

Symposium : Infectious diseases

Guest Editor : **Mark C. Steinhoff**, John Hopkins University, USA.

Escherichia coli that cause diarrhea

Myron M. Levine and Pablo Vial

The Division of Infectious Disease and Tropical Pediatrics, Department of Pediatrics, University of Maryland School of Medicine, Baltimore, U.S.A.

Although *Escherichia coli* plays an important role in maintaining normal gut physiology, there exist within this species primary pathogens that cause various syndromes of diarrheal disease. Five categories of diarrheagenic *E. coli* are now recognized which manifest distinct virulence properties, interact with the intestinal mucosa in different ways, cause distinct clinical syndromes, differ in their epidemiology, and fall into distinct O : H serotypes.

The five categories of diarrheagenic *E. coli* include 1. Enteropathogenic (EPEC; a frequent cause of infant diarrhea) 2. Enterotoxigenic (ETEC; major cause of traveler's diarrhea and infant diarrhea in less-developed countries) 3. Enteroinvasive (EIEC; cause dysentery) 4. Enterohemorrhagic (EHEC; cause of hemorrhagic colitis and hemolytic uremic syndrome) 5. Enteroadherent-aggregative; (EA-Aggec) common cause of infant and traveler's diarrhea).

While the five categories of diarrheagenic *E. coli* are quite distinct, they nevertheless have certain underlying commonalities from the point of view of pathogenesis.¹ These include the importance of plasmids in encoding critical virulence properties; characteristic interactions with intestinal mucosa; the production of enterotoxins or cytotoxins; a marked propensity to fall within certain O : H serotypes.

In the 1940s, Kauffman² proposed a scheme to differentiate *E. coli* on the basis of lipopolysaccharide O, flagellar H and polysaccharide K antigens. We presently recognize 171 O serogroups and 56 H types. Together these designate the O : H serotype which is important role in studying the epidemiology and pathogenesis of *E. coli* infection.

Enteropathogenic (*E. coli*)

Enteropathogenic *E. coli* was the term coined by Neter³ to refer to certain *E. coli* strains identified in the 1940s and 1950s by serological methods and incriminated as causes of infantile diarrhea both in out-

Reprint requests : Dr. Myron M. Levine, Center for Vaccine Development, 10 S. Pine Street, Room 9-34, Baltimore, Maryland 21201, U.S.A.

breaks and in sporadic cases.⁴ The major O serogroups that contain EPEC serotypes are shown in the Table. Except for O142, this list represents a compilation from reports of Ewing *et al*⁵ and Taylor⁶ who, in the 1950s, headed the enteric reference laboratories at the Center for Disease Control in Atlanta and at the Central Public Health Laboratory in Colindale, respectively; these O serogroups represent "classical" (Class I) EPEC O serogroups. O142 EPEC were described in the late 1960s and early 1970s.^{1,4} Certain other O serogroups were recognized by both Ewing *et al* and Taylor as being less strongly incriminated as pathogens; these include O18, O44, O112 and O114, among others (Table).

Volunteer studies in the early 1950s established the pathogenicity of several common classical EPEC O serogroups, including O55, O111 and O127 strains.⁴ However, by the tests and animal models available up to the mid 1960s, it was not possible to differentiate these *E. coli* from normal flora strains and the virulence properties by which they cause diarrhea remained unknown. Therefore, until the 1970s, O serogrouping remained the only diagnostic tool to detect EPEC.

EPEC strains do not elaborate heat-labile (LT) or heat-stable (ST) enterotoxins of ETEC nor do they exhibit the epithelial cell invasiveness of EIEC.^{1,4} Rather, they cause diarrhea by other mechanisms.⁷ EPEC cause a distinctive

Table. Features of the five distinct categories of *E. coli* that cause diarrhea

Category	O Serogroups	Clinical syndromes	Plasmid-mediated virulence properties
Enterotoxigenic	6, 8, 15, 20, 25, 27, 63, 78, 80, 85, 115, 128ac, 139, 148, 153, 159, 167	Infant diarrhea in less-developed countries; Adult traveler's diarrhea	LT & ST; fimbrial colonization factors
Enteroinvasive	28ac, 29, 124, 136, 143, 144, 152, 164, 167	dysentery, all ages	Shigella-like invasiveness of epithelial cells
Enteropathogenic			
Class I	55, 86, 111, 119, 125, 126, 127, 128ab, 142	Acute and protracted infant diarrhea	Attachment to epithelial cells
Class II	18, 44, 114	Acute infant diarrhea	?
Enterohemorrhagic	26, 111, 157	Hemorrhagic colitis, hemolytic-uremic syndrome, all ages	Novel colonization fimbriae
Enteroadherent-aggregative	O untypable (new O groups)	Infant diarrhea in less developed countries	O antigen and novel fimbriae

ultrastructural histopathological lesion in the intestine which involves destruction of the microvilli, typically without further invasion. Bacteria are often closely adherent to the membrane of the enterocyte with the membrane partially enveloping the bacterium.⁴ A very important practical observation (because it provides an alternative diagnostic tool to serotyping) is that EPEC adhere to HEp-2 cells in tissue culture in a characteristic pattern of microcolonies called localized adherence, a property not found among other *E. coli*.⁸⁻¹¹ The name EPEC Adherence Factor, or EAF, has been given to this property and the genes that encode it are found in certain plasmids. Localized adherence to HEp-2 and HeLa cells must be differentiated from diffuse adherence,⁹⁻¹¹ more recently, a third pattern of adherence to HEp-2 cells, the "aggregative" pattern, has been described and identifies a distinct new category of diarrheagenic *E. coli* Enteroadherent-aggregative *E. coli*²¹.

The EAF plasmid is necessary for full expression of the pathogenicity of most (but not all) EPEC strains.¹³ The EAF plasmid encodes the expression of a bacterial surface (outer membrane) protein that appears to be critical in the pathogenesis of EPEC diarrhea and perhaps in mediating protective immunity.¹³ This protein has been found in all the important EPEC serotypes, such as those in serogroups O55, O111, O119, O127, and O142, but it is not found in other pathogenic *E. coli*. An attempt to purify the 94 Kd protein is underway to prepare a potent and plentiful antibody to it that could be used in a simple diagnostic test such as one based on agglutination.

Some EPEC strains have been reported to elaborate moderate quantities of a cytotoxin very similar (or identical) to

Shigella dysenteriae I toxin. It has been suggested that this toxin may play a role in the pathogenesis of EPEC disease.^{14,15}

Clinically, EPEC illness is characterized by fever, malaise, vomiting and diarrhea with prominent amounts of mucus but without gross blood. EPEC illness tends to be clinically more severe than many other diarrheal infections in infants, some of whom develop prolonged diarrhea that persists for more than 14 days. Recent studies from several countries in South America where improved diagnostic techniques were employed have shown EPEC to be either the first or second most important bacterial cause of diarrhea in infants.^{16,17} EPEC illness is rare beyond infancy.

Unfortunately, at present, Class I EPEC can be identified only by O serogrouping, recognition of localized adherence to HEp-2 cells in tissue culture or by use of the EAF gene probe.

Enterotoxigenic *E. coli*

ETEC came to prominence in the late 1960s and early 1970s, largely based on work carried out in Calcutta.^{18,19} ETEC are a major cause of infant diarrhea in less-developed countries (some reports of infants prospectively followed by frequent household surveillance suggest that as many as 2-3 clinical ETEC infections per child/year occur during the first 2-3 years of life),²⁰ one of the main bacterial causes of dehydrating infant diarrhea in developing areas;²¹ and an infection correlated with adverse nutritional consequences.²² ETEC are also the most frequent agent responsible for traveler's diarrhea.²³⁻²⁵ Within developed countries ETEC infection is rare, although occasional outbreaks have been reported.

ETEC infection is acquired via the ingestion of contaminated food or water which allows the bacteria to reach the proximal small intestine, the critical site of host-parasite interaction. Here they colonize by means of fimbrial colonization factors and elaborate LT or ST. LT closely resembles cholera toxin in structure and action and immunologically.²⁶ ST is a small polypeptide that is not immunogenic in the course of natural infection.²⁶ The clinical features of ETEC infection are watery diarrhea, nausea, abdominal cramps and low-grade fever. Occasionally, particularly with strains that are prevalent in the Indian subcontinent, severe cholera-like purging can occur.

ETEC from diverse geographic areas fall within a limited number of O : H serotypes.¹ While many other serotypes can also be toxigenic, the recurrent O : H serotypes appear to be successful ETEC clones that have spread far and wide. Usually these serotypes elaborate both LT and ST and possess fimbrial colonization factors.¹ The major O serogroups associated with ETEC are shown in Table.

ETEC possess attachment or colonization factors that allow them to overcome the peristaltic defense mechanism of the small intestine. Heretofore, all the characterized colonization factors have proven to be fimbriae, i.e. hair-like, filamentous organelles on the surface of the *E. coli* that are notably thinner than flagellae.

ETEC are identified by detecting the presence of LT or ST or the genes that encode these toxins. Several practical tests for LT are available, including enzyme-linked immunosorbent assays (ELISAs²⁷ immunodiffusion assays,²⁸ latex particle tests, tissue culture cell lines that change their morphology in the presence

of LT (Y-1 adrenal and Chinese Hamster Ovary cells), and DNA probes.²⁹ Tests for ST are less practical and include ELISA,³⁰ the infant mouse assay³¹ and DNA probes.²⁹

Enteroinvasive *E. coli*

Certain *E. coli* strains cause an invasive, dysenteric form of diarrheal illness. These strains, of serotypes distinct from ETEC and EPEC (Table 1), were found to closely resemble *Shigella* in many ways. Like *Shigella*, their cardinal pathogenic feature is the capacity to invade and proliferate within epithelial cells, leading to cell death. The invasive capacity of both EIEC and *Shigella* is dependent on the presence of large plasmids which code for the production of several outer membrane proteins involved in the invasiveness process;¹ the proteins are antigenically closely related (if not identical) in EIEC and *Shigella*. EIEC often resemble *Shigella* in being unable to ferment lactose and non-motile. Furthermore, EIEC and *Shigella* O antigens show many cross-reactions.

EIEC have a predilection for colonic mucosa as the favored site of host parasite interaction. Clinically, the illness is marked by fever, severe abdominal cramps, malaise, toxemia, and watery diarrhea followed by gross dysentery consisting of scanty stools of blood and mucus. A simple stain of the fecal mucus reveals sheets of polymorphonuclear leukocytes.

EIEC can be diagnosed by serotyping suspect *E. coli* strains,³² by an ELISA based on detection of the invasiveness-associated outer membrane proteins³³ and by DNA probes that detect the invasiveness genes.³⁴

Enterohemorrhagic *E. coli*

In 1982 an outbreak of hemorrhagic colitis in the USA drew attention to an unusual clinical syndrome of diarrheal disease and a new bacterial enteric pathogen³⁵ the causative organism, *Escherichia coli* O157 : H7, was a serotype not previously recognized as a cause of diarrheal disease in humans. The clinical syndrome was notable in that bloody but copious diarrhea, unaccompanied by fecal leukocytes, was seen in afebrile patients;³⁵ these features distinguish it from classic dysentery due to *Shigella* or enteroinvasive *Escherichia coli* (EIEC) which are characterized by fever and scanty stools of blood and mucus containing many fecal leukocytes. Since 1982, some knowledge has been gleaned on the epidemiology of O157 : H7 infections as they occur in North America and Europe and considerable progress has been made on elucidating its pathogenesis. There has also been a strong incrimination of O157 : H7 as a cause of hemolytic-uremic syndrome (HUS).^{1,36} O157 : H7 has emerged as an enteric pathogen of public health importance in Canada and the USA with multiple reports of outbreaks of hemorrhagic colitis, hemolytic uremic syndrome and diarrhea in nursing homes, day care centers, schools, and the community.¹ So far, little is known of the epidemiology of EHEC in less-developed countries.

O157 : H7 strains from persons with hemorrhagic colitis and HUS have been shown to elaborate phage-encoded potent cytotoxins active on HeLa and Vero cells.^{1,36} One of these toxins, so-called Shiga-like toxin 1 (SLTI) or Verotoxin 1 (VT1), is apparently identical to the potent cyto-

toxin/neurotoxin/enterotoxin produced by *S. dysenteriae* 1 (Shiga toxin and reacts with and is neutralized by Shiga antitoxin.¹ Many strains also elaborate a second potent cytotoxin (Shiga-like toxin 2 or Verotoxin 2) that is not neutralized by Shiga antitoxin.¹ In addition, O157 : H7 strains possess a plasmid that plays a role in virulence by encoding the production of a newly-recognized variety of fimbriae that mediates attachment to gut-derived epithelial cells in tissue culture.³⁷

Several animal models have been developed which demonstrate the pathologic features of O157 : H7 infection.¹ In electron photomicrographs,³⁸ attached and effaced enterocytes are evident with destruction of the microvilli, a lesion resembling that due to classic serotype enteropathogenic *E. coli* (EPEC). Nevertheless, in gnotobiotic piglets, the two types of infection, BHEC versus EPEC, can be clearly differentiated by anatomic site of involvement, severity of lesions, and degree of polymorphonuclear cells infiltration. EPEC involve the entire intestine of piglets, EHEC only the cecum and colon; EPEC lesions are generally less severe; some infiltration by leukocytes is seen with EPEC, but not with EHEC, infection.

The term BHEC refers to strains such as O157 : H7 which manifest the above-mentioned clinical, epidemiologic and pathogenetic features. Heretofore, it has been difficult to undertake studies of the epidemiology of BHEC infections, other than outbreak investigations, because of the lack of suitable methods for screening large numbers of stool cultures for O157 : H7 strains and because of the lack of knowledge regarding what other serotypes may also be enterohemorrhagic

and how to identify them as well. One other serotype, in particular, O26 : H11, is now recognized as EHEC; this serotype was previously considered to be EPEC. O26 : H11 is usually an abundant producer of VT, possesses a plasmid that does not hybridize with the EPEC EAF gene probe, and sometimes is associated with bloody diarrhea. Preliminary studies have shown considerable homology among the O157 : H7 and O26 : H11 plasmids and have led to the development of a sensitive and specific DNA probe to identify EHEC.^{13,6}

Enteroadherent-aggregative *E. coli*

The newest category of diarrheagenic *E. coli* are strains that show a characteristic "aggregative" pattern of adherence to HEp-2 cells. In this pattern, the bacteria form aggregates that give a "stacked brick" appearance.¹² Bacterial clusters are visible on the glass, as well as attached to HEp-2 cells. Enteroadherent aggregative *E. coli* (Auto EC) do not elaborate LT or ST, do not manifest Shigella-like invasiveness, do not cause the histopathological lesion of EPEC and are negative with the EPEC, EPEC, EIEC and ETEC DNA probes. EA-AggEC cause a distinct histopathological lesion of the intestine discernable by light microscopy.^{3,9} AutoEC occur in distinct O : H serotypes but the O antigens are new and as yet untypable. Plasmids encode critical virulence properties.

EA-AggEC have been found to be an important cause of acute diarrhea in young children in South America and are suspected to cause protracted diarrhea in young children. At least one of the "enteroadherent" *E. coli* strains fed to volunteers by Mathewson et al⁴⁰ is now known to

be EA-AggEC. It is likely that many of the "enteroadherent" *E. coli* referred to by Mathewson et al are in fact EA-AggEC strains.

While EA-AggEC can be readily identified by their characteristic pattern of adherence to HEp-2 cells, antisera and DNA probes are also being prepared to facilitate epidemiologic studies.

From the early days of the 1940s when *E. coli* were first convincingly associated with human diarrhea, there has come to exist an impressive fund of knowledge on the several categories of diarrheagenic *E. coli*, including information on their clinical features, epidemiology, O : H serotypes and, their pathogenesis. From a previous state of some confusion about their role as enteric pathogens, diarrheagenic *E. coli* are now recognized as being among the best understood bacterial enteropathogens.

References

1. Levine MM. *Escherichia coli* that cause diarrhea : enterotoxigenic, enteropathogenic, enteroinvasive, enterohemorrhagic, and enteroadherent. *J Infect Dis* 1987; **155** : 377-389
2. Kauffmann F. The serology of the coli group. *J Immunol* 1947; **57** : 71-100
3. Neter E, Westphal O, Luderitz O, Needell MH. Demonstration of antibodies against enteropathogenic *Escherichia coli* in sera of children of various ages. *Pediatrics* 1955; **16** : 801-7
4. Levine MM, Edelman R. Enteropathogenic *Escherichia coli* of classical serotypes associated with infant diarrhea-epidemiology and pathogenesis. *Epidemiol Rev* 1984; **6** : 31-51
5. Ewing WH, Davis BR, Montague TS. Studies on the occurrence of *Escherichia coli* serotype associated with diarrheal disease. Atlanta, GA : US Department of Health, Education and Welfare, Public Health Service, Communicable Disease Centre, 1963
6. Taylor J. Host specificity and enteropathogenicity of *Escherichia coli*. *J Appl Bacteriol* 1961; **3** : 316-25

7. Levine MM, Bergguist EJ, Nalin DR, Waterman DH, Hornick RB, Young CR, Sotman S, Rowe B. *Escherichia coli* strains that cause diarrhea but do not produce heat-labile or heat-stable enterotoxins and are non-invasive. *Lancet* 1978; **1** : 1119-22.
8. Cravioto A, Gross RJ, Scotland SM, Rowe B. An adhesive factor found in strains of *Escherichia coli* belonging to the traditional infantile enteropathogenic serotypes. *Curr Microbiol* 1979; **3** : 95-9
9. Scaletsky ICA, Silva MLM, Trabulsi LR. Distinctive patterns of adherence of enteropathogenic *Escherichia coli* to HeLa cells. *Infect Immun* 1984; **45** : 534-6
10. Nataro JP, Scaletsky ICA, Kaper JB, Levine MM, Trabulsi LR. Plasmid-mediated factors conferring diffuse and localized adherence of enteropathogenic *Escherichia coli*. *Infect Immun* 1985; **48** : 378-83
11. Nataro JP, Baldini MM, Kaper JB, Black RE, Levine MM. Detection of an adherence factor of enteropathogenic *Escherichia coli* with a DNA probe. *J Infect Dis* 1985; **152** : 560-5
12. Nataro JP, Kaper JB, Robins-Browne R, Prado V, Vial P, Levine MM. Patterns of adherence of diarrheagenic *Escherichia coli* to HEP-2 cells. *J Pediatr Infect Dis* 1987 (in press)
13. Levine MM, Nataro JP, Baldini MM, Kaper JB, Black RE, Clements ML, O'Brien AD. The diarrheal response of humans to some classical serotype enteropathogenic *Escherichia coli* is dependent on a plasmid encoding an enteroadhesiveness factor. *J Infect Dis* 1985; **152** : 550-9
14. Marques LRM, Moore MA, Wells J, Wachsmuth IK, O'Brien AD. Production of Shiga-like toxin by *Escherichia coli*. *J Infect Dis* 1986; **154** : 338-41
15. Cleary TG, Mathewson JJ, Faris E, Pickering LK. Shiga-like cytotoxin production by enteropathogenic *Escherichia coli* serogroups. *Infect Immun* 1985; **47** : 335-7
16. Toledo MRF, Alvariza MDB, Murahovschi J, Ramos SRTS, Trabulsi LR. Enteropathogenic *Escherichia coli* serotypes and endemic diarrhea in infants. *Infect Immun* 1983; **39** : 586-9
17. Prado V, Braun S, Bosch P, Bercovich M, Reyes M, Sawada M. Analisis de *Escherichia coli* enteropatogeno clasico (E.C.E.P.) como causa endemica de diarrea aguda en ninos Chilenos. *Rev Chil Pediatr* 1984; **55** : 171-5
18. Gorbach SL, Banwell JG, Chatterjee BD, Jacobs B, Sack RB. Acute undifferentiated diarrhea in the tropics : I. Alterations in intestinal microflora. *J Clin Invest* 1971; **50** : 881-9
19. Sack RB. Enterotoxigenic *Escherichia coli* : Identification and characterization. *J Infect Dis* 1980; **142** : 279-86
20. Black RE, Brown KH, Becker S, Alim Abdul ARM, Huq I. Longitudinal studies of infectious diseases and physical growth of children in rural Bangladesh. II. Incidence of diarrhea and association with well known pathogens. *Am J Epidemiol* 1982; **115** : 315-24
21. Black RE, Merson MH, Huq I, Alim ARMA, Yunus M. Incidence and severity of rotavirus and *Escherichia coli* diarrhoeal in rural Bangladesh. *Lancet* 1981; **i** : 141-3
22. Black RE, Brown KH, Becker S, Alim AR, Huq I. Longitudinal studies of infectious diseases and physical growth of children in rural Bangladesh. II. Incidence of diarrhea and association with known pathogens. *Am J Epidemiol* 1982; **115** : 315-24
23. Merson JH, Morris GK, Sack DA, Wells JG, Feeley JC, Sack RB, Creech WB, Kapikian AZ, Gangarosa EJ. Travelers diarrhea in Mexico : a prospective study of physicians and family members attending a congress. *New Engl J Med* 1976; **294** : 1299-1305
24. Sack DA, Kaminsky DC, Sack RB, Itotia JN, Arthur RR, Kapikian AZ, Orskov F, Orskov I. Prophylactic doxycycline for travelers' diarrhea. *New Engl J Med* 1978; **298** : 758-63
25. DuPont HL, Olarte J, Evans DG, Pickering LK, Galindo E, Evans DJ. Comparative susceptibility of Latin American and United States students to enteric pathogens. *New Engl J Med* 1976; **295** : 1520-1
26. Levine MM, Kaper JB, Black RE, Clements ML. New knowledge on pathogenesis of bacterial enteric infections as applied to vaccine development. *Microbiol Rev* 1983; **47** : 510-50
27. Ristaino PA, Levine MM, Young CR. Improved GMI-enzyme linked immunoabsorbent assay for detection of *Escherichia coli* heatlabile enterotoxin. *J Clin Microbiol* 1983; **18** : 808-815

28. Honda T, Taga S, Takeda Y, Miwatani T. Modified Elek test for detection of heat-labile enterotoxin of enterotoxigenic *Escherichia coli*. *J Clin Microbiol* 1981; **13** : 1-5
29. Lanata CF, Kaper JB, Baldini MM, Black RE, Levine MM. Sensitivity and specificity of DNA probes with the stool blot technique for detection of *Escherichia coli* enterotoxins. *J Infect Dis* 1985; **152** : 1087-1090
30. Thompson MR, Jordan RL, Luttrell MA, Brandwein H, Kaper JB, Levine MM, Giannella RA. Blinded two-laboratory comparative analysis of *Escherichia coli* heat-stable enterotoxin production by using monoclonal antibody, enzyme-linked immunosorbent assay, radioimmunoassay, suckling mouse assay, and gene probes. *J Clin Microbiol* 1986; **24** : 753-758
31. Dean AG, Ching Y-C, Williams RG, Harden LB. Test for *Escherichia coli* enterotoxin using infant mice : application in a study of diarrhea in children in Honolulu. *J Infect Dis* 1972; **125** : 407-411
32. Toledo MRF, Trabulsi L. Correlation between biochemical and serological characteristics of *Escherichia coli* and results of the Sereny test. *J Clin Microbiol* 1983; **17** : 419-21
33. Pal T, Pacsa AS, Emody L, Voros S, Selley. Modified enzyme-linked immunosorbent assay for detecting enteroinvasive *Escherichia coli* and virulent *Shigella* species. *J Clin Microbiol* 1985; **21** : 415-8
34. Wood PK, Morris Jr JG, Small PC, Sethabutr O, Trabulsi L, Kaper J. Comparison of DNA probes with the Sereny test in the identification of invasive strains of *Shigella* and *Escherichia coli*. *J Clin Microbiol* 1986; **24** : 498-500
35. Riley LW, Remis RS, Helgerson SD, MoGee HB, Wells JG, Davis BR, Hebert RJ, Olcott ES, Johnson LM, Hargrett NT, Blake PA, Cohen ML. Hemorrhagic colitis associated with a rare *Escherichia coli* serotype. *New Engl J Med* 1982; **308** : 681-5
36. Levine MM, Xu J-G, Kaper JB, Lior H, Prado V, Tall B, Nataro JP, Karch H, Wachsmuth IK. A DNA probe to identify enterohemorrhagic *Escherichia coli* of 0157 : H7 and other serotypes that cause hemorrhagic colitis and hemolytic uremic syndrome. *J Infect Dis* 1987 ; **156** (in press)
37. Karch H, Heeseman J, Laufs R, O'Brien AD, Tacket CO, Levine MM. A plasmid of enterohemorrhagic *Escherichia coli* 0157 : H7 is required for expression of a new fimbrial antigen and for adhesion to epithelial cells. *Infect Immun* **55** : 455-461
38. Tzipori S, Wachsmuth KI, Chapman C, Birner R, Brittingham J, Jackson C, Hogg J. Studies on the pathogenesis of haemorrhagic colitis caused by *Escherichia coli* 0157 : H7 in gnotobiotic piglets. *J Infect Dis* 1986; **154** : 712-716
39. Vial P, Prado V, Robins-Browne R, Lior H, Maneval D, Tall B, Kaper J, Nataro J, Levine M. Autoagglutinating *E. Coli* (AutEC) —A new enteric pathogen. Abstract, 28th Interscience Conference on Antimicrobial Agents and Chemotherapy, New York October, 1987.
40. Mathewson JJ, Johnson PC, DuPont HL, Satterwhite TK, Winsor DK. Pathogenicity of enteroadherent *Escherichia coli* in adult volunteers. *J Infect Dis* 1986; **154** : 524-7