form contact allergens by autoxidation

Contact dermatitis caused by low molecular weight compounds requires the formation of antigenic hapten-protein complexes. The potential of a low molecular weight compound to become a hapten is thus determined by its chemical reactivity towards skin proteins. Some compounds will react directly (e.g. nickel), while others require activation, either metabolically inside the skin or externally [132]. The latter are classified either as pro- or prohaptens, depending on the mode of activation. Non-sensitising compounds that require metabolic activation are prohaptens, while prohaptens are compounds with no or low sensitising potential that are activated externally [133].

Examples for prohaptens are found among the unsaturated hydrocarbons and ethers such as common fragrance terpenes, diterpenes in colophony and ethoxylated surfactants. Patch tests revealed some of these substances to be potent skin sensitisers following their activation by autoxidation. Autoxidation of limonene (from citrus) and linalool (from lavender), two frequently used fragrances, results in the formation of the corresponding hydroperoxides [134–136]. Multicentre studies imply that oxidised limonene and oxidised linalool are among the most

common causes for ACD, while the compounds themselves rarely cause sensitisation [22–24, 137, 138].

Prohaptens are metabolically activated in the skin and thus activation could vary depending on the individuals' enzymatic expression patterns. Well-known examples of prohaptens are cinnamyl alcohol (3-phenyl-2-propen-1-ol) and urushiols [139, 140]. Some compounds are prehaptens as well as prohaptens. Depending on the way of activation the resulting haptens can have different potentials for skin sensitisation. A well-studied example is the moderate sensitizer geraniol, which is used in the basic fragrance mix for the diagnosis of contact allergy. Studies showed it to act as a prehapten that is activated by autoxidation, as well as being a hapten when activated by CYPs. The two major haptens formed by both processes are geranial and neral. However, autoxidation results in the additional formation of a sensitising hydroperoxide, while enzymatic activation produces sensitising epoxides and aldehydes [141, 142].

Considering the importance of oxidation for the formation of haptens autoxidation and CYP-mediated metabolism should be part of the hazard identification for potential contact allergens. This can be achieved by predicting autoxidation using structure activity relationships (SAR) and by *in vitro* CYP activity assays. A recently developed CYP cocktail is based on cutaneous CYP enzymes and thus allows studying part of the skin metabolism *in vitro* [143]. Furthermore, diagnosis of contact allergens should include patch tests with oxidised forms of the corresponding substances.

Contact sensitisation: hazard identification, assessment of potency and opportunities for the development of alternative methods

In the past decades hundreds of chemicals have been implicated as contact allergens. Hence there is a need to identify potential skin sensitizers. Most approaches are based upon an appreciation of the cellular and molecular mechanisms involved in the acquisition of skin sensitization. LCs are of particular interest as they are now known to play important roles in both the initiation and orchestration of cutaneous immune responses to chemical allergens. The activation, mobilisation, migration and subsequent presentation of antigen in regional lymph nodes results in a clonal expansion of allergen-responsive T lymphocytes and the development of sensitization [87, 144–148].

The activation and proliferation of T lymphocytes in skin-draining lymph nodes during skin sensitization can be measured using the local lymph node assay (LLNA) [149]. In this assay cell turnover is measured as a function of the incorporation of ³H-thymidine. On this basis chemicals that elicit a three-fold or greater increase in T lymphocyte proliferation are categorised as skin sensitizers. The LLNA

provides a reliable test for the identification of skin-sensitising chemicals and has been added to the OECD testing guidelines [guideline 429, see ref. 150]. In addition, the LLNA can be used for evaluation of the relative skin-sensitising potency of contact allergens, and this in turn provides a sound basis for the development of accurate quantitative risk assessments. For this latter purpose EC3 values are derived from consideration of dose responses in the assay [151, 152].

The LLNA is the most useful tool for the identification and characterisation of skin-sensitising substances. However, there is a lack of comparable *in vitro* methods, and substantial efforts have been, and are being, made worldwide to develop alternative assays. The main challenge is to address the required level of integration of the molecular and cellular processes that are underlying skin sensitisation. The development of contact allergy follows a four-step process as the allergen has to (1) achieve epidermal bio-availability, (2) stimulate a local trauma that leads to cytokine expression, (3) form a hapten–protein conjugate and (4) be inherently immunogenic to induce the activation of T lymphocytes. It will probably take several separate *in vitro* assays to achieve a sufficient level of experimental integration for these steps (I.K., personal communication).

In vitro methods as alternatives to animal testing in predicting and characterising the allergenic potency of chemicals

Characterising allergenic hazards and assessing allergy risks: defining the role of alternatives

Adequate *in vitro* assays should allow the assessment of allergen potency, a requirement introduced in 2009 by the ‘Globally Harmonised System’ [153]. In the absence of an agreed standard data set, this will prove to be challenging as the dose metrics employed *in vitro* need extrapolation to match the *in vivo* situation.

Effective risk management requires quantitative risk assessment and thus information about allergen potency, i.e. EC3 values from a LLNA. Other test alternatives to the LLNA are the Magnusson and Kligman maximisation test and the occluded patch test of Buehler, which use guinea pigs as test system [154, 155]. These tests predict skin-sensitising substances (EU-label R43) with 85–90% accuracy, although the LLNA is the only test formally validated. Decisions on the classification of allergens follow a weight of evidence approach. They are primarily based on the *in vivo* test results, but include the chemical structure as well as clinical data in order to reduce the number of false positives and false negatives [156]. This strategy has been successful in identifying major skin

sensitisers. However, it fails to spot potential allergens that fail to generate a test response of sufficient magnitude.

Skin sensitisers: chemical reactivity as a tool for hazard and potency prediction

Chemical reactivity has been seen as a key parameter for allergenic sensitisation since it was first discussed in the 1930s [157]. ‘Quantitative Structure Activities Relationships’ (QSAR) can be used to evaluate the sensitising potential of allergens based on physicochemical parameters like chemical and thermodynamic constants, reactivity and the partition coefficient. Predictions tend to be accurate for molecules that share the same reaction mechanism. However, QSAR can be difficult for substance classes that have more than one option on how to react with the reaction partner, e.g. aldehydes [158, 159]. Together with nucleophilic proteins saturated aldehydes form Schiff bases while α,β -unsaturated aldehydes also have the option of undergoing a Michael addition reaction [160]. In the latter case QSAR-predicted EC3 values can differ significantly from the ones obtained *in vivo* [161].

Peptide assays can be used to look into the reactivity of potential allergens in more detail. The reactivity of potential chemical allergens was compared using glutathione and synthetic peptides containing lysine, histidine or cysteine [162]. The assay showed the highest sensitivity with cysteine as functional group, while histidine was the least sensitive. Notably the assay identified some substances of low and moderate protein reactivity, which are known to be negative in the LLNA, i.e. 2-hydroxypropyl methacrylate, 1-bromobutane, 2-acetylcyclohexanone, propylparaben or vanillin. However, the assay inherently failed to detect prohaptens such as aminophenol or 3,4-dihydrocoumarin. While most skin sensitisers reacted with cysteine, some were found to react with lysine and others with amino groups in general. Substances binding to any polypeptide correlated well with known potent sensitisers. The best results were achieved with a prediction system based on reactivity thresholds towards the functional groups of cysteine and lysine [163].

In vitro identification of allergens

The migration of LCs is a key process during contact sensitisation and can be used for alternative testing strategies. LC migration in human skin biopsies has been successfully used to distinguish allergens from irritants and to assess allergen potency [164, 165]. However, as an *in vivo* method it is laborious and unsuitable for high throughput screenings because of the limited availability of suitable human skin.

One alternative is the DC culture. Peripheral blood monocytes can be differentiated to monocyte-derived dendritic cells (MoDCs) by adding granulocyte macrophage colony-stimulating factor and IL-4 to the culture medium [166]. Effects of allergens added to the maturing MoDCs can be followed by measuring the expression levels of the dendritic cell maturation marker CD83, the co-stimulatory molecule CD86 and the chemokine CXCL8 [167]. Exposure to allergens like NiSO₄, CrCl₃, CuSO₄ and DNCB lead to elevated levels for all three markers. The expression of CXCL8 was increased by seven out of eight allergens, but was not affected by the addition of irritants, i.e. DMSO, SDS or 1-propanol. This shows that the system can be used to distinguish allergens from irritants. Other increased markers were identified recently and include IL-8, TRIM16 and AKR1C2 [168]. Furthermore, systems based on DCs are a suitable tool to identify potential contact allergens. Transcriptomic profiling of CD34⁺ DCs following allergen exposure resulted in the identification of 13 genes, most prominently CREM and CCR. Within the initial set of 21 substances this marker set detected skin sensitisers with a concordance of 89% and a specificity of 97% [169]. Intriguingly gene expression levels seem partially to correlate with sensitising potency, thus allowing a preliminary classification of the test results [170]. Another study recently identified a biomarker signature of 200 genes in MUTZ-3 cells, following a 24 h-treatment with 20 sensitisers and 20 non-sensitizers respectively [171]. In a comparative study MoDCs and DCs from CD34⁺ precursors recognised 76 and 67% of all tested contact allergens. In addition, using the leukaemic THP-1 cell line, the histiocytic lymphoma U-937 cell line and the acute myeloid leukaemia MUTZ-3 cell line, 70, 83 and 100% of contact allergens, respectively, could be identified (T.R., unpublished). Two of these test systems are currently pre-validated for regulatory purposes, namely the myeloid 'U-937 Skin Sensitisation Test' (MUSST) [172, 173] and the THP-1 based 'human Cell Line Activation Test' (hCLAT) [174, 175]. Both test systems use the expression of CD86 as readout for dendritic cell activation, supplemented by the adhesion molecule CD54 for hCLAT [176]. Comparative studies recently highlighted the use of the latter system in regard to surfactants, a substance group for which the LLNA is known to report false positives [177].

The T cell polarising potential of contact allergens was investigated using oxazolone, DNCB and NiSO₄. Stimulation of MoDC cultures with the latter induced high levels of the Th1 polarising cytokines TNF- α and CXCL10 [167]. Analysis of the IL-12p70 (Th1)/IL-10 (Th2) ratio showed no effect for oxazolone, whereas DNCB enhanced a Th1 response. Notably NiSO₄ induced a Th2 response, which is the exception for contact allergens but had been suggested earlier by clinical studies [178]. As contact allergen nickel

has the ability to activate the NF- κ B pathway, which usually leads to the release of inflammatory cytokines by DCs [179]. Altogether it appears that the activation of DCs is influenced by intrinsic properties of the respective allergens. This is supported by the observation that contact and respiratory allergens tend to cause Th1- or Th2-associated diseases respectively [167, 178].

While DCs are crucial during initial sensitisation any subsequent allergic reaction is caused as a consequence of effective T cell stimulation following allergenic re-exposure. Complex as it might be, stimulation of naïve T cells is thus an effective target for any in vitro testing strategy. Naïve T cells can either polarise to cytotoxic T cells (CD3⁺ CD8⁺) or Th cells (CD3⁺ CD4⁺). The latter further specialise to Th1, Th2 and IL-17 releasing Th cells (Th17), or to regulatory T cells. Two further T cell populations, Th22 and Th9, were described recently, and their role in hypersensitivity remains to be determined [180–183]. Matters are further complicated by the fact that, to an as yet undefined extent, contact hypersensitivity is regulated by the balance of inflammatory (Th) versus inhibitory (T_{reg}) cells present at the site of antigen exposure. This balance is in turn regulated by the expression of P- and E-selectin ligands and chemokine receptors on T cells proliferating in the lymph nodes draining that site [Fuhlbrigge unpublished, 184, 185]. Current assays thus mainly focus on the priming of CD45RA⁺ T cells, using cell proliferation and production of IFN- γ or IL-5 as readouts [for a recent review please see ref. 186]. Depletion of regulatory CD25⁺- or CD4⁺-T cells in vivo increases the number of IFN- γ -producing T cells during sensitisation. Likewise the sensitivity of the corresponding in vitro assays can be increased by using systems depleted of CD25⁺ T cells [187, 188].

Another potential target for in vitro testing is Th cells. Promotion of Th17 involves several cytokines (IL-6, IL-1 β , TGF- β and IL-23) and can be driven by LCs following the stimulation of TLR 2 [189]. The involvement of TLR 2 links Th17 to dermal inflammation. Nevertheless Th17 cells seem likewise to be involved in contact allergy as IL-17 was shown to promote type-IV hypersensitivity to DNFB [190, 191]. Furthermore, NiSO₄ stimulates LCs to release IL-6, IL-1 β and IL-23 (M.P., unpublished), and IL-17 was previously found in nickel-specific T cells [192]. Allergens like cinnamal and DNCB fail to induce a Th17 phenotype in naïve T cells, suggesting a different mechanism (M.P., unpublished). Contact hypersensitivity thus involves Th cell subpopulations, other than just Th1, which could be considered for the design of future in vitro assays.

Other cells like NK cells are also implicated to be effector cells for allergic inflammatory responses of the skin [193, 194]. In T/B cell-deficient mice, NK cells can initiate allergic responses to DNFB, oxazolone and picryl

chloride. These responses are contact allergen-specific and can be recalled weeks after sensitisation. In the absence of T and B cells contact allergens thus induce memory-like responses that depend on liver NK cells expressing the chemokine receptor CXCR6. Further on NK cells also infiltrate the skin of *wt* mice [193, 194] and humans during allergic contact dermatitis [195]. In human contact dermatitis they seem to amplify the allergic reaction.

Dose response and threshold issues in chemically induced skin sensitisation and its implications in regulatory toxicology

Assessing contact allergen potency and thresholds in the local lymph node assay

Sensitisation to an allergen depends on its potency and allergenic exposure in terms of frequency, duration and site. The LLNA assesses sensitisation potency and thus allows the identification and comparison of potential allergens (Fig. 3). This was further demonstrated by ranking the sensitising potential of rubber chemicals [196]. In this study an EC3 value was estimated from a dose response curve by fitting non-linear regression models. Uncertainty was limited to a 90% confidence interval by parametric bootstrapping. The resulting 5th percentile of the EC3 value from the bootstrapping method represents the dose where an allergic reaction will occur with a 5% probability and is similar to the benchmark dose limit (BMDL). Ultimately the data of LLNA allow to set exposure thresholds, defining a sensitising dose per unit area [197].

Generally LLNA thresholds correlate well with human thresholds. A critical question is whether prolonged exposure to an allergen below its threshold can cause sensitisation. Therefore mice were subjected to an extended LLNA, exposing them to 2,4-dinitro-1-chlorobenzene, benzocaine and tetramethylthiuram disulfide below the corresponding EC3 values. After 56 days the lymph nodes showed no increased cell proliferation [198]. Similar negative results were seen with paraformaldehyde and hexamethylenetetramine. However, formaldehyde, 2-chloro-*N*-(hydroxymethyl)acetamide and quaternium-15 showed positive results in an extended LLNA [199]. The underlying mechanisms leading to sensitisation at exposure levels below the LLNA thresholds are unclear. Nevertheless the results show that the use of EC3 values as thresholds for no risk of sensitisation has to be evaluated carefully.

The LLNA as a tool to assess respiratory allergens

The LLNA has been proven to be a reliable test for the identification of dermal sensitisers. In addition it shows

positive test results for almost all known respiratory sensitisers [200]. This implies firstly that respiratory allergens could induce allergies following dermal exposure and secondly, that dermal tests can be used to identify potential respiratory allergens.

This was tested by adapting the dermal LLNA to fit respiratory exposure. BALB/c mice were exposed head/nose-only to various allergens (respiratory and dermal) during 3 consecutive days. Allergen exposure was at a constant concentration for 45, 90, 180 or 360 min/day. Three days after the last exposure cell proliferation was determined in the mandibular lymph nodes, which drain the respiratory tract [201]. The respiratory allergens trimellitic anhydride (TMA), phthalic anhydride (PA), hexamethylene diisocyanate (HDI), toluene diisocyanate (TDI) and isophorone diisocyanate (IPDI) showed a more than three-fold induction of T cell proliferation, as did the contact allergens DNCB and oxazolone. For TDI, HDI, IPDI and oxazolone proliferation values were even higher in the auricular lymph nodes. It is assumed that these substances have an increased dermal absorption due to their lipophilicity. Other substances, like the dermal allergen formaldehyde, the irritant methylsalicylate and the unclassified trimeric IPDI, were found to be negative in the respiratory LLNA. Altogether the contact allergens turned out to be as potent as the respiratory allergens in the respiratory LLNA, although the resulting potency ranking differed from that of a dermal LLNA [201]. Therefore all substances testing positively in the LLNA should be considered to be a potential respiratory allergen as well as a dermal allergen.

Risk assessment and risk management for skin-sensitising chemicals

Quantitative risk assessment for fragrance compounds Fragrances are among the most frequent sensitisers in cosmetic products. The corresponding risk assessments are based on the no-observed-effect-level (NOAEL) and a qualitative assessment of exposure (i.e. the NOAEL for substances with non-skin contact and NOAEL/10 for substances with skin contact). This categorisation into just two product groups prevents any further distinction of possible exposure or differences in the experienced dose due to e.g. different applications. Hence a recent approach suggested the use of a quantitative risk assessment (QRA) for fragrance compounds [202, 203]. The method is based on the following four steps: (1) hazard identification, (2) dose/response relationship, (3) exposure assessment and (4) risk characterisation. Hazard identification reviews experimental data and clinical data to identify the potential hazard of a fragrance substance to cause sensitisation. The dose/response relationship then

uses a weight of evidence approach to determine the ‘no expected sensitisation induction level’ (NESIL) and derives a sensitisation assessment factor (SAF) based on the most likely scenario of exposure. The latter will include (1) the inter-individual variability (i.e. age, gender, genetics), (2) matrix effects from varying product formulations (i.e. irritant or skin damaging) and (3) differences of the exposure scenario in the experimental setup (i.e. application on sensitive skin areas). Each parameter is factored between 1 and 10, the SAF being the product of all three factors. Division of the NESIL by the SAF aims to provide an ‘acceptable exposure level’ (AEL). The AEL is finally compared to the expected consumer exposure level (CEL), based on estimates about the amount of product used, the frequency of application and the duration of use. For safe products the CEL should be smaller than, or equal to, the AEL [203]. The approach was formally implemented by the ‘International Fragrance Association’s (IFRA) code of practice in 2006 and to date more than 80 standards have been evaluated using QRA. However, QRA remains a predictive model and has not been adequately assessed against clinical or epidemiological data.

Risk management for contact allergens In the recent years Europe has implemented a whole set of regulations aimed at reducing the exposure of the workforce and consumers to contact allergens. Examples are the ‘Nickel Directive’, limiting the release of nickel in contact with

skin to $0.5 \mu\text{g}/(\text{cm}^2 \text{ per week})$ [10], and the ‘Chromium Directive’, limiting Cr^{VI} to 2 ppm in the total dry weight of cement [29]. The directive on detergents requires the listing of preservatives and listed fragrances if their content in detergents and similar household products exceeds 100 ppm [204]. Detergents are thus treated as rinse-off cosmetics. Furthermore, details of the product formulation have to be released when necessary, i.e. to investigate adverse reactions. As a result of this regulation the preservative MDGN was banned from all cosmetic products in 2008. Prior to the ban cosmetic products were allowed to contain up to 0.1% MDGN, a level that was found to cause elicitation. A reassessment of MDGN failed to establish a safe level of use and thus recommended the ban of the substance [205].

A decision on the ban/restriction of PPD and other ingredients of hair dyes is still pending. The median prevalence of contact dermatitis against PPD in Europe is 2–6% [32]. The EU Commission’s Scientific Committee on Consumer Products (SCCP) assessed the skin sensitising properties of 48 hair dye substances in 2006, finding 27 of them to be skin sensitising according to the European classification R43. In conclusion the SCCP stated that products containing these substances might not be safe for consumer use [206]. Industry has suggested to introduce sensitivity self-testing as a regulatory requirement. However, this approach is problematic due to the possibility of false-negative results, the induction of skin sensitisation and ethical reasons [207]. Further, on application of hair dyes to the skin, the product is being used for in vivo diagnostic purposes and is thus outwith the legal framework for cosmetics. Discussions continue on the safety of PPD and similar ingredients in hair dyes

Likewise further regulation is needed for fragrance substances. The labelling of fragrances was first addressed in the 7th amendment of the first European cosmetics directive [208] and subsequently included in the new European cosmetics regulation [27]. As a consequence, the industry introduced the concept of the aforementioned QRA for the evaluation of fragrance substances. Undoubtedly this will be a useful approach for new substances. However, concerns remain that QRA fails to protect previously sensitised consumers and that aggregate exposures through multiple products are not considered in the basic form of the QRA. Therefore epidemiological and clinical data continue to represent a critical decision point in risk assessment and risk management [209].

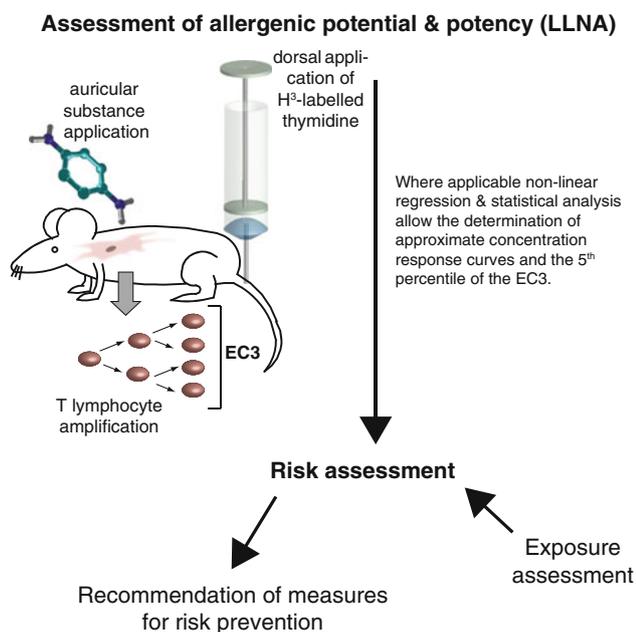


Fig. 3 Use of the LLNA for the regulatory risk assessment of potential allergens. The 5th percentile of the EC₃ is similar to a probabilistic BMDL. Alternatively the respective data might be used to estimate a threshold concentration, similar to a ‘lowest adverse effect level’ (LOAEL)

Conclusion

Skin sensitisation and subsequent contact dermatitis is a significant problem for consumers and workers. It is

clear that the immune response to contact allergens is more complex than previously thought and described. Different allergens elicit different immune responses and mechanisms for the activation of allergens can differ substantially. Research efforts are underway to elucidate the complex biochemistry and molecular biology underlying contact dermatitis. Several *in vivo* systems have been established that are able to identify potential allergens reliably and to assess their potency. At the same time *in vitro* tests are developed because of public demand to replace *in vivo* tests, animal welfare and costs. However, their regulatory acceptance will depend on a thorough validation, not only against other methods (internal validation) but also against human observational data from clinical epidemiological surveillance systems (external validation). Such validation is the indispensable gold standard for any predictive safety assessment. Legislation has to focus on the protection of consumers and workers against potential allergens, and it is adapted continuously as our understanding of allergic contact dermatitis progresses. In this context clinical data and the epicutaneous patch test as published by Jadassohn more than 100 years ago [210] are invaluable as they highlight substances and problems missed by other approaches.

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