E. ANDREAKOS, I. UDALOVA, S. SACRE & B.M. FOXWELL

THE ROLE OF NF-κB IN INFLAMMATORY DISEASES

1. MOLECULAR AND CELLULAR BASIS OF INFLAMMATION AND ITS ROLE IN PATHOLOGY

The term inflammation was used as early as 1600 BC in Egyptian papyrus writings to describe a complex series of events induced by a wound or an infection. These events can be recognized by the redness, swelling, fever and pain observed, and are now known to reflect the vasodilation, increased capillary permeability and influx of phagocytic cells. Thus, under most circumstances, inflammation is a normal physiological and very beneficial response, employed by our organism to fight infectious agents and protect us from disease. Inflammation involves the activation of both the innate and adaptive branches of the immune system and is under the control of a large number of cellular and molecular components. Major cellular components include B-cells, T-cells, dendritic cells and macrophages (Table 1).

Major molecular components involved in inflammation comprise cytokines, adhesion molecules, acute phase proteins, kinins, prostaglandins/leukotrienes, nitric oxide (NO) and inducible NO synthase (iNOS), and histamine. Cytokines are small proteins or glycoproteins that act as local messengers in inflammation and immunity, but also in many other important biological processes such as cell growth, development, repair and fibrosis. Cytokines are considered to play a central role in the development of inflammatory responses, as they regulate both the recruitment and activation of the cellular components, as well as the expression of the molecular components of inflammation. Their own expression, in turn, is subject to control by the other molecular components, thus setting up a regulatory loop that can either reduce or extend the persistence of the inflammatory response. Adhesion molecules are essential for the adhesion, activation and transmigration of leukocytes to inflammatory sites as well as the costimulation and activation process of T- and B-cells upon recognition of antigen (Ley, 2001). Acute phase proteins are serum proteins released in response to tissue damage. C-reactive protein, in particular, is produced by the liver and binds to the C-polysaccharide cell wall component of bacteria and fungi, activating complement, mediating phagocytosis and increasing pathogen clearance (Mortensen, 2001).

Kinins are small peptides, found in blood plasma in an inactive form, that become activated in response to tissue injury to cause vasodilation and increased capillary permeability (Couture et al., 2001). Some kinins such as bradykinin, also stimulate pain receptors in the skin. Finally, prostaglandins and leukotrienes are eicosanoid lipid mediators derived from phospholipase-released arachidonic acid that are involved in numerous homeostatic and inflammatory functions, including vasodilation, chemotaxis, fever and pain (Funk, 2001). NO is also a mediator of inflammation that is important in bronchodilation, leukocyte adhesion and extravasation to tissues and antimicrobial activity through its oxidant derivatives (Bogdan, 2001). Production of NO is under the control of two constitutive NO synthase enzymes and one inducible NO synthase enzyme, which renders its study very difficult.

Table 1. Major functions of immune system cells.

Cell type		Function
Lymphoid	Helper T-lymphocytes	Stimulate B-lymphocyte growth and differentiation, and activate macrophages
	Cytotoxic T-lymphocytes	Kill virally infected or tumor cells and activate macrophages
	B-lymphocytes	Present antigen to T-lymphocytes and produce antibodies
	Large granular lymphocytes	Involved in the early responses to virally
	or natural killer (NK) cells	infected or tumor cells and mediate antibody- dependent cellular cytotoxicity
Myeloid	Mononuclear phagocytes	Phagocytose foreign particles and dead cells,
	(monocytes/macrophages)	present antigen to T-lymphocytes, induce
		fibroblast and vascular endothelial cell proli-
		feration important in tissue repair, and recruit other inflammatory cells
	Neutrophils	Phagocytose foreign particles and dead cells
	Eosinophils	Phagocytose IgE-coated particles and mediate immediate-type hypersensitivity reactions
	Basophils/mast cells	Involved in IgE-mediated hypersensitivity
	Platelets	Mediate blood clotting
	Dendritic cells	Present antigen to T-lymphocytes
Endothelial		Regulate the recruitment of effector cells and
		the blood-clotting process

Although inflammation is a very beneficial response, prolonged or deregulated inflammation can lead to extensive tissue damage, disability or even death. A number of human diseases have been attributed to sustained inflammatory responses and include rheumatoid arthritis, inflammatory bowel disease, inflammatory lung diseases (asthma and fibrosing alveolitis), inflammatory skin diseases (psoriasis and incontinentia pigmenti), atherosclerosis, diabetes and neurodegenerative diseases such as Alzheimer's disease and multiple sclerosis. Understanding

what regulates the inflammatory response in these conditions may lead to novel therapeutics of increased efficacy and reduced toxicity.

2. NF-κB IN THE CONTROL OF MOLECULAR FACTORS OF INFLAMMATION

In the past, most studies of inflammation were focusing on the role of mediators, such as cytokines, adhesion molecules, kinins and prostaglandins, in the development of the inflammatory response. However, in recent years, the development of powerful molecular biology tools has made it possible to study not only how distinct mediators regulate each other, but also which signaling pathways and transcription factors are involved and how they interact with each other. Central to the development of the inflammatory response was found to be the transcription factor NF- κ B.

Table 2. Genes involved in inflammation and regulated by NF-κB.

Function	Gene
Activation of immune cells and inflammation	TNFα, IL-1, IL-2, IL-6, IL-12, IL-17, IL-18, IFNγ, GM-CSF
Chemotaxis/recruitment of immune cells	IL-8, MIP- $1\alpha/\beta$, MCP-1, Gro- $\alpha/\beta/\gamma$, eotaxin, ICAM-1, VCAM-1, E-selectin
Angiogenesis/neovascularization	VEGF
Vasodilation/increased capillary permeability	B1 kinin receptor, iNOS, COX-2, PGE2, PLA ₂
Tissue damage	C-reactive protein, hsp32, hsp70
Tissue remodeling	MMP-1, MMP-3, MMP-9, MMP-13
Survival and proliferation	TRAF1/2, c-IAP1/2, XIAP, A1/Bfl-1, bcl-x _L , c-Myc, cyclin D

NF- κ B is essential for host defense and inflammatory responses to microbial and viral infections (Barnes & Karin, 1997; Ghosh et al., 1998). Mouse models deficient in different NF- κ B subunits have demonstrated the pivotal role that NF- κ B plays in lymphoid organ development, immunity and inflammation (Beg et al., 1995; Sha et al., 1995; Weih et al., 1997; Franzoso et al., 1998). In response to pathogens, coordinated activation of NF- κ B occurs in almost every cell type known and includes cells involved in immunity such as neutrophils, macrophages, lymphocytes, endothelial cells and epithelial cells. NF- κ B activation, in turn, induces the expression of numerous inflammatory mediators such as cytokines, adhesion molecules, chemokines, cyclooxygenase 2 (COX-2), nitric oxide and others that contain NF- κ B-binding sites in their promoters (Table 2) (Pahl, 1999). As many of these inflammatory mediators such as TNF α and IL-1 can also activate NF- κ B, a positive regulatory loop is created that maintains the activation of NF- κ B and amplifies the inflammatory response (Karin, 1999). NF- κ B is tightly regulated and provides an important mechanism by which the fine balance

between beneficial immune responses to fight infections and pathological immune responses that lead to tissue damage are achieved. Here possible mechanisms of highly specific NF- κ B-dependent regulation of gene expression during inflammation are discussed.

Homodimers and heterodimers of five distinct Rel proteins form the family of transcription factors collectively referred to as NF-kB. All five proteins are characterized by the presence of a common Rel homology domain responsible for DNA binding and dimerization. Nevertheless there are two distinct classes within the family. The first consists of p65, RelB and c-Rel proteins that are synthesized as mature proteins and encompass the 'transactivating domain'. The second group includes p50 and p52 proteins that first are synthesized as large precursors p105 and p100, respectively, and then undergo proteolytic processing. Neither p50 nor p52 protein contain the transactivating domain. This diversity of NF-κB complexes lays the first basis for specificity of gene regulation. Indeed, there is growing body of evidence that the p50/p50 homodimer (lacking transactivating potential) exerts an inhibitory effect on gene transcription (Schmitz & Baeuerle, 1991; Franzoso et al., 1992; Franzoso et al., 1993; Udalova et al., 2000), possibly due to a competition with p50/p65 for binding to DNA. Overexpression of p50 blocked lipopolysaccharide (LPS)-induced transcription from $TNF\alpha$ promoter reporter constructs, suggesting that this factor is an inhibitor of the TNF α gene (Baer et al., 1998).

It has been also observed that expression of IFN β , which is transcriptionally regulated by NF- κ B in response to a viral infection, is increased in p50 knockout mice (Sha et al., 1995). Which NF- κ B dimer is more likely to interact with a given promoter depends both on cell-specific protein distribution and DNA-binding specificity of different NF- κ B sites, which significantly differ between the dimers (Kunsch et al., 1992; Udalova et al., 2002a).

p65 and c-Rel-containing NF-κB dimers are held in the cytoplasm in an inactive form by specific inhibitors, termed as inhibitors of NF-κB (IκB) proteins such as $I\kappa B\alpha$, $I\kappa B\beta$ and $I\kappa B\epsilon$, which undergo rapid ubiquitin-dependent degradation after the cell becomes exposed to a stimulus. Tissue-specific distribution of IκB molecules together with their different preference for NF-κB dimers may constitute another level of fine gene regulation by virtue of inhibiting specific NF-κB subsets in different cells. Various IκB proteins target different NF-κB species: $I\kappa B\alpha$ and $I\kappa B\beta$ bind preferentially to p50/p65 and p50/c-Rel heterodimers (Thompson et al., 1995b), IkBe associates only with homo- or heterodimeric complexes containing p65 and c-Rel (Simeonidis et al., 1997; Whiteside et al., 1997), and Bcl-3, another member of the IkB family, interacts specifically with p50 and p52 homodimers (Franzoso et al., 1992; Wulczyn et al., 1992). Bcl-3 can form ternary complexes with DNA-bound p52/p52, and enables it to activate transcription directly (Bours et al., 1993; Fujita et al., 1993), while interaction of this unusual member of the IkB family with DNA-bound p50/p50 is believed to induce its dissociation from DNA, allowing other NF-κB species to bind instead (Franzoso et al., 1992; Franzoso et al., 1993; Pan & McEver,

1995). Direct phosphorylation of Rel proteins constitutes another mode of NF- κ B regulation (Schmitz et al., 2001). It has been suggested that Rel phosphorylation is critical for full NF- κ B activation and that this can occur independently of I κ B degradation. Both protein kinase A (PKA)-mediated and casein kinase II (CKII)-mediated phosphorylation of p65 have been demonstrated (Zhong et al., 1997; Wang et al., 2000). Such integration of different signaling pathways allows for activation of distinct Rel proteins to take place in response to different proinflammatory signals.

Upstream kinase cascades that result in I κ B phosphorylation and degradation constitute another level of control of NF- κ B activation as their activity is tightly regulated. Central to this cascade is the I κ B kinase complex (IKK), a large multisubunit kinase complex that comprises IKK α , IKK β and IKK γ , and that is the point of convergence of multiple signals (e.g. microbial and viral products, cytokines and T-cell receptor engagement) leading to I κ B phosphorylation and NF- κ B activation (Karin, 1999). The spectrum of adaptor proteins and kinases that signal downstream from the receptors to the IKK complex differ between microbial (Toll-like receptor), cytokine (e.g. TNF receptors) and T-cell receptor-signaling pathways. Many of the signaling molecules in these pathways are yet to be identified. However, recent studies identified a common molecule, namely receptor-interacting protein kinase (RIP1/RIP2), which is involved in signaling from Toll-like, T-cell and TNF receptors, thus integrating signals for both innate and adaptive immune responses during the inflammation (Fig. 1) (Kelliher et al., 1998; Chin et al., 2002; Kobayashi et al., 2002).

A different mechanism of NF-kB RelB/p52 translocation into the nucleus has recently emerged. RelB is associated with p100 precursor in nonstimulated cells. p105 and p100-containing complexes are held in the cytoplasm by the C-terminal IkB-like domain of the proteins, which is proteolytically processed by ubiquitindependent partial degradation (Karin & Ben-Neriah, 2000). Whether the processing of p105 to p50 is inducible remains in question (Belich et al., 1999; Ciechanover et al., 2001; Dumitru et al., 2000). However, there is clear evidence for regulated processing of p100 (Senftleben et al., 2001; Xiao et al., 2001). Lymphotoxins α and β (LT α , LT β) are essential in both lymphoid organ development and inflammation (Ruddle, 1999). Signaling from the LT β receptor (LT β R) activates the MAP3K protein NIK, which in turn activates IKK α , leading to phosphorylation, ubiquitination and degradation of the C-terminal domain of p100. While the canonical NF-κB pathway, which is essential for innate immunity (Senftleben et al., 2001), is mostly dependent on IKK β (Li et al., 1999a-b), this alternative pathway depends on IKK α and is mostly involved in lymphoid organ development and adaptive immunity (Senftleben et al., 2001). At the same time, signaling through the LT β R has also been shown to potentially use IKK β to induce IκBα phosphorylation and NF-κB activation (Smith et al., 2001). It remains to be determined whether two different NF-κB pathways activate distinct subsets of genes and whether the proinflammatory genes are likely to be regulated by one selected pathway.

A different mechanism of NF- κ B RelB/p52 translocation into the nucleus has recently emerged. RelB is associated with the p100 precursor in nonstimulated cells. p105- and p100-containing complexes are held in the cytoplasm by the C-terminal I κ B-like domain of the proteins, which is proteolytically processed by ubiquitin-dependent partial degradation (Karin & Ben-Neriah, 2000).

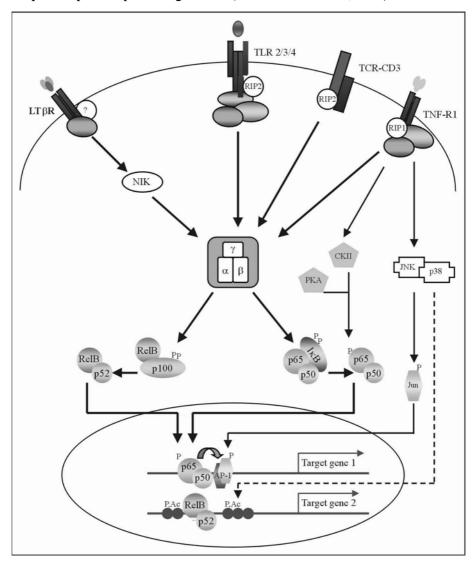


Fig. 1. Integration and specificity in NF- κ B-dependent activation of the proinflammatory genes. In this model different proinflammatory stimuli are recognized by different receptors at the cell surface: TLR (for microbial lipoproteins, LPS, dsRNA), TNF-R1 and LT β R (for cytokines TNF α and LT α LT β 2 trimer) and TCR (for T-cell-specific signaling). They

activate intracellular signaling cascades through distinct sets of adaptor proteins and kinases, with RIP1/2 integrating signals between the cascades. The IKK complex congregates the pathways. The activated IKK complex phosphorylates IκB, mostly by IKKβ, leading to IκB ubiquitination and degradation and the release of free NF-κB p50/p65. Its translocation into the nucleus is accompanied by PKA or CKII-dependent phosphorylation of p65. LTβR-NIK signaling is the only example to date of acting through IKKα subunit of the IKK, resulting in phosphorylation and degradation of p100 and nuclear translocation of NF-κB RelB/p52. In the nucleus p50/p65 and RelB/p52 complexes (and other NF-κB/Rel dimers not described in this simplified model) find specific DNA-binding sites in the promoters of inflammatory mediators (cytokines, adhesion molecules, nitric oxide, macrophage proteins, etc.) based on DNA sequence preferences, association with other transcription factors (e.g. AP-1) and high-order chromatin structure. The chromatin structure of the promoters can either be immediately accessible to NF-κB binding (e.g. the TNFα promoter) or require stimulus-dependent modification of histones (e.g. p38-dependent phosphorylation and acetylation of H3 histone in the IL-6 promoter). Binding of a specific NF-kB complex and its interaction with promoter-specific transcription factors leads to recruitment of coactivators and chromatin remodeling factors and results in highly specific signal-dependent activation of inflammatory gene transcription.

Whether the processing of p105 to p50 is inducible, remains in question (Belich et al., 1999; Dumitru et al., 2000; Ciechanover et al., 2001). However, there is clear evidence for regulated processing of p100 (Senftleben et al., 2001; Xiao et al., 2001). Lymphotoxin α and β (LT α /LT β) are essential in both lymphoid organ development and inflammation (Ruddle, 1999). Signaling from the LT β R activates the MAP3K protein NIK, which in turn activates IKK α , leading to phosphorylation, ubiquitination and degradation of the C-terminal domain of p100.

While the canonical NF- κ B pathway, which is essential for innate immunity (Senftleben et al., 2001), is mostly dependent on IKK β (Li et al., 1999a-b), this alternative pathway depends on IKK α and is mostly involved in lymphoid organ development and adaptive immunity (Senftleben et al., 2001). At the same time, signaling through the LT β R has also been shown to potentially use IKK β to induce I κ B α phosphorylation and NF- κ B activation (Smith et al., 2001). It remains to be determined whether two different NF- κ B pathways activate distinct subsets of genes and whether the proinflammatory genes are likely to be regulated by one selected pathway.

Another mode of specificity in NF- κ B-dependent gene activation lies in its ability to orchestrate gene expression in concert with other transcription factors. Eukaryotic genes are regulated largely through the cooperative interactions of distinct transcription factors and their assembly into multicomponent enhancer complexes (Carey, 1998). The repertoire of proteins NF- κ B interacts with determines the timing, level and duration of gene expression. For instance, the production of IFN β in response to virus induction requires the correct assembly of NF- κ B p50/p65, ATF-2, c-Jun, IRF-3, IRF-7 and HMG-I(Y) into enhanceosome (Thanos & Maniatis, 1995), in which both the composition and the orientation of transcription complexes are essential (Thanos & Maniatis, 1995; Falvo et al.,

2000). Moreover, such assembly leads to ordered recruitment of coactivator molecules, histone acetylases and chromatin remodeling factors and general transcription machinery, that determines activation time and duration of the gene transcription (Agalioti et al., 2000). DNA-binding studies reveal a requirement for NF- κ B and a small group of other transcriptional activators for maximal levels of adhesion molecules during inflammatory responses. The organization of the cytokine-inducible element in the E-selectin promoter is remarkably similar to the IFN β gene in that both promoters require NF- κ B, ATF-2 and HMG-I(Y) (Whitley et al., 1994), whereas vascular cell adhesion molecule-1 (VCAM-1) expression is induced through interactions of NF- κ B with IRF-1 and HMG-I(Y), which also depends on constitutively present SP-1 (Neish et al., 1995a-b). NF- κ B physically associates with the transcription factor NF-IL-6 in synergistic activation of transcription of the inflammatory cytokine IL-6 (Matsusaka et al., 1993), and together with AP-1 of IL-8 (Roebuck, 1999).

The ability of NF- κ B to physically interact with another ubiquitous transcription factor AP-1, whose transcriptional induction follows a distinct signal transduction pathway involving phosphorylation of c-Jun kinase (JNK) (Chang & Karin, 2001), is of particular importance, as it may play a general role in regulation of the inflammatory response by simultaneous activation of the majority of inflammatory mediators (Stein et al., 1993). Indeed, many of the inflammatory genes require these two transcription factors working cooperatively, including VCAM-1 (Ahmad et al., 1998), IL-8 (Roebuck, 1999), COX-2 (Allport et al., 2000), macrophage chemoattractant protein-1 (MCP-1) (Martin et al., 1997) and collagenase-3 (Mengshol et al., 2000).

Both the extensive histone-DNA contacts in individual nucleosomes and the folding of the nucleosomal chain into higher order chromatin structures limit the accessibility of the genome to DNA-binding proteins, including NF-kB. It has been proposed that stimulation with microbial products or proinflammatory cytokines may result in two distinct waves of NF-kB recruitment to target promoters: a fast recruitment to constitutively and immediately accessible promoters (e.g. the TNFα promoter) and a late recruitment to promoters requiring stimulus-dependent modifications in chromatin structure to make NF-kB sites accessible (Saccani et al., 2001). The later recruitment was shown to selectively occur on the promoters of a subset of cytokine and chemokine genes, including IL-6, IL-8 and MCP-1, through p38 MAP kinase-dependent phosphorylation and phosphoacetylation of histone H3 (Saccani et al., 2002). p38 is not a general modulator of NFκB recruitment to chromatin as other cryptic NF-κB sites, e.g. in the macrophage inflammatory protein-1 (MIP-1) promoter, are switched into an accessible state in a p38-independent manner. A highly promoter-specific facilitation of NF-κB recruitment to selected binding sites introduces a novel mechanism of gene activation specificity during inflammatory responses.

The mechanism of gene expression optimization that has received relatively little attention so far is the effect of genetic variations on the level of response to NF- κ B. A naturally occurring polymorphism at nucleotide -863 in the human

TNF α promoter region provided an opportunity to dissect the functional NF- κ B interactions at a single binding site (Udalova et al., 2000). It was found that this site normally binds both p50/p65 and p50/p50 NF-kB complexes, but a single base change specifically inhibited p50/p50 binding. Reporter gene analysis indicated that the variant allele acts to elevate inducible TNF α production. This is of fundamental interest, as it raises the question of how the genetic background may influence the immune gene regulation and consequently result in balancing of the inflammatory response. For instance, patients with severe rheumatoid arthritis, the chronic inflammatory disease in which TNF α has an essential role in pathogenesis, were twice as likely to possess the TNF α -863 allele than the control group (Udalova & Kwiatkowski, 2002). Predictions of the NF-κB-binding sites that are likely to have a functional effect were done for the human chromosome 22 and identified about fifty single nucleotide polymorphisms that would alter NF-κB binding to DNA (Udalova et al., 2002b). Together with a number of other observations of functional consequences of nucleotide polymorphisms that alter protein-DNA interactions (Knight et al., 1999; Farzaneh-Far et al., 2001; Hohjoh & Tokunaga, 2001; Price et al., 2001; van Heel et al., 2002), this novel genetic mechanism of gene regulation may have important implications in disease pathogenesis.

3. NF-κB AND ITS ROLE IN DISEASE PATHOGENESIS

3.1. Scope

The profound effect that NF- κ B has on the regulation of numerous genes involved in inflammation and the ability of many such genes to potently activate NF- κ B themselves, led to the suggestion that NF- κ B may also be essential for the development and/or progression of inflammatory diseases. Currently, an increasing amount of data on the role of NF- κ B is being generated by studies in patients and patient tissue, and is being complemented by studies in relevant animal models.

3.2. Rheumatoid arthritis

The rheumatoid synovium is a site of active inflammation. It is infiltrated by a diverse spectrum of cells such as activated T-lymphocytes, macrophages, fibroblasts, endothelial cells and plasma cells, and is enriched with almost every cytokine and inflammatory mediator known (Feldmann & Maini, 2001). Cytokines such as TNF α and IL-1 that activate NF- κ B are elevated in the synovium, and activated NF- κ B has been detected in human synovial tissue from both early and later stage patients in macrophage- and fibroblast-like synoviocytes (Asahara et al., 1995; Handel et al., 1995; Marok et al., 1996). In particular, macrophage-like synoviocytes that localize in the synovial lining layer and the vascular endothelium have been shown to contain p65 and p50 NF- κ B subunits in their nucleus (Handel et al., 1995). In animal models of arthritis, such as collagen-induced

arthritis in mice or adjuvant-induced arthritis, pristane-induced arthritis and streptococcal cell wall (SCW)-induced arthritis in rats, NF-κB is also activated (Tsao et al., 1997; Han et al., 1998; Miagkov et al., 1998; Palombella et al., 1998).

The activation of NF-kB has important functional consequences. Studies that we performed in dissociated synovial membrane cultures from rheumatoid arthritis patients with active disease demonstrated that overexpression of $I\kappa B\alpha$, the natural inhibitor of NF- κ B, significantly reduces the production of TNF α (Foxwell et al., 1998). As TNF α is central to the inflammatory cascade that takes place in the rheumatoid synovium, and is an already established and proven therapeutic target for the reduction of disease activity, this suggested that NF-kBblocking agents may also result in therapeutic benefit in these patients. Interestingly, inhibition of NF-kB in these cultures also inhibited the production of a large number of other proinflammatory cytokines such as IL-1\beta, IL-6 and IL-8, without significantly affecting the production of the anti-inflammatory cytokines IL-10 and IL-11, or the release of soluble TNF receptors (Bondeson et al., 1999). In addition, inhibition of NF-κB blocked the secretion of matrix metalloproteinases (MMPs) 1, 3 and 13 (Bondeson et al., 1999). These observations suggested that NF-κB and NF-κB-dependent gene expression are central to the production of a large number of proinflammatory, but not of anti-inflammatory mediators in the rheumatoid synovium essential for disease pathogenesis (Fig. 2). These proinflammatory mediators in turn induce NF-κB activation, thus establishing a vicious circle that maintains NF-kB activation and perpetuates inflammation.

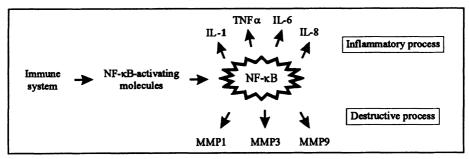


Fig. 2. NF- κB is central to the regulation of inflammatory and destructive processes in the rheumatoid synovium. Therapeutics aimed at blocking NF- κB or upstream NF- κB -activating molecules constitute a promising new approach to the treatment of rheumatoid arthritis and possibly other chronic inflammatory diseases.

NF-κB may also have a prominent role in rheumatoid arthritis joint destruction as it promotes osteoclast maturation and increased bone-resorbing activity (Iotsova et al., 1997), and inhibits at the same chondrocyte differentiation and repair of damaged cartilage tissue (de Crombrugghe et al., 2000; Murakami et al., 2000). In animal models of arthritis, the genetic inactivation of c-Rel and NF-κB1 in rel-/- and nfkb1-/- deficient mice protected the animals from the development of collagen-induced arthritis (CIA) (Campbell et al., 2000), as did the

transgenic expression of (superrepressor) $I_{\kappa}B\alpha$ in cells of the T-cell lineage (Seetharaman et al., 1999). Similarly, the administration of NF- κ B decoy oligonucleotides suppressed the severity of already established disease in both CIA and SCW rat arthritis by inhibiting the production of TNF α and IL-1 within the joints and by reducing paw swelling and disease activity (Miagkov et al., 1998; Tomita et al., 1999). A proteosome inhibitor that prevents $I_{\kappa}B$ degradation also afforded protection in SCW-induced rat arthritis (Palombella et al., 1998). Thus, blocking NF- κB may reduce the inflammatory response and restore the cytokine equilibrium in the rheumatoid joint by reducing at the same time cartilage and bone damage.

3.3. Inflammatory bowel disease (IBD)

IBD comprises a heterogeneous group of diseases including Crohn's disease (CD) and ulcerative colitis (UC). Several genetic and environmental factors are implicated in the pathophysiology of IBD that ultimately result in a common final manifestation, which is mucosal inflammation (Fiocchi, 1998). Deregulated cytokine production and signaling mechanisms by epithelial cells, mucosal lymphocytes and macrophages seem to play a major role in the inflammatory response that takes place in the colonic mucosa. NF-kB, in particular, has been found to be increased in patients with IBD and p65 to be localized in epithelial cells and lamina propria macrophages in biopsies of patients (Neurath et al., 1996; Neurath & Pettersson, 1997; Rogler et al., 1998; Schreiber et al., 1998). Increased NFκB DNA-binding activity has been demonstrated in nuclear extracts from both biopsy specimens and isolated lamina propria mononuclear cells from CD patients (Schreiber et al., 1998), and in vitro treatment of lamina propria macrophages with p65 antisense oligonucleotides was shown to be more effective in downregulating the expression of TNF α , IL-1 β and IL-6 than 5-aminosalicylic acid or glucocorticoids (Neurath et al., 1996; Neurath & Pettersson, 1997). Collectively, these observations suggested that NF- κ B activation may be important in the generation and maintenance of mucosal inflammation, although its activation may not be specific to IBD. Indeed, increased NF-κB levels have also been found in nonspecific colitis or diverticulitis biopsy speciments and were not significantly different from those found in CD or UC specimens (Rogler et al., 1998).

NF-κB was also found to be activated in several animal models that closely resemble human IBD. Thus, in TNBS-induced colitis in mice, increased NF-κB DNA-binding activity, that consisted mainly of the p65 and p50 NF-κB subunits, was detected in nuclear extracts from lamina propria macrophages (Neurath et al., 1996). Inhibition of this activity in vivo by intravenous or local colonic administration of p65 antisense oligonucleotides abrogated the clinical signs of established colitis (Neurath et al., 1996; Neurath & Pettersson, 1997). Strikingly, the efficacy of p65 antisense oligonucleotides in treating TNBS-induced colitis appeared to be superior than the administration (single or daily) of glucocorticoids. In addition, in IL-10-deficient mice, another model of chronic intestinal

inflammation (Kuhn et al., 1993), increased levels of NF-κB DNA-binding activity and increased levels of p65 protein expression were found in lamina propria macrophages. Again, administration of p65 antisense oligonucleotides improved the clinical and histological signs of established colitis (Neurath et al., 1996). Finally, in DSS-induced colitis in mice, intravenous or oral administration of an inhibitor that blocks NF-κB nuclear translocation, reduced both the inflammation and injury scores of the diseased colon (Fujihara et al., 2000). These in vivo studies suggest that specific blockade of NF-κB may also have therapeutic efficacy in the human forms of colonic intestinal inflammation such as CD and UC.

3.4. Psoriasis

Psoriasis is a chronic remitting-relapsing skin disease. It is caused by increased epidermal proliferation and infiltration of inflammatory/immune cells into the epidermis that result in raised scaly plaques on the skin surface and skin lesions. This condition is often seen in conjunction with nail disease and arthritis. Numerous inflammatory mediators have been reported to be upregulated in skin biopsy speciments from psoriatic lesions, including TNF α , IL-1 and IL-15 (Danning et al., 2000). The p65 NF-kB subunit has also been shown to be activated in psoriatic lesions, although it is not known whether this is a cause or consequence of the inflammatory process taking place (Danning et al., 2000). Certainly, experiments in an animal model of psoriasis using a selective proteasome inhibitor that prevents IkB degradation reduced the severity of psoriasis, indicating that NF-κB activation may indeed be essential in that process (Zollner et al., 2002). In addition, the observation that cyclosporin, glucocorticoids and fumaric acid esters, commonly used for the treatment of psoriasis in humans (Asadullah et al., 2002), all can inhibit NF-kB activation, also supports this possibility.

3.5. Inflammatory lung diseases

Acute and chronic alveolar and/or bronchial inflammation also takes place in a number of diseases of the lung, collectively termed inflammatory lung diseases. These include acute lung injury (ALI), acute respiratory distress syndrome (ARDS), fibrosing alveolitis (FA) and asthma, and are characterized by an inflammatory response that involves the recruitment to the lungs and the activation of immune cells. This is accompanied by the upregulation of adhesion molecules and the secretion of cytokines, chemokines, oxidants and other inflammatory mediators, and results in lung injury. Airway epithelial cells contribute to that process as they are also able to produce inflammatory mediators (Thompson et al., 1995a).

NF-κB seems to be important in lung inflammation as increased NF-κB activity has been detected in bronchoalveolar lavage biopsies from ALI (Blackwell & Christman, 1997), ARDS (Schwartz et al., 1996; Christman et al., 1998), FA

(Conron et al., 2002) and asthma patients (Hart et al., 1998) compared to healthy volunteers (Farver et al., 1998). In asthma patients in particular, increased NF- κB activity is also present in the submucosal cells, in the endothelium and the bronchial epithelium (Barnes & Adcock, 1997). Few studies have been performed to evaluate the role of NF-kB in lung inflammation. In bronchoalveolar lavage biopsies from FA patients, we recently showed that overexpression of $I\kappa B\alpha$ inhibits the constitutive production of both macrophage and T-cell-derived cytokines such as TNF α , IL-6, IL-8 and IFN γ , demonstrating that NF- κ B regulates the expression of a number of inflammatory mediators in this system (Conron et al., 2001). The examination of putative kinases that may be involved in that process revealed that IKK2, but not NIK was required (Conron et al., 2002). Several animal models of lung inflammation have also been used. In rat and mouse models of endotoxic shock, intraperitoneal administration of endotoxin induces NF-kB activation in alveolar macrophages and lung tissue that is associated with lung neutrophilia, epithelial permeability and lipid peroxidation (Blackwell et al., 1996; Blackwell et al., 1999). This was suppressed by treatment with antioxidants that inhibit NF-κB activation (Blackwell et al., 1996). Similarly, hemorrhage-induced shock in mice resulted in rapid NF-κB activation in intraparenchymal lung mononuclear cells and in lung inflammation (Shenkar & Abraham, 1997). This may occur through stimulation of α -adrenergic receptors and the generation of reactive oxygen species. Finally, mice deficient in the p50 NF-κB subunit of NF-κB are incapable of mounting eosinophilic airway inflammation. This is due to the lack of production of IL-5 and eotaxin, MIP-1 α and MIP-1\(\beta\), but not to T-cell activation or adhesion molecule expression (Yang et al., 1998). The same was true for c-Rel-deficient mice that were resistant to allergen-induced pulmonary inflammation, airway hyperresponsiveness and eosinophilia (Donovan et al., 1999). As these are common features of asthma, it is likely that p50, c-rel and probably other NF-kB subunits are also important in the human disease. Although the stimuli for NF-kB activation in asthmatic lungs have not been defined, agents that correlate with asthma exacerbations, such as various allergens, ozone, rhinoviruses and respiratory syncytial viruses, have all been shown to activate NF-kB (Christman et al., 2000).

3.6. Atherosclerosis

Atherosclerosis and its clinical manifestations of heart attack, stroke and peripheral vascular insufficiency can be viewed as a multistep, chronic inflammatory disease. The development of atherosclerotic lesions can be subdivided into three phases, viz. the initiation phase (formation of small fatty streaks), the expansion phase (endothelial cell growth and coalescence of fatty streaks) and the progression phase to plaques (recruitment of smooth muscle cells, deposition of collagen and formation of a fibrous cap). This process is under the control of cytokines, chemokines, growth factors and other soluble mediators and adhesion molecules (produced within the lesion) that regulate the recruitment and proliferation of

monocytes and macrophages to the vessel wall, the migration of smooth muscle cells from the media to the intima of the vessel, and the proliferation of vascular endothelial cells. As many of these inflammatory mediators can be regulated by NF-κB, it has been suggested that NF-κB may also play a major role in the pathogenesis of atherosclerosis (Collins & Cybulsky, 2001). Indeed, activated NF-kB has been detected in human atherosclerotic plaques, whereas it is almost absent in healthy vessels devoid of atherosclerosis. Studies using an antibody that recognizes p65 dissociated from IkB revealed that activated p65 localizes in the intima and media of atheromatous areas of atherosclerotic lesions in smooth muscle cells, macrophages and endothelial cells (Brand et al., 1996). This is in contrast to healthy vessels where p65 and p50 are mainly localized in the cytoplasm. Elevated NF-kB levels have also been detected in arterial smooth muscle cells after injury and have been suggested to account for the expression of several genes, including ICAM-1, VCAM-1 and MCP-1, all of which facilitate the recruitment of monocytes and macrophages to the atherosclerotic lesions (Landry et al., 1997).

Few studies have also examined the activation of NF-κB in animal models of atherosclerosis. One study demonstrated activated NF-kB in intimal cells from coronary arteries of pigs fed with a hypercholesterolemic diet (Wilson et al., 2000). Interestingly, another study in mice suggested that there is elevated NF- κ B expression in vessel regions of high probability for the development of atherosclerotic plagues when compared to adjacent regions where plagues are unlikely to develop (Hajra et al., 2000). As these high probability regions coincide with areas exposed to high shear stress, the authors of the study proposed that shear stress may prime certain regions of the vessel wall by inducing steady state levels of NF-κB that may upregulate the expression of cytokines and adhesion molecules upon an additional atherogenic stimulus. In humans, many atherogenic stimuli or risk factors have been shown to activate NF-kB. Thus, oxidative stress (Li & Karin, 1999), oxidized LDL and VLDL (Brand et al., 1997; Calara et al., 1998; Dichtl et al., 1999), hypertension and angiotensin II (Han et al., 1999; Pueyo et al., 2000), hyperglycemia and elevated advanced glycation end products (Morigi et al., 1998; Schmidt et al., 1999), homocysteine and various infectious agents such as cytomegalovirus and Chlamydia pneumonia (Welch & Loscalzo, 1998) have all been shown to induce NF-kB activation in vitro or in vivo. In summary, there is good evidence that deregulated NF-kB may be involved in the development, but also some of the pathological features of atherosclerosis, although more studies are needed to demonstrate that blockade of NF-kB can result in therapeutic benefit in animal models of the disease and ultimately patients.

3.7. Diabetes

There are two forms of diabetes, viz. type 1 (often referred to as insulin-dependent diabetes mellitus or juvenile diabetes) and type 2, known as insulin-independent diabetes or adult onset diabetes. Type 1 is the most serious and is caused by

the autoimmune apoptotic destruction of the insulin-producing beta cells in the islets of Langerhans. This involves the activation of T-cells and antigen-presenting cells, and the production of cytokines such as $\text{INF}\gamma$, $\text{TNF}\alpha$ and $\text{IL-1}\beta$ (Igaz et al., 2000). Type 2 diabetes develops slowly and is probably due to alterations in fat metabolism as the body gains weight and becomes resistant to insulin. Common complications of diabetes include hypertension and cardiovascular disease.

Inappropriate NF-κB activation has recently been implicated in both types of diabetes. In type 1 diabetes, peripheral blood mononuclear cells from patients have increased NF-κB DNA-binding activity (Hofmann et al., 1998; Bierhaus et al., 2001) and increased NF-κB DNA-binding activity has been associated with more pronounced diabetic complications such as diabetic nephropathy (Hofmann et al., 1999; Mohamed et al., 1999). In human islets, the expression of a mutated nondegradable form of the $I\kappa B\alpha$ or the antiapoptotic gene A20 (that also inhibits NF-κB in this system) prevents cytokine-induced beta cell death (Grey et al., 1999; Giannoukakis et al., 2000; Heimberg et al., 2001) by blocking the apoptotic pathway and probably by preventing the NF-kB-dependent upregulation of the Fas gene (Darville & Eizirik, 2001). Evidence for an important role of NFκB in type 1 diabetes also comes from studies in animal models. In chemically induced diabetes (alloxan and steptozotocin-induced diabetes) in mice, administration of NF-kB decoy oligonucleotides (Quan et al., 2001) or other NF-kB inhibitors such as dietary zinc and Amomum xanthoides extract (Ho et al., 2001; Park & Park, 2001) inhibited the development of diabetes. In addition, nonobese diabetic (NOD) mice that are commonly used to model human type 1 diabetes exhibit elevated levels of NF-kB activation that leads to enhanced DC and T-cell function and increased autoimmune destruction of the islets of Langerhans (Hayashi et al., 1993; Weaver et al., 2001; Poligone et al., 2002). Polymorphisms in the NF-κB locus have recently been associated with human type 1 diabetes (Hegazy et al., 2001).

NF- κ B has also been implicated in the development of insulin-resistance that leads to type II diabetes. The discovery that salicylates used to treat insulin-resistance potently inhibit IKK β (Yin et al., 1998) and the observation that IKK β -deficient mice are protected from the development of fat-induced insulin resistance (Kim et al., 2001) suggest that NF- κ B activation may be essential for that process.

3.8. Neurodegenerative diseases

3.8.1. Scope

Most cells within the nervous system (including neurons, astrocytes, microglia and oligodendrocytes) express functional NF- κ B (Kaltschmidt et al., 1994; O'Neill & Kaltschmidt, 1997). This can be upregulated by a wide spectrum of stimuli that include reactive oxygen species (Schreck et al., 1992), inflammatory cytokines (Barger et al., 1995), viral and bacterial products, depolarization

(Guerrini et al., 1995; Kaltschmidt et al., 1995), nerve growth factor (Carter et al., 1996), β -amyloid (Barger & Mattson, 1996; the secreted form of β -amyloid precursor protein) and dopamine (Weingarten et al., 2001). Recently, NF- κ B activation was suggested to play an important role in the development of many neurodegenerative diseases by regulating survival and growth of neurons, microglia and astrocytes.

3.8.2. Multiple sclerosis (MS)

MS is a demyelinating disease where myelin in the central nervous system is lost. This is a result of an immune response that develops against oligodendrocytes (the cells that produce myelin) and effectively inhibits their function and induces their depletion (Kieseier et al., 1999). The mechanisms, however, that regulate these events remain poorly understood. Studies by immunohistochemistry have shown that NF- κ B is upregulated and localized in the nuclei of microglia and oligodendrocytes in chronic active MS lesions (Gveric et al., 1998; Bonetti et al., 1999). As IL-1 and TNF α are also localized in active MS lesions, these cytokines are likely to account for the observed upregulation of NF- κ B (Hofman et al., 1989; Selmaj et al., 1991; Woodroofe & Cuzner, 1993; Liu et al., 1996; Bierhaus et al., 2001). Upregulated NF- κ B, in turn, may amplify the inflammatory process taking place in MS by increasing the expression of NF- κ B-controlled adhesion molecules and cytokines. Ultimately, this leads to disease progression, disability and even death.

In experimental encephalomyelitis (EAE), an animal model of MS and autoimmune neural damage, NF- κ B activation has also been reported (Pahan & Schmid, 2000). Interestingly, NF- κ B-deficient mice are resistant to induced EAE, possibly due to a deficiency in the differentiation of myelin oligodendrocyte glycoprotein-specific T-cells to Th1- or Th2-type effector cells (Hilliard et al., 1999). However, although inflammatory factors that become upregulated by NF- κ B may act to promote autoimmune disease, NF- κ B may at the same time upregulate many antiapoptotic genes, thus protecting neurons and oligodendrocytes from cell death and further damage (Beg & Baltimore, 1996; Van Antwerp et al., 1996; Wang et al., 1996; Bonetti et al., 1997; Bonetti & Raine, 1997). The pathogenesis of this disease may thus rely on the balance between the 'beneficial' and 'pathological' effects of NF- κ B.

3.8.3. Alzheimer's disease (AD)

AD is a neurodegenerative disease that affects the elderly. The main symptoms are memory loss and dementia, with the severity of disease being linked to the amplitude of neuronal and synaptic loss. Aggregates of amyloid beta peptides build up forming plaques along with neurofibrilary tangles in neurons that interfere with the normal function of the brain (Walsh et al., 2002). Interestingly, the areas of the brain that are affected by AD pathology also demonstrate increased NF- κ B activity (Yan et al., 1995). Thus, p65 NF- κ B has been localized in neurons and astrocytes next to amyloid plaques found in brain sections from AD

patients (Terai et al., 1996; Kaltschmidt et al., 1997; O'Neill & Kaltschmidt, 1997; Ferrer et al., 1998). A mechanism by which NF- κ B may worsen AD has been proposed. NF- κ B can be activated by a number of stimuli that include soluble amyloid precursor protein (Barger & Mattson, 1996), β -amyloid peptides possibly through activation of RAGE (Kaltschmidt et al., 1997), IL-1 β and glutamate (Grilli et al., 1996a). This creates a positive regulatory loop as NF- κ B also upregulates the expression of the amyloid precursor protein gene (Grilli et al., 1996a), producing higher levels of β -amyloid peptide that further activates NF- κ B and induces neuronal cell death. The observation, however, that NF- κ B can also be beneficial in AD by upregulating the expression of genes like SOD (Mattson et al., 1997; Keller et al., 1998) and other antiapoptotic proteins that may aid neuronal survival (Barger et al., 1995; Zong et al., 1999) has complicated matters. Therefore, it is still unclear whether NF- κ B is a potential therapeutic target in AD, and more studies are needed.

4. NF-κB IN THERAPY

Although there is a large amount of data that deregulated NF- κ B is central to the pathogenesis of a number of inflammatory diseases, the therapeutic potential of NF- κ B blockade has not yet been tested in the clinic. Evidence, however, from other immunosuppressive and anti-inflammatory drugs already used for the treatment of these diseases (Table 3) suggests that it is likely to be very beneficial, as the common property of these drugs is their ability to inhibit NF- κ B activation among their multiple effects.

Table 3. Drugs used for the treatment of inflammatory diseases that have been proposed to block $NF-\kappa B$.

Disease	Treatment
Rheumatoid arthritis	Glucocorticoids, NSAIDs, cyclosporin A and tacromilus, methotrexate, sulfasalazine, leflunomide, gold and D-penicillamine compounds, anti-TNF α blocking agents
IBD	Glucocorticoids, NSAIDs, cyclosporin A and tacromilus, methotrexate, sulfasalazine, anti-TNF α blocking agents
Psoriasis	Glucocorticoids, NSAIDs, cyclosporin A and tacromilus, methotrexate, anti-TNF blocking agents
Inflammatory lung diseases Atherosclerosis	Glucocorticoids, NSAIDs, cyclosporin A and tacromilus NSAIDs
Insulin resistance (diabetes) Neurodegenerative diseases	NSAIDs Glucocorticoids, NSAIDs, cyclosporin A and tacromilus

Thus glucocorticoids, such as dexamethasone and prednisone, which are frequently used for the treatment of rheumatoid arthritis, IBD and inflammatory lung diseases (Geier & Miner, 1992; Caesar et al., 1997; Keatings et al., 1997; Norbiato et al., 1997; Wilckens & De Rijk, 1997) owe, at least in part, their

therapeutic effects to their ability to inhibit NF- κ B-dependent gene expression. Glucocorticoids have been shown to upregulate the transcription and expression of $I\kappa B\alpha$, possibly by yet unidentified glucocorticoid-responsive elements in the $I\kappa B\alpha$ promoter (Auphan et al., 1995; Scheinman et al., 1995a), and to physically interact with p65, thereby preventing transactivation of NF- κ B (Caldenhoven et al., 1995; Scheinman et al., 1995b; De Bosscher et al., 1997; Heck et al., 1997).

In addition, NSAIDs were recently suggested to inhibit NF- κ B activity, an effect that could account for their anti-inflammatory action. Thus, sodium salicylates and aspirin were shown to inhibit I κ B α phosphorylation and degradation by preventing ATP-binding to IKK2 at concentrations found in the serum of patients treated for inflammatory conditions (Kopp & Ghosh, 1994; Grilli et al., 1996b; Pierce et al., 1996; Yin et al., 1998). At the same time, methotrexate, sulfasalazine and leflunomide were shown to inhibit I κ B α phosphorylation and degradation, probably by interfering with the IKK complex (Egan & Sandborn, 1998; Wahl et al., 1998; Manna et al., 2000; Majumdar & Aggarwal, 2001), and demonstrating that certain NSAIDs may exert their anti-inflammatory effects by blocking the activation of NF- κ B.

Similarly, gold and D-penicillamine compounds and thalidomine, that are used drugs, especially for the treatment of rheumatoid arthritis, all have inhibitory effects on NF-κB activation (Yang et al., 1995; Keifer et al., 2001). Immunosuppressive agents such as cyclosporin A and tacrolimus (FK-506), that are mainly employed in transplantation, but also in inflammatory diseases such as rheumatoid arthritis, were also able to block the activation of NF-kB in addition to their effects in other pathways such as the activation of NF-AT. Cyclosporin A has been claimed to prevent IκBα degradation and NF-κB activation by interfering with the proteasome function (Marienfeld et al., 1997; Meyer et al., 1997), whereas tacromilus seems to specifically inhibit c-Rel, but not RelB or p50 nuclear translocation (Venkataraman et al., 1995). Finally, anticytokine biologicals such as anti-TNF α blocking agents, shown to be highly successful in the treatment of a number of inflammatory diseases (Andreakos et al., 2002), may owe part of their efficacy to the prevention of cytokine-induced NF-κB activation and NF-kB-dependent gene expression. Support for that comes from a study in refractory Crohn's disease, where anti-TNFα blocking agents have been demonstrated to induce disease remission in 30-50% of the patients. It was shown that the anti-TNF α blocking agent infliximab reduced significantly nuclear p65 expression in the colonic mucosa within one week after a single infusion, and an extended downregulation of p65 seemed to be associated with a sustained induction of remission (Nikolaus et al., 2000). In contrast, reaccumulation of nuclear p65 correlated with the increase in TNF α secretion, and was usually observed before patients presented with a clinical relapse. These findings indicated that the activation of NF-κB may be secondary to the secretion of TNFα, and suggested a possible mechanism by which infliximab improves Crohn's disease.

However, as the majority of these drugs is associated with various side effects, especially in the long term, the specific inhibition of NF- κ B may result in

increased therapeutic efficacy and reduced toxicity. Initial attempts to identify novel NF-κB inhibitors focused on interfering with proteasome function or using natural plant products such as flavonoids to prevent IκB degradation and NF-κB activation. However, it became apparent that these inhibitors would also lack specificity and thus be subject to more severe side effects. Current attempts are focusing on the design of specific NF-κB inhibitors that block NF-κB without affecting other pathways. This requires the identification of upstream signaling components that lead to the activation of NF-kB, the determination of which of these are activated and rate-limiting in disease states, as well as the elucidation of their molecular structure and biochemical properties. In that way, some of the main worries concerning NF-κB-specific drugs, such as liver toxicity or interference with host defences in the long term (Beg et al., 1995; Li et al., 1999a-b; Tanaka et al., 1999; Lavon et al., 2000; Rudolph et al., 2000) may be overcome. Certainly, the fact that NSAIDs or glucocorticoids, that block NF-κB, can be tolerated long-term indicates that the use of NF- κ B inhibitors is a valid strategy. Of course, under conditions where NF-κB inhibitors will be needed for shortterm therapy, these worries are not of much concern. Ultimately, clinical trials will be needed to resolve these issues and assess the potential of NF- κ B inhibitors as novel anti-inflammatory agents as well as their therapeutic efficacy.

5. REFERENCES

- Agalioti T, Lomvardas S, Parekh B, Yie J, Maniatis T & Thanos D (2000) Ordered recruitment of chromatin modifying and general transcription factors to the IFN-β promoter. Cell 103: 667-678.
- Ahmad M, Theofanidis P & Medford RM (1998) Role of activating protein-1 in the regulation of the vascular cell adhesion molecule-1 gene expression by tumor necrosis factor- α . J. Biol. Chem. 273: 4616-4621.
- Allport VC, Slater DM, Newton R & Bennett PR (2000) NF-κB and AP-1 are required for cyclo-oxygenase 2 gene expression in amnion epithelial cell line (WISH). Mol. Hum. Reprod. 6: 561-565.
- Andreakos E, Foxwell B, Brennan F, Maini R & Feldmann M (2002) Cytokines and anti-cytokine biologicals in autoimmunity: present and future. Cytokine Growth Factor Rev. 13: 299-314.
- Asadullah K, Volk HD & Sterry W (2002) Novel immunotherapies for psoriasis. Trends Immunol. 23: 47-53.
- Asahara H, Asanuma M, Ogawa N, Nishibayashi S & Inoue H (1995) High DNA-binding activity of transcription factor NF-κB in synovial membranes of patients with rheumatoid arthritis. Biochem. Mol. Biol. Int. 37: 827-832.
- Auphan N, DiDonato JA, Rosette C, Helmberg A & Karin M (1995) Immunosuppression by gluco-corticoids: inhibition of NF-κB activity through induction of IκB synthesis. Science 270: 286-290.
- Baer M, Dillner A, Schwartz RC, Sedon C, Nedospasov S & Johnson PF (1998) Tumor necrosis factor α transcription in macrophages is attenuated by an autocrine factor that preferentially induces NF- κ B p50. Mol. Cell. Biol. 18: 5678-5689.
- Barger SW, Horster D, Furukawa K, Goodman Y, Krieglstein J & Mattson MP (1995) Tumor necrosis factors α and β protect neurons against amyloid β -peptide toxicity: evidence for involvement of a κ B-binding factor and attenuation of peroxide and Ca²⁺ accumulation. Proc. Natl. Acad. Sci. USA 92: 9328-9332.
- Barger SW & Mattson MP (1996) Induction of neuroprotective κ B-dependent transcription by secreted forms of the Alzheimer's β -amyloid precursor. Brain Res. Mol. Brain Res. 40: 116-126.
- Barnes PJ & Adcock IM (1997) NF-κB: a pivotal role in asthma and a new target for therapy. Trends

- Pharmacol. Sci. 18: 46-50.
- Barnes PJ & Karin M (1997) Nuclear factor-κB: a pivotal transcription factor in chronic inflammatory diseases. N. Engl. J. Med. 336: 1066-1071.
- Beg AA, Sha WC, Bronson RT, Ghosh S & Baltimore D (1995) Embryonic lethality and liver degeneration in mice lacking the RelA component of NF-κB. Nature 376: 167-170.
- Beg AA & Baltimore D (1996) An essential role for NF- κ B in preventing TNF- α -induced cell death. Science 274: 782-784.
- Belich MP, Salmeron A, Johnston LH & Ley SC (1999) TPL-2 kinase regulates the proteolysis of the NF-κB-inhibitory protein NF-κB1 p105. Nature 397: 363-368.
- Bierhaus A, Schiekofer S, Schwaninger M, Andrassy M, Humpert PM, Chen J, Hong M, Luther T, Henle T, Kloting I, Morcos M, Hofmann M, Tritschler H, Weigle B, Kasper M, Smith M, Perry G, Schmidt AM, Stern DM, Haring HU, Schleicher E & Nawroth PP (2001) Diabetes-associated sustained activation of the transcription factor nuclear factor-kB. Diabetes 50: 2792-2808.
- Blackwell TS, Blackwell TR, Holden EP, Christman BW & Christman JW (1996) In vivo antioxidant treatment suppresses nuclear factor-κB activation and neutrophilic lung inflammation. J. Immunol. 157: 1630-1637.
- Blackwell TS & Christman JW (1997) The role of nuclear factor-κB in cytokine gene regulation. Am. J. Respir. Cell Mol. Biol. 17: 3-9.
- Blackwell TS, Yull FE, Chen CL, Venkatakrishnan A, Blackwell TR, Hicks DJ, Lancaster LH, Christman JW & Kerr LD (1999) Use of genetically altered mice to investigate the role of nuclear factor-κB activation and cytokine gene expression in sepsis-induced ARDS. Chest 116: 73S-74S.
- Bogdan C (2001) Nitric oxide and the immune response. Nat. Immunol. 2: 907-916.
- Bondeson J, Foxwell B, Brennan F & Feldmann M (1999) Defining therapeutic targets by using adenovirus: blocking NF-κB inhibits both inflammatory and destructive mechanisms in rheumatoid synovium but spares anti-inflammatory mediators. Proc. Natl. Acad. Sci. USA 96: 5668-5673.
- Bonetti B, Pohl J, Gao YL & Raine CS (1997) Cell death during autoimmune demyelination: effector but not target cells are eliminated by apoptosis. J. Immunol. 159: 5733-5741.
- Bonetti B & Raine CS (1997) Multiple sclerosis: oligodendrocytes display cell death-related molecules in situ but do not undergo apoptosis. Ann. Neurol. 42: 74-84.
- Bonetti B, Stegagno C, Cannella B, Rizzuto N, Moretto G & Raine CS (1999) Activation of NF-κB and c-jun transcription factors in multiple sclerosis lesionsImplications for oligodendrocyte pathology. Am. J. Pathol. 155: 1433-1438.
- Bours V, Franzoso G, Azarenko V, Park S, Kanno T, Brown K & Siebenlist U (1993) The oncoprotein Bcl-3 directly transactivates through κB motifs via association with DNA-binding p50B homodimers. Cell 72: 729-739.
- Brand K, Page S, Rogler G, Bartsch A, Brandl R, Knuechel R, Page M, Kaltschmidt C, Baeuerle PA & Neumeier D (1996) Activated transcription factor nuclear factor-κB is present in the atherosclerotic lesion. J. Clin. Invest. 97: 1715-1722.
- Brand K, Eisele T, Kreusel U, Page M, Page S, Haas M, Gerling A, Kaltschmidt C, Neumann FJ, Mackman N, Baeurele PA, Walli AK & Neumeier D (1997) Dysregulation of monocytic nuclear factor-κB by oxidized low-density lipoprotein. Arterioscler. Thromb. Vasc. Biol. 17: 1901-1909.
- Caesar I, Gross V, Roth M, Andus T, Schmidt C, Raedsch R, Weber A, Gierend M, Ewe K & Scholmerich J (1997) Treatment of active and postactive ileal and colonic Crohn's disease with oral pH-modified-release budesonide. German Budesonide Study Group. Hepatogastroenterology 44: 445-451
- Calara F, Dimayuga P, Niemann A, Thyberg J, Diczfalusy U, Witztum JL, Palinski W, Shah PK, Cercek B, Nilsson J & Regnstrom J (1998) An animal model to study local oxidation of LDL and its biological effects in the arterial wall. Arterioscler. Thromb. Vasc. Biol. 18: 884-893.
- Caldenhoven E, Liden J, Wissink S, Van de Stolpe A, Raaijmakers J, Koenderman L, Okret S, Gustafsson JA & Van der Saag PT (1995) Negative cross-talk between RelA and the glucocorticoid receptor: a possible mechanism for the antiinflammatory action of glucocorticoids. Mol. Endocrinol. 9: 401-412.
- Campbell IK, Gerondakis S, O'Donnell K & Wicks IP (2000) Distinct roles for the NF-κB1 (p50) and

- c-Rel transcription factors in inflammatory arthritis. J. Clin. Invest. 105: 1799-1806.
- Carey M (1998) The enhanceosome and transcriptional synergy. Cell 92: 5-8.
- Carter BD, Kaltschmidt C, Kaltschmidt B, Offenhauser N, Bohm-Matthaei R, Baeuerle PA & Barde YA (1996) Selective activation of NF-κB by nerve growth factor through the neurotrophin receptor p75. Science 272: 542-545.
- Chang L & Karin M (2001) Mammalian MAP kinase signalling cascades. Nature 410: 37-40.
- Chin AI, Dempsey PW, Bruhn K, Miller JF, Xu Y & Cheng G (2002) Involvement of receptor-interacting protein 2 in innate and adaptive immune responses. Nature 416: 190-194.
- Christman JW, Lancaster LH & Blackwell TS (1998) Nuclear factor κB: a pivotal role in the systemic inflammatory response syndrome and new target for therapy. Intensive Care Med. 24: 1131-1138.
- Christman JW, Sadikot RT & Blackwell TS (2000) The role of nuclear factor-κB in pulmonary diseases. Chest 117: 1482-1487.
- Ciechanover A, Gonen H, Bercovich B, Cohen S, Fajerman I, Israel A, Mercurio F, Kahana C, Schwartz AL, Iwai K & Orian A (2001) Mechanisms of ubiquitin-mediated limited processing of the NF-κB1 precursor protein p105. Biochimie 83: 341-349.
- Collins T & Cybulsky MI (2001) NF-κB: pivotal mediator or innocent bystander in atherogenesis? J. Clin. Invest. 107: 255-264.
- Conron M, Bondeson J, Pantelidis P, Beynon HL, Feldmann M, duBois RM & Foxwell BM (2001) Alveolar macrophages and T cells from sarcoid but not normal lung are permissive to adenovirus infection and allow analysis of NF-κB-dependent signaling pathways. Am. J. Respir. Cell. Mol. Biol. 25: 141-149.
- Conron M, Andreakos E, Pantelidis P, Smith C, Beynon HL, Dubois RM & Foxwell BM (2002) Nuclear factor-κB activation in alveolar macrophages requires IκB kinase-β but not nuclear factor-κB inducing kinase. Am. J. Respir. Crit. Care Med. 165: 996-1004.
- Couture R, Harrisson M, Vianna RM & Cloutier F (2001) Kinin receptors in pain and inflammation. Eur. J. Pharmacol. 429: 161-176.
- Danning CL, Illei GG, Hitchon C, Greer MR, Boumpas DT & McInnes IB (2000) Macrophage-derived cytokine and nuclear factor κB p65 expression in synovial membrane and skin of patients with psoriatic arthritis. Arthritis Rheum. 43: 1244-1256.
- Darville MI & Eizirik DL (2001) Cytokine induction of Fas gene expression in insulin-producing cells requires the transcription factors NF-κB and C/EBP. Diabetes 50: 1741-1748.
- De Bosscher K, Schmitz ML, Vanden Berghe W, Plaisance S, Fiers W & Haegeman G (1997) Gluco-corticoid-mediated repression of nuclear factor-κB-dependent transcription involves direct interference with transactivation. Proc. Natl. Acad. Sci. USA 94: 13504-13509.
- de Crombrugghe B, Lefebvre V, Behringer RR, Bi W, Murakami S & Huang W (2000) Transcriptional mechanisms of chondrocyte differentiation. Matrix Biol. 19: 389-394.
- Dichtl W, Nilsson L, Goncalves I, Ares MP, Banfi C, Calara F, Hamsten A, Eriksson P & Nilsson J (1999) Very low-density lipoprotein activates nuclear factor-κB in endothelial cells. Circ. Res. 84: 1085-1094.
- Donovan CE, Mark DA, He HZ, Liou HC, Kobzik L, Wang Y, De Sanctis GT, Perkins DL & Finn PW (1999) NF-κB/Rel transcription factors: c-Rel promotes airway hyperresponsiveness and allergic pulmonary inflammation. J. Immunol. 163: 6827-6833.
- Dumitru CD, Ceci JD, Tsatsanis C, Kontoyiannis D, Stamatakis K, Lin JH, Patriotis C, Jenkins NA, Copeland NG, Kollias G & Tsichlis PN (2000) TNF-α induction by LPS is regulated posttranscriptionally via a Tpl2/ERK-dependent pathway. Cell 103: 1071-1083.
- Egan LJ & Sandborn WJ (1998) Inhibition of nuclear factor κB by sulfasalazine: a new target for inflammatory bowel disease therapy? Gastroenterology 115: 1295-1296.
- Falvo JV, Uglialoro AM, Brinkman BM, Merika M, Parekh BS, Tsai EY, King HC, Morielli AD, Peralta EG, Maniatis T, Thanos D & Goldfeld AE (2000) Stimulus-specific assembly of enhancer complexes on the tumor necrosis factor α gene promoter. Mol. Cell. Biol. 20: 2239-2247.
- Farver CF, Raychaudhuri B, Buhrow LT, Connors MJ & Thomassen MJ (1998) Constitutive NF-κB levels in human alveolar macrophages from normal volunteers. Cytokine 10: 868-871.
- Farzaneh-Far A, Davies JD, Braam LA, Spronk HM, Proudfoot D, Chan SW, O'Shaughnessy KM,

- Weissberg PL, Vermeer C & Shanahan CM (2001) A polymorphism of the human matrix γ -carboxyglutamic acid protein promoter alters binding of an activating protein-1 complex and is associated with altered transcription and serum levels. J. Biol. Chem. 276: 32466-32473.
- Feldmann M & Maini RN (2001) Anti-TNF α therapy of rheumatoid arthritis: what have we learned? Annu. Rev. Immunol. 19: 163-196.
- Ferrer I, Marti E, Lopez E & Tortosa A (1998) NF- κ B immunoreactivity is observed in association with β A4 diffuse plaques in patients with Alzheimer's disease. Neuropathol. Appl. Neurobiol. 24: 271-277.
- Fiocchi C (1998) Inflammatory bowel disease: etiology and pathogenesis. Gastroenterology 115: 182-205.
- Foxwell B, Browne K, Bondeson J, Clarke C, de Martin R, Brennan F & Feldmann M (1998) Efficient adenoviral infection with $I_{\kappa}B_{\alpha}$ reveals that macrophage tumor necrosis factor α production in rheumatoid arthritis is NF- κ B dependent. Proc. Natl. Acad. Sci. USA 95: 8211-8215.
- Franzoso G, Bours V, Park S, Tomita-Yamaguchi M, Kelly K & Siebenlist U (1992) The candidate oncoprotein Bcl-3 is an antagonist of p50/NF-κB-mediated inhibition. Nature 359: 339-342.
- Franzoso G, Bours V, Azarenko V, Park S, Tomita-Yamaguchi M, Kanno T, Brown K & Siebenlist U (1993) The oncoprotein Bcl-3 can facilitate NF-κB-mediated transactivation by removing inhibiting p50 homodimers from select κB sites. EMBO J. 12: 3893-3901.
- Franzoso G, Carlson L, Poljak L, Shores EW, Epstein S, Leonardi A, Grinberg A, Tran T, Scharton-Kersten T, Anver M, Love P, Brown K & Siebenlist U (1998) Mice deficient in nuclear factor (NF)-κB/p52 present with defects in humoral responses germinal center reactions and splenic microarchitecture. J. Exp. Med. 187: 147-159.
- Fujihara SM, Cleaveland JS, Grosmaire LS, Berry KK, Kennedy KA, Blake JJ, Loy J, Rankin BM, Ledbetter JA & Nadler SG (2000) A D-amino acid peptide inhibitor of NF-κB nuclear localization is efficacious in models of inflammatory disease. J. Immunol. 165: 1004-1012.
- Fujita T, Nolan GP, Liou HC, Scott ML & Baltimore D (1993) The candidate proto-oncogene bcl-3 encodes a transcriptional coactivator that activates through NF-κB p50 homodimers. Genes Dev. 7: 1354-1363.
- Funk CD (2001) Prostaglandins and leukotrienes: advances in eicosanoid biology. Science 294: 1871-1875.
- Geier DL & Miner PB (1992) New therapeutic agents in the treatment of inflammatory bowel disease. Am. J. Med. 93: 199-208.
- Ghosh S, May MJ & Kopp EB (1998) NF-κB and Rel proteins: evolutionarily conserved mediators of immune responses. Annu. Rev. Immunol. 16: 225-260.
- Giannoukakis N, Rudert WA, Trucco M & Robbins PD (2000) Protection of human islets from the effects of interleukin- 1β by adenoviral gene transfer of an IkB repressor. J. Biol. Chem. 275: 36509-36513.
- Grey ST, Arvelo MB, Hasenkamp W, Bach FH & Ferran C (1999) A20 inhibits cytokine-induced apoptosis and nuclear factor κB-dependent gene activation in islets. J. Exp. Med. 190: 1135-1146.
- Grilli M, Goffi F, Memo M & Spano P (1996a) Interleukin-1β and glutamate activate the NF-κB/Rel binding site from the regulatory region of the amyloid precursor protein gene in primary neuronal cultures. J. Biol. Chem. 271: 15002-15007.
- Grilli M, Pizzi M, Memo M & Spano P (1996b) Neuroprotection by aspirin and sodium salicylate through blockade of NF-κB activation. Science 274: 1383-1385.
- Guerrini L, Blasi F & Denis-Donini S (1995) Synaptic activation of NF-κB by glutamate in cerebellar granule neurons in vitro. Proc. Natl. Acad. Sci. USA 92: 9077-9081.
- Gveric D, Kaltschmidt C, Cuzner ML & Newcombe J (1998) Transcription factor NF- κ B and inhibitor I κ B α are localized in macrophages in active multiple sclerosis lesions. J. Neuropathol. Exp. Neurol. 57: 168-178.
- Hajra L, Evans AI, Chen M, Hyduk SJ, Collins T & Cybulsky MI (2000) The NF-κB signal transduction pathway in aortic endothelial cells is primed for activation in regions predisposed to atherosclerotic lesion formation. Proc. Natl. Acad. Sci. USA 97: 9052-9057.
- Han Y, Runge MS & Brasier AR (1999) Angiotensin II induces interleukin-6 transcription in vascular

- smooth muscle cells through pleiotropic activation of nuclear factor-κB transcription factors. Circ. Res. 84: 695-703.
- Han Z, Boyle DL, Manning AM & Firestein GS (1998) AP-1 and NF-κB regulation in rheumatoid arthritis and murine collagen-induced arthritis. Autoimmunity 28: 197-208.
- Handel ML, McMorrow LB & Gravallese EM (1995) Nuclear factor-κB in rheumatoid synovium. Localization of p50 and p65. Arthritis Rheum. 38: 1762-1770.
- Hart LA, Krishnan VL, Adcock IM, Barnes PJ & Chung KF (1998) Activation and localization of transcription factor nuclear factor-κB in asthma. Am. J. Respir. Crit. Care Med. 158: 1585-1592.
- Hayashi T, Sekine T & Okamoto T (1993) Identification of a new serine kinase that activates NFκB by direct phosphorylation. J. Biol. Chem. 268: 26790-26795.
- Heck S, Bender K, Kullmann M, Gottlicher M, Herrlich P & Cato AC (1997) IκBα-independent downregulation of NF-κB activity by glucocorticoid receptor. EMBO J. 16: 4698-4707.
- Hegazy DM, O'Reilly DA, Yang BM, Hodgkinson AD, Millward BA & Demaine AG (2001) NFκB polymorphisms and susceptibility to type 1 diabetes. Genes Immun. 2: 304-308.
- Heimberg H, Heremans Y, Jobin C, Leemans R, Cardozo AK, Darville M & Eizirik DL (2001) Inhibition of cytokine-induced NF- κ B activation by adenovirus-mediated expression of a NF- κ B super-repressor prevents β -cell apoptosis. Diabetes 50: 2219-2224.
- Hilliard B, Samoilova EB, Liu TS, Rostami A & Chen Y (1999) Experimental autoimmune encephalomyelitis in NF-κB-deficient mice: roles of NF-κB in the activation and differentiation of autoreactive T cells. J. Immunol. 163: 2937-2943.
- Ho E, Quan N, Tsai YH, Lai W & Bray TM (2001) Dietary zinc supplementation inhibits NFκB activation and protects against chemically induced diabetes in CD1 mice. Exp. Biol. Med. 226: 103-111.
- Hofman FM, Hinton DR, Johnson K & Merrill JE (1989) Tumor necrosis factor identified in multiple sclerosis brain. J. Exp. Med. 170: 607-612.
- Hofmann MA, Schiekofer S, Kanitz M, Klevesath MS, Joswig M, Lee V, Morcos M, Tritschler H, Ziegler R, Wahl P, Bierhaus A & Nawroth PP (1998) Insufficient glycemic control increases nuclear factor-κB binding activity in peripheral blood mononuclear cells isolated from patients with type 1 diabetes. Diabetes Care 21: 1310-1316.
- Hofmann MA, Schiekofer S, Isermann B, Kanitz M, Henkels M, Joswig M, Treusch A, Morcos M, Weiss T, Borcea V, Abdel Khalek AK, Amiral J, Tritschler H, Ritz E, Wahl P, Ziegler R, Bierhaus A & Nawroth PP (1999) Peripheral blood mononuclear cells isolated from patients with diabetic nephropathy show increased activation of the oxidative-stress sensitive transcription factor NF-κB. Diabetologia 42: 222-232.
- Hohjoh H & Tokunaga K (2001) Allele-specific binding of the ubiquitous transcription factor OCT-1 to the functional single nucleotide polymorphism (SNP) sites in the tumor necrosis factor- α gene (TNFA) promoter. Genes Immun. 2: 105-109.
- Igaz P, Falus A, Glaz E & Racz K (2000) Cytokines in diseases of the endocrine system. Cell Biol. Int 24: 663-668.
- Iotsova V, Caamano J, Loy J, Yang Y, Lewin A & Bravo R (1997) Osteopetrosis in mice lacking NF-κB1 and NF-κB2. Nat. Med. 3: 1285-1289.
- Kaltschmidt B, Uherek M, Volk B, Baeuerle PA & Kaltschmidt C (1997) Transcription factor NF- κ B is activated in primary neurons by amyloid β peptides and in neurons surrounding early plaques from patients with Alzheimer disease. Proc. Natl. Acad. Sci. USA 94: 2642-2647.
- Kaltschmidt C, Kaltschmidt B, Neumann H, Wekerle H & Baeuerle PA (1994) Constitutive NF-κB activity in neurons. Mol. Cell. Biol. 14: 3981-3992.
- Kaltschmidt C, Kaltschmidt B & Baeuerle PA (1995) Stimulation of ionotropic glutamate receptors activates transcription factor NF-κB in primary neurons. Proc. Natl. Acad. Sci. USA 92: 9618-9622.
- Karin M (1999) How NF- κB is activated: the role of the I κB kinase (IKK) complex. Oncogene 18: 6867-6874.
- Karin M & Ben-Neriah Y (2000) Phosphorylation meets ubiquitination: the control of NFκB activity. Annu. Rev. Immunol. 18: 621-663.

- Keatings VM, Jatakanon A, Worsdell YM & Barnes PJ (1997) Effects of inhaled and oral glucocorticoids on inflammatory indices in asthma and COPD. Am. J. Respir. Crit. Care Med. 155: 542-548.
- Keifer JA, Guttridge DC, Ashburner BP & Baldwin AS (2001) Inhibition of NF-κB activity by thalidomide through suppression of IκB kinase activity. J. Biol. Chem. 276: 22382-22387.
- Keller JN, Kindy MS, Holtsberg FW, St Clair DK, Yen HC, Germeyer A, Steiner SM, Bruce-Keller AJ, Hutchins JB & Mattson MP (1998) Mitochondrial manganese superoxide dismutase prevents neural apoptosis and reduces ischemic brain injury: suppression of peroxynitrite production lipid peroxidation and mitochondrial dysfunction. J. Neurosci. 18: 687-689.
- Kelliher MA, Grimm S, Ishida Y, Kuo F, Stanger BZ & Leder P (1998) The death domain kinase RIP mediates the TNF-induced NF-κB signal. Immunity 8: 297-303.
- Kieseier BC, Storch MK, Archelos JJ, Martino G & Hartung HP (1999) Effector pathways in immune mediated central nervous system demyelination. Curr. Opin. Neurol. 12: 323-336.
- Kim JK, Kim YJ, Fillmore JJ, Chen Y, Moore I, Lee J, Yuan M, Li ZW, Karin M, Perret P, Shoelson SE & Shulman GI (2001) Prevention of fat-induced insulin resistance by salicylate. J. Clin. Invest. 108: 437-446.
- Knight JC, Udalova I, Hill AV, Greenwood BM, Peshu N, Marsh K & Kwiatkowski D (1999) A polymorphism that affects OCT-1 binding to the TNF promoter region is associated with severe malaria. Nat. Genet. 22: 145-150.
- Kobayashi K, Inohara N, Hernandez LD, Galan JE, Nunez G, Janeway CA, Medzhitov R & Flavell RA (2002) RICK/RIP2/CARDIAK mediates signalling for receptors of the innate and adaptive immune systems. Nature 416: 194-199.
- Kopp E & Ghosh S (1994) Inhibition of NF-κB by sodium salicylate and aspirin. Science 265: 956-959.
- Kuhn R, Lohler J, Rennick D, Rajewsky K & Muller W (1993) Interleukin-10-deficient mice develop chronic enterocolitis. Cell 75: 263-274.
- Kunsch C, Ruben SM & Rosen CA (1992) Selection of optimal κB/Rel DNA-binding motifs: interaction of both subunits of NF-κB with DNA is required for transcriptional activation. Mol. Cell. Biol. 12: 4412-4421.
- Landry DB, Couper LL, Bryant SR & Lindner V (1997) Activation of the NF-κB and IκB system in smooth muscle cells after rat arterial injury. Induction of vascular cell adhesion molecule-1 and monocyte chemoattractant protein-1. Am. J. Pathol. 151: 1085-1095.
- Lavon I, Goldberg I, Amit S, Landsman L, Jung S, Tsuberi BZ, Barshack I, Kopolovic J, Galun E, Bujard H & Ben-Neriah Y (2000) High susceptibility to bacterial infection but no liver dysfunction in mice compromised for hepatocyte NF-κB activation. Nat. Med. 6: 573-577.
- Ley K (2001) Pathways and bottlenecks in the web of inflammatory adhesion molecules and chemoattractants. Immunol. Res. 24: 87-95.
- Li N & Karin M (1999) Is NF-κB the sensor of oxidative stress? FASEB J. 13: 1137-1143.
- Li Q, Van Antwerp D, Mercurio F, Lee KF & Verma IM (1999a) Severe liver degeneration in mice lacking the I_KB kinase 2 gene. Science 284: 321-325.
- Li ZW, Chu W, Hu Y, Delhase M, Deerinck T, Ellisman M, Johnson R & Karin M (1999b) The IKK β subunit of I κ B kinase (IKK) is essential for nuclear factor κ B activation and prevention of apoptosis. J. Exp. Med. 189: 1839-1845.
- Liu ZG, Hsu H, Goeddel DV & Karin M (1996) Dissection of TNF receptor 1 effector functions: JNK activation is not linked to apoptosis while NF-κB activation prevents cell death. Cell 87: 565-576
- Majumdar S & Aggarwal BB (2001) Methotrexate suppresses NF- κ B activation through inhibition of I κ B α phosphorylation and degradation. J. Immunol. 167: 2911-2920.
- Manna SK, Mukhopadhyay A & Aggarwal BB (2000) Leflunomide suppresses TNF-induced cellular responses: effects on NF-κB activator protein-1: c-Jun N-terminal protein kinase and apoptosis. J. Immunol. 165: 5962-5969.
- Marienfeld R, Neumann M, Chuvpilo S, Escher C, Kneitz B, Avots A, Schimpl A & Serfling E (1997) Cyclosporin A interferes with the inducible degradation of NF-κB inhibitors but not with the processing of p105/NF-κB1 in T cells. Eur. J. Immunol. 27: 1601-1609.

- Marok R, Winyard PG, Coumbe A, Kus ML, Gaffney K, Blades S, Mapp PI, Morris CJ, Blake DR, Kaltschmidt C & Baeuerle PA (1996) Activation of the transcription factor nuclear factor-κB in human inflamed synovial tissue. Arthritis Rheum. 39: 583-591.
- Martin T, Cardarelli PM, Parry GC, Felts KA & Cobb RR (1997) Cytokine induction of monocyte chemoattractant protein-1 gene expression in human endothelial cells depends on the cooperative action of NF-κB and AP-1. Eur. J. Immunol. 27: 1091-1097.
- Matsusaka T, Fujikawa K, Nishio Y, Mukaida N, Matsushima K, Kishimoto T & Akira S (1993) Transcription factors NF-IL6 and NF-κB synergistically activate transcription of the inflammatory cytokines interleukin 6 and interleukin 8. Proc. Natl. Acad. Sci. USA 90: 10193-10197.
- Mattson MP, Goodman Y, Luo H, Fu W & Furukawa K (1997) Activation of NF-κB protects hippocampal neurons against oxidative stress-induced apoptosis: evidence for induction of manganese superoxide dismutase and suppression of peroxynitrite production and protein tyrosine nitration. J. Neurosci. Res. 49: 681-697.
- Mengshol JA, Vincenti MP, Coon CI, Barchowsky A & Brinckerhoff CE (2000) Interleukin-1 induction of collagenase 3 (matrix metalloproteinase 13) gene expression in chondrocytes requires p38: c-Jun N-terminal kinase and nuclear factor κB: differential regulation of collagenase 1 and collagenase 3. Arthritis Rheum. 43: 801-811.
- Meyer S, Kohler NG & Joly A (1997) Cyclosporine A is an uncompetitive inhibitor of proteasome activity and prevents NF-κB activation. FEBS Lett. 413: 354-358.
- Miagkov AV, Kovalenko DV, Brown CE, Didsbury JR, Cogswell JP, Stimpson SA, Baldwin AS & Makarov SS (1998) NF-κB activation provides the potential link between inflammation and hyperplasia in the arthritic joint. Proc. Natl. Acad. Sci. USA 95: 13859-13864.
- Mohamed AK, Bierhaus A, Schiekofer S, Tritschler H, Ziegler R & Nawroth PP (1999) The role of oxidative stress and NF-κB activation in late diabetic complications. Biofactors 10: 157-167.
- Morigi M, Angioletti S, Imberti B, Donadelli R, Micheletti G, Figliuzzi M, Remuzzi A, Zoja C & Remuzzi G (1998) Leukocyte-endothelial interaction is augmented by high glucose concentrations and hyperglycemia in a NF-κB-dependent fashion. J. Clin. Invest. 101: 1905-1915.
- Mortensen RF (2001) C-reactive protein inflammation and innate immunity. Immunol. Res. 24: 163-176.
- Murakami S, Lefebvre V & de Crombrugghe B (2000) Potent inhibition of the master chondrogenic factor Sox9 gene by interleukin-1 and tumor necrosis factor-α. J. Biol. Chem. 275: 3687-3692.
- Neish AS, Khachigian LM, Park A, Baichwal VR & Collins T (1995a) Sp1 is a component of the cytokine-inducible enhancer in the promoter of vascular cell adhesion molecule-1. J. Biol. Chem. 270: 28903-28909.
- Neish AS, Read MA, Thanos D, Pine R, Maniatis T & Collins T (1995b) Endothelial interferon regulatory factor 1 cooperates with NF-κB as a transcriptional activator of vascular cell adhesion molecule 1. Mol. Cell. Biol. 15: 2558-2569.
- Neurath MF, Pettersson S, Meyer zum Buschenfelde KH & Strober W (1996) Local administration of antisense phosphorothioate oligonucleotides to the p65 subunit of NF-κB abrogates established experimental colitis in mice. Nat. Med. 2: 998-1004.
- Neurath MF & Pettersson S (1997) Predominant role of NF-κB p65 in the pathogenesis of chronic intestinal inflammation. Immunobiology 198: 91-98.
- Nikolaus S, Raedler A, Kuhbacker T, Sfikas N, Folsch UR & Schreiber S (2000) Mechanisms in failure of infliximab for Crohn's disease. Lancet 356: 1475-1479.
- Norbiato G, Bevilacqua M, Vago T & Clerici M (1997) Glucocorticoids and Th-1: Th-2 type cytokines in rheumatoid arthritis osteoarthritis asthma atopic dermatitis and AIDS. Clin. Exp. Rheumatol. 15: 315-323.
- O'Neill LA & Kaltschmidt C (1997) NF-κB: a crucial transcription factor for glial and neuronal cell function. Trends Neurosci. 20: 252-258.
- Pahan K & Schmid M (2000) Activation of nuclear factor-κB in the spinal cord of experimental allergic encephalomyelitis. Neurosci. Lett. 287: 17-20.
- Pahl HL (1999) Activators and target genes of Rel/NF-κB transcription factors. Oncogene 18: 6853-6866.

- Palombella VJ, Conner EM, Fuseler JW, Destree A, Davis JM, Laroux FS, Wolf RE, Huang J, Brand S, Elliott PJ, Lazarus D, McCormack T, Parent L, Stein R, Adams J & Grisham MB (1998) Role of the proteasome and NF-κB in streptococcal cell wall-induced polyarthritis. Proc. Natl. Acad. Sci. USA 95: 15671-15676.
- Pan J & McEver RP (1995) Regulation of the human P-selectin promoter by Bcl-3 and specific homodimeric members of the NF-κB/Rel family. J. Biol. Chem. 270: 23077-23083.
- Park BH & Park JW (2001) The protective effect of Amomum xanthoides extract against alloxaninduced diabetes through the suppression of NFκB activation. Exp. Mol. Med. 33: 64-68.
- Pierce JW, Read MA, Ding H, Luscinskas FW & Collins T (1996) Salicylates inhibit $I\kappa B-\alpha$ phosphorylation endothelial-leukocyte adhesion molecule expression and neutrophil transmigration. J. Immunol. 156: 3961-3969.
- Poligone B, Weaver DJ, Sen P, Baldwin AS & Tisch R (2002) Elevated NF-κB activation in nonobese diabetic mouse dendritic cells results in enhanced APC function. J. Immunol. 168: 188-196.
- Price SJ, Greaves DR & Watkins H (2001) Identification of novel functional genetic variants in the human matrix metalloproteinase-2 gene: role of Sp1 in allele-specific transcriptional regulation. J. Biol. Chem. 276: 7549-7558.
- Pueyo ME, Gonzalez W, Nicoletti A, Savoie F, Arnal JF & Michel JB (2000) Angiotensin II stimulates endothelial vascular cell adhesion molecule-1 via nuclear factor-κB activation induced by intracellular oxidative stress. Arterioscler. Thromb. Vasc. Biol. 20: 645-651.
- Quan N, Ho E, La W, Tsai YH & Bray T (2001) Administration of NF-κB decoy inhibits pancreatic activation of NF-κB and prevents diabetogenesis by alloxan in mice. FASEB J. 15: 1616-1618.
- Roebuck KA (1999) Regulation of interleukin-8 gene expression. J. Interferon Cytokine Res. 19: 429-438.
- Rogler G, Brand K, Vogl D, Page S, Hofmeister R, Andus T, Knuechel R, Baeuerle PA, Scholmerich J & Gross V (1998) Nuclear factor κB is activated in macrophages and epithelial cells of inflamed intestinal mucosa. Gastroenterology 115: 357-369.
- Ruddle NH (1999) Lymphoid neo-organogenesis: lymphotoxin's role in inflammation and development. Immunol. Res. 19: 119-125.
- Rudolph D, Yeh WC, Wakeham A, Rudolph B, Nallainathan D, Potter J, Elia AJ & Mak TW (2000) Severe liver degeneration and lack of NF-κB activation in NEMO/IKKγ-deficient mice. Genes Dev. 14: 854-862.
- Saccani S, Pantano S & Natoli G (2001) Two waves of nuclear factor κB recruitment to target promoters. J. Exp. Med. 193: 1351-1359.
- Saccani S, Pantano S & Natoli G (2002) p38-Dependent marking of inflammatory genes for increased NF-κB recruitment. Nat Immunol. 3: 69-75.
- Scheinman RI, Cogswell PC, Lofquist AK & Baldwin AS (1995a) Role of transcriptional activation of IκBα in mediation of immunosuppression by glucocorticoids. Science 270: 283-286.
- Scheinman RI, Gualberto A, Jewell CM, Cidlowski JA & Baldwin AS (1995b) Characterization of mechanisms involved in transrepression of NF-κB by activated glucocorticoid receptors. Mol. Cell. Biol. 15: 943-953.
- Schmidt AM, Yan SD, Wautier JL & Stern D (1999) Activation of receptor for advanced glycation end products: a mechanism for chronic vascular dysfunction in diabetic vasculopathy and atherosclerosis. Circ. Res. 84: 489-497.
- Schmitz ML & Baeuerle PA (1991) The p65 subunit is responsible for the strong transcription activating potential of NF-κB. EMBO J. 10: 3805-3817.
- Schmitz ML, Bacher S & Kracht M (2001) IκB-independent control of NF-κB activity by modulatory phosphorylations. Trends Biochem. Sci. 26: 186-190.
- Schreck R, Albermann K & Baeuerle PA (1992) Nuclear factor κB: an oxidative stress-responsive transcription factor of eukaryotic cells (a review). Free Radic. Res. Commun. 17: 221-237.
- Schreiber S, Nikolaus S & Hampe J (1998) Activation of nuclear factor κB inflammatory bowel disease. Gut 42: 477-484.
- Schwartz MD, Moore EE, Moore FA, Shenkar R, Moine P, Haenel JB & Abraham E (1996) Nuclear factor- κ B is activated in alveolar macrophages from patients with acute respiratory distress

- syndrome. Crit. Care Med. 24: 1285-1292.
- Seetharaman R, Mora AL, Nabozny G, Boothby M & Chen J (1999) Essential role of T cell NF-κB activation in collagen-induced arthritis. J. Immunol. 163: 1577-1583.
- Selmaj K, Raine CS, Cannella B & Brosnan CF (1991) Identification of lymphotoxin and tumor necrosis factor in multiple sclerosis lesions. J. Clin. Invest. 87: 949-954.
- Senftleben U, Cao Y, Xiao G, Greten FR, Krahn G, Bonizzi G, Chen Y, Hu Y, Fong A, Sun SC & Karin M (2001) Activation by IKKα of a second evolutionary conserved NF-κB signaling pathway. Science 293: 1495-1499.
- Sha WC, Liou HC, Tuomanen EI & Baltimore D (1995) Targeted disruption of the p50 subunit of NF-κB leads to multifocal defects in immune responses. Cell 80: 321-330.
- Shenkar R & Abraham E (1997) Hemorrhage induces rapid in vivo activation of CREB and NF-κB in murine intraparenchymal lung mononuclear cells. Am. J. Respir. Cell Mol. Biol. 16: 145-152.
- Simeonidis S, Liang S, Chen G & Thanos D (1997) Cloning and functional characterization of mouse IκBε. Proc. Natl. Acad. Sci. USA 94: 14372-14377.
- Smith C, Andreakos E, Crawley JB, Brennan FM, Feldmann M & Foxwell BM (2001) NF- κ B-inducing kinase is dispensable for activation of NF- κ B in inflammatory settings but essential for lymphotoxin β receptor activation of NF- κ B in primary human fibroblasts. J. Immunol. 167: 5895-5903.
- Stein B, Baldwin AS, Ballard DW, Greene WC, Angel P & Herrlich P (1993) Cross-coupling of the NF-κB p65 and Fos/Jun transcription factors produces potentiated biological function. EMBO J. 12: 3879-3891.
- Tanaka M, Fuentes ME, Yamaguchi K, Durnin MH, Dalrymple SA, Hardy KL & Goeddel DV (1999) Embryonic lethality liver degeneration and impaired NF-κB activation in IKK-β-deficient mice. Immunity 10: 421-429.
- Terai K, Matsuo A & McGeer PL (1996) Enhancement of immunoreactivity for NF-κB in the hippocampal formation and cerebral cortex of Alzheimer's disease. Brain Res. 735: 159-168.
- Thanos D & Maniatis T (1995) Virus induction of human IFN β gene expression requires the assembly of an enhanceosome. Cell 83: 1091-1100.
- Thompson AB, Robbins RA, Romberger DJ, Sisson JH, Spurzem JR, Teschler H & Rennard SI (1995a) Immunological functions of the pulmonary epithelium. Eur. Respir. J. 8: 127-149.
- Thompson JE, Phillips RJ, Erdjument-Bromage H, Tempst P & Ghosh S (1995b) $I\kappa B-\beta$ regulates the persistent response in a biphasic activation of NF- κ B. Cell 80: 573-582.
- Tomita T, Takeuchi E, Tomita N, Morishita R, Kaneko M, Yamamoto K, Nakase T, Seki H, Kato K, Kaneda Y & Ochi T (1999) Suppressed severity of collagen-induced arthritis by in vivo transfection of nuclear factor κB decoy oligodeoxynucleotides as a gene therapy. Arthritis Rheum. 42: 2532-2542.
- Tsao PW, Suzuki T, Totsuka R, Murata T, Takagi T, Ohmachi Y, Fujimura H & Takata I (1997)
 The effect of dexamethasone on the expression of activated NF-κB in adjuvant arthritis. Clin. Immunol. Immunopathol. 83: 173-178.
- Udalova IA, Richardson A, Denys A, Smith C, Ackerman H, Foxwell B & Kwiatkowski D (2000) Functional consequences of a polymorphism affecting NF-κB p50-p50 binding to the TNF promoter region. Mol. Cell. Biol. 20: 9113-9119.
- Udalova IA, Mott R, Field D & Kwiatkowski D (2002a) Quantitative prediction of NF-κB DNA-protein interactions. Proc. Natl. Acad. Sci. USA 99: 8167-8172.
- Udalova IA, Richardson A, Ackerman H, Wordsworth P & Kwiatkowski D (2002b) Association of accelerated erosive rheumatoid arthritis with a polymorphism that alters NF-κB binding to the TNF promoter region. Rheumatology 41: 830-831.
- Van Antwerp DJ, Martin SJ, Kafri T, Green DR & Verma IM (1996) Suppression of TNF-α-induced apoptosis by NF-κB. Science 274: 787-789.
- van Heel DA, Udalova IA, De Silva AP, McGovern DP, Kinouchi Y, Hull J, Lench NJ, Cardon LR, Carey AH, Jewell DP & Kwiatkowski D (2002) Inflammatory bowel disease is associated with a TNF polymorphism that affects an interaction between the OCT1 and NF-κB transcription factors. Hum. Mol. Genet. 11: 1281-1289.

- Venkataraman L, Burakoff SJ & Sen R (1995) FK506 inhibits antigen receptor-mediated induction of c-rel in B and T lymphoid cells. J. Exp. Med. 181: 1091-1099.
- Wahl C, Liptay S, Adler G & Schmid RM (1998) Sulfasalazine: a potent and specific inhibitor of nuclear factor κB. J. Clin. Invest. 101: 1163-1174.
- Walsh DM, Klyubin I, Fadeeva JV, Cullen WK, Anwyl R, Wolfe MS, Rowan MJ & Selkoe DJ (2002) Naturally secreted oligomers of amyloid β protein potently inhibit hippocampal long-term potentiation in vivo. Nature 416: 535-539.
- Wang CY, Mayo MW & Baldwin AS (1996) TNF- and cancer therapy-induced apoptosis: potentiation by inhibition of NF-kB. Science 274: 784-787.
- Wang D, Westerheide SD, Hanson JL & Baldwin AS (2000) Tumor necrosis factor α -induced phosphorylation of RelA/p65 on Ser529 is controlled by casein kinase II. J. Biol. Chem. 275: 32592-32597.
- Weaver DJ, Poligone B, Bui T, Abdel-Motal UM, Baldwin AS & Tisch R (2001) Dendritic cells from nonobese diabetic mice exhibit a defect in NF-κB regulation due to a hyperactive IκB kinase. J. Immunol. 167: 1461-1468.
- Weih F, Durham SK, Barton DS, Sha WC, Baltimore D & Bravo R (1997) p50-NF-κB complexes partially compensate for the absence of RelB: severely increased pathology in p50^{-/-}relB^{-/-} double-knockout mice. J. Exp. Med. 185: 1359-1370.
- Weingarten P, Bermak J & Zhou QY (2001) Evidence for non-oxidative dopamine cytotoxicity: potent activation of NF-κB and lack of protection by anti-oxidants. J. Neurochem. 76: 1794-1804.
- Welch GN & Loscalzo J (1998) Homocysteine and atherothrombosis. N. Engl. J. Med. 338: 1042-1050.
- Whiteside ST, Epinat JC, Rice NR & Israel A (1997) $I\kappa B\epsilon$, a novel member of the $I\kappa B$ family, controls RelA and cRel NF- κB activity. EMBO J. 16: 1413-1426.
- Whitley MZ, Thanos D, Read MA, Maniatis T & Collins T (1994) A striking similarity in the organization of the E-selectin and β interferon gene promoters. Mol. Cell. Biol. 14: 6464-6475.
- Wilckens T & De Rijk R (1997) Glucocorticoids and immune function: unknown dimensions and new frontiers. Immunol. Today 18: 418-424.
- Wilson SH, Caplice NM, Simari RD, Holmes DR, Carlson PJ & Lerman A (2000) Activated nuclear factor- κ B is present in the coronary vasculature in experimental hypercholesterolemia. Atherosclerosis 148: 23-30.
- Woodroofe MN & Cuzner ML (1993) Cytokine mRNA expression in inflammatory multiple sclerosis lesions: detection by non-radioactive in situ hybridization. Cytokine 5: 583-588.
- Wulczyn FG, Naumann M & Scheidereit C (1992) Candidate proto-oncogene bcl-3 encodes a subunitspecific inhibitor of transcription factor NF-κB. Nature 358: 597-599.
- Xiao G, Harhaj EW & Sun SC (2001) NF-κB-inducing kinase regulates the processing of NF-κB2 p100. Mol. Cell 7: 401-409.
- Yan SD, Yan SF, Chen X, Fu J, Chen M, Kuppusamy P, Smith MA, Perry G, Godman GC, Nawroth P, Zweiter JL & Stern D (1995) Non-enzymatically glycated tau in Alzheimer's disease induces neuronal oxidant stress resulting in cytokine gene expression and release of amyloid βpeptide. Nat. Med. 1: 693-699.
- Yang JP, Merin JP, Nakano T, Kato T, Kitade Y & Okamoto T (1995) Inhibition of the DNA-binding activity of NF-κB by gold compounds in vitro. FEBS Lett. 361: 89-96.
- Yang L, Cohn L, Zhang DH, Homer R, Ray A & Ray P (1998) Essential role of nuclear factor κB in the induction of eosinophilia in allergic airway inflammation. J. Exp. Med. 188: 1739-1750.
- Yin MJ, Yamamoto Y & Gaynor RB (1998) The anti-inflammatory agents aspirin and salicylate inhibit the activity of IκB kinase-β. Nature 396: 77-80.
- Zhong H, SuYang H, Erdjument-Bromage H, Tempst P & Ghosh S (1997) The transcriptional activity of NF-κB is regulated by the IκB-associated PKAc subunit through a cyclic AMP-independent mechanism. Cell 89: 413-424.
- Zollner TM, Podda M, Pien C, Elliott PJ, Kaufmann R & Boehncke WH (2002) Proteasome inhibition reduces superantigen-mediated T cell activation and the severity of psoriasis in a SCID-hu model. J. Clin. Invest. 109: 671-679.

Zong WX, Edelstein LC, Chen C, Bash J & Gelinas C (1999) The prosurvival Bcl-2 homolog Bfl-1/A1 is a direct transcriptional target of NF- κ B that blocks TNF α -induced apoptosis. Genes Dev. 13: 382-387.

6. AFFILIATIONS

Imperial College of Science, Technology and Medicine, Faculty of Medicine, Kennedy Institute of Rheumatology, 1 Aspenlea Road, Hammersmith, London W6 8LH, UK (E. Andreakos, S. Sacre & B.M. Foxwell); The John Radcliffe Hospital, University Department of Paediatrics, Headley Way, Headington, Oxford OX3 9DU, UK (I. Udalova).