Cellular and Humoral Markers of Tissue Damage

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Introduction

Patients with sepsis who are hospitalized in intensive care units frequently develop multiple organ dysfunction failure syndrome (MODFS) with a poor prognosis (1-3). It is believed that injury in MOFDS is brought about by factors which under normal conditions play a regulatory role in homeostatic mechanisms (Table 1).

Table 1.	Factors	involved	in	septic	shock
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Cytokines
Growth factors
Adhesins
CD antigens
Lipid mediators
Gas mediators
Franscription factors
Dncogenes
Genes

Some of these factors are humoral like cytokines, growth factors, lipid and gas mediators while others like oncogenes and their protein products, transcription factors (i.e. nuclear factor kappa B, NF κ B) or DNA nuclear material are widely recognized as cellular ones. Adhesins might be viewed both as cellular (or membrane-bound) and soluble (humoral) factors. The distinction between cellular and humoral factors tends to be more and more vague.

A widely used classification for the cluster of differentiation (CD) antigens involves not only cellular markers but also some cytokines and their receptors in membrane-bound and soluble forms (4). The best example of the dual role of cellular markers is class I major histocompatibility antigens (MHC or HLA) (5). These were first recognized as membrane-bound molecules on all nucleated cells, and the existence of soluble HLA antigens has also been reported. HLA antigens are known markers of severe brain damage or of the onset of transplant rejection (5, 6).

Many of the mediators associated with MODFS are generated by macrophages which being maximally stimulated already escaped physiological regulatory mechanisms and are genetically recoded to their death. This state is known as programmed cell death or apoptosis (7, 8). At a cellular level apoptosis and necrosis are responsible for the release of cellular markers to the body fluids. Apoptosis regulates cell number and eliminates damaged or infected cells. Also shedding off the cell-bound molecules from macrophages, or from other cells with the fast membrane turnover might be a source of soluble forms of cellular markers.

Baue and Faist suggested the following stages of development of MODFS: infection, increased permeability of intestinal mucosa, involvement of immune system (macrophages, cytokines, antibodies, receptors); generalized inflammation associated with a damage to endothelium, oedema and impaired oxygen availability; and finally ischaemia with reduced microcirculatory flow, necrosis and eventually MODFS (2).

This systemic inflammatory response syndrome (SIRS) caused by infection is characterized by the exacerbation of the production of pro- and anti-inflammatory mediators (9). The cascade of cellular and humoral factors (Table 1) released by endotoxin (LPS) includes thromboxane A_2 (TXA₂), nitric oxide (NO), oxygen free radicals (O₂-, OH'), lipid peroxides (LOO'), leukotrienes (LTB₄-E₄), platelet-activating factor (PAF), tumour necrosis factor (TNF α), interleukins (IL-1, IL-4, IL-6, IL-8, IL-10, IL-12, IL-13), interferons α and γ (IFN α , IFN γ), cytoadhesins (ELAM, ICAM, sPAGEM), soluble TNF (sTNF R) and IL-1 (IL-1ra) receptors, transforming growth factor β (TGF β) and many others.

Although in many animal models of septic shock removal of the above mediators has conferred successful protection, so far very poor results have been achieved in humans. Current understanding of the setting of an antiinflammatory response by the host seems to indicate that the host organism is responsible for putting into motion most of the necessary regulatory processes. Indeed, both pro- and anti-inflammatory mediators appear to be markers of the severity of the disease. However, after exhaustion of adaptive mechanisms, therapeutic help in SIRS is required.

Various therapeutic approaches have been proposed to interfere with the development of SIRS (9-11). These include antibodies to: endotoxin (LPS), anti-IL-1, anti-IL-8, anti-IL-12, anti-CD14, anti-73kDa LPS receptor, anti-IFN γ , monoclonal antibodies to TNF α , macrophage migration inhibitory factor (MIF) and lymphocyte migration inhibitory factor (LIF), anti-CD11b, anti-ICAM-1, anti-ELAM-1, anti-superoxide dismutase, IL-1 receptor antagonists, sTNF R, IL-1 β converting enzyme inhibitors (ICE), aprotinin, hirudin, heparin, platelet

activating factor (PAF) antagonists, cyclooxygenases (COX-1 and COX-2) inhibitors (nonsteroidal anti-inflammatory drugs, NSAID), steroid hormones, lipooxygenase inhibitors and NO synthase (NOS) inhibitors, prostacyclin analogues, tyrosine kinase inhibitors, lipid X, lipoaminoacids, as well as infusions of various electrolytes or buffers containing catecholamines, vasopressin, angiotensin II or sometimes phenoxybenzamine (10, 12-22). A relapse of time between an infection and administration of a putative drug is crucial for its efficacy. Lack of success in the treatment of MODFS with curative molecules which were effective in the experimental models of septic shock might be explained, at least partially, by a late stage of SIRS at which the therapy is usually implemented (9).

Out of a broad scope of tissue damage markers which are important in MODFS we will focus on: i) LPS/ceramide CD14 system in the induction of septic shock, ii) cytokines - inhibition of their production and cytokine-binding proteins, iii) serum HLA class I antigens, iv) lipid mediators, v) NO as a marker of septic shock and vi) apoptosis.

LPS/ceramide CD14 system

Most cases of sepsis and septic shock are secondary to infections with Gram-negative bacteria (23, 24). The Gram-negative bacterial cell wall consists of inner and outer membranes, the latter portion of which contains proteins as well as LPS (14, 25). It is useful to review briefly the structure of LPS. LPSs are complex molecules composed of three major parts: a polysaccharide side chain (O-antigen), which is attached via a bridging (core) polysaccharide to a glucosamine-based phospholipid (lipid A). The most variable part of the LPS structure is the O antigen. In contrast, the lipid A and to some extend also core region are more conserved (26). This is why antibodies are produced which may cross-react.

Endotoxin is measured utilizing the haemolymph of amoebocytes from the Limus horseshoe crab (27). This substance contains a proenzyme which is directly activated by endotoxin leading to visible gelling of the mixture. Spectrophotometric modification of the Limus amoebocyte lysate (LAL) assay detects endotoxin to less than 10 pg/dl. Unfortunately, the accuracy of the LAL assay is affected by numerous circulating plasma proteins (i.e. antithrombin III, anti-endotoxic antibodies, etc.) and it can be positive in patients with Gram-positive bacterial and fungal infections (28). This may be related to the gut leak which accompanies these infections. Measurement of LPS levels in diagnostic practice shows that endotoxin is often present in sepsis; its presence might correlate with the severity of clinical manifestations and end-organ dysfunction. However, the same literature consistently finds out that endotoxaemia, as detected by LAL assay, is not invariably present in clinical sepsis as the results of LAL assay must be viewed with scepticism (14). In clinical practice, useful markers that may indicate the presence of Gram-negative bacteria sepsis are: blood culture for Gram-negative microbe or culture or Gram stain positive for a Gram-negative microbe at a local site of infection (26).

It has been shown that nerve growth factor (NGF), TNF α , IL-1 β or Fas ligand after binding to their specific CD40 transmembrane receptors stimulate cells by releasing the intracellular messenger ceramide (29-32). This stimulation is completed through activating neutral sphingomyelinases. Agonist-induced hydrolysis of sphingomyelin puts in motion the sphingomyelin cycle analogous to the signal transduction by the phosphatidyloinositol and glycerophospholipid system (33).

Recent studies on the composition of the outer leaflet of the outer membrane of some Gram-negative bacteria emphasize the presence of a glycosphingolipid but not the LPS. This bacterium has been named *Sphingomonas* (34). The substitution of sphingolipids for LPS in this bacterium confirms that the physical properties of sphingolipids may be interchangeable with those of LPS. Indeed, the examination of the structure of LPS and ceramide have revealed a strong similarity between their molecules. Computer molecular modelling and conformational dynamics yielded a solution structure for the acylated glucosamine I of LPS with a strong similarity to that of ceramide (35). Identical chirality at two optical active carbon centres and approximately similar lengths of the hydrocarbon chains make the molecules of LPS and ceramide very much alike. LPS along with TNF and IL-1 cause common events in cells such as: (i) mitogen-activated protein kinase activation; (ii) NF- κ B translocation; (iii) activator protein 1 stimulation; (iv) phospholipase A₂ activation and (v) TNF gene expression (35).

For recognition of endotoxin, a binding protein/receptor system involving LPS-binding protein (LBP) and CD14 molecule has been postulated. However, effects of LPS occur also in CD14-negative cells, and not all of them depend on the presence of LPB. This is why not all pathways in recognition of endotoxin are already defined. Several cellular structures have been found to bind LPS. For instance, the scavenger receptor or CD11/CD18 are involved in the detoxification of LPS (36). Several not well characterized membrane proteins with 18, 25, 38, 55, and 65 kDa were found to bind LPS. Also, 40- and 80 kDa, as well as 70- to 80 kDa proteins have been postulated in LPS recognition by various cell types (25). The CD14 molecule, however, has been unequivocally established to be a LPS receptor (37). CD14 is found as a 53 kDa glycoprotein on the cell surface (mCD14) of all mature myeloid cells. The gene for CD14 is located on the fifth chromosome in a region known to encode several cytokines including granulocyte/macrophage colony stimulating factor (GM-CSF), CSF-1, IL-3, endothelial cell growth factor (ECGF), as well as receptors like CSF-1 receptor, platelet-derived growth factor receptor, FMS (c-fms photooncogen) and B-adrenergic receptor. The 32 identified amino acid residues have 40 kDa and almost completely match the amino acid sequence deduced from the CD14 cDNA. In the plasma of healthy adults 4-6 μ g/ml of soluble CD14 (sCD14) are

found and sCD14 is highly elevated up to 200 μ g/ml in the plasma of septic patients. Two slightly different soluble forms of CD14 molecules exist which can be found in normal serum, most likely due to shedding of mCD14 as it was shown for endothelial cells (38).

Endotoxins or LPS are undoubtedly the molecules responsible for most of the pathophysiological phenomena associated with Gram-negative infections. Experimental and clinical observations indicate that endotoxin may exert its deleterious effects upon the host to a large part by provoking the release of a variety of endogenous mediators, although direct toxicity may also occur. In 1968, Chedid et al. proposed and used an antibody capable of neutralizing endotoxins. In 1982, Ziegler et al. used antiserum prepared in healthy volunteers injected with J5 LPS of rough mutant of Escherichia coli and substantially reduced deaths from bacteraemia (10, 11, 14). Modern technology has enabled researchers to prepare monoclonal antibodies (mAb). Many mAbs have been generated and studied. The most well known are those which reached the stage of clinical studies. E5 mAb from Xoma Corporation and HA.1A mAb from Centocor were reported to be effective for patients with sepsis and Gram-negative bacteraemia (39, 40). The neutralizing therapy is extremely expensive and the results of double blind randomized placebo controlled multicentre trials are indicative but not conclusive. The current limitations of treatment of septic patients with anti-LPS antibodies, beside the not trivial financial impact, are connected with the fact that an increasing proportion of cases of sepsis are related to infection with Gram-positive bacteria; it is difficult to demonstrate cross-reactivity against rough mutants and so far neutralization of the effect of endotoxin by antibodies has not been described. The theoretical premise that LPS antibodies are cross-protective is attractive but requires further investigation (14).

Soluble CD14 can act as an inhibitor at the monocyte/macrophage level. In contrast, sCD14 is involved in the activation of endothelial cells by LPS (41). This observation limits the therapeutic use of sCD14 but leaves room for the use of anti-CD14 antibodies. It was reported that the anti-CD14 strategy reduced hypotension, lowered cytokine production and prevented pulmonary oedema.

Proinflammatory cytokine in sepsis

While the initiating event in sepsis may be the release of endotoxin, many of the clinical symptoms of sepsis result from the release of endogenous mediators such as TNF, IL-1, IL-6, IL-8, PAF etc., mononuclear phagocytes and other cells, including endothelial cells. Each of these mediators stimulates both its own release and the release of other mediators and acts in concert to produce symptoms of sepsis. The proinflammatory cytokines TNF α and IL-1 β have been studied extensively. It is generally accepted that these cytokines play a central role in the pathogenesis of sepsis (13, 19, 20, 42).

Plasma levels of TNF are a marker of sepsis and depend on the severity of disease. After bolus injection of endotoxin, blood levels of TNF α typically increase at 30 min and peak at 90 min (9, 19, 20). TNF is known to stimulate not only the release of several other cytokines (e.g. IL-1, IL-6, PAF), but also the production of endothelial adhesion molecules which promote neutrophilendothelial cell adherences. Likewise, TNF enhances neutrophil phagocytosis and injures endothelial cells as manifested by an increase of endothelial cell permeability. TNF is rapidly cleared from the systemic circulation due to its short half-life (14-18 min in humans) and returns to base levels within a few hours (43). This is why plasma TNF activity alone is not a reliable prognostic factor in septicaemia and supports the concept that $TNF\alpha$ is an early mediator that initiates the changes that lead to the extensive cellular injury (44). However, the strongest evidence supporting the role of the TNF in the pathogenesis of septic shock, SIRS and MODFS comes from studies that employed anti-TNF antibodies (10). The favourable effect of anti-TNF antibodies in experimental septic shock were most prominent when antibody was administered prior to the infusion of LPS. Based on these observations, it is clear that for TNF antibodies to be effective, the antibody must be given before or very soon after the onset of bacterial infection (14).

The polypeptide hormone IL-1 is another important factor in host defence and exists in two forms: IL-1 α and IL-1 β . Both are potent proinflammatory monokines with biological effects that include endothelial cell activation and increase of adhesion molecule receptor expression. In addition, IL-1 promotes the release of other cytokines (TNF, IL-6, PAF) and acts synergistically with TNF in the production of many of its biological and inflammatory effects such as hypotension, endothelial cell injury, increased vascular permeability and, finally, death (45). Like TNF, serum levels of IL-1 rise after endotoxin infusion. However, in contrast with TNF, serum levels of IL-1 reach their peak 3-4 h after LPS challenge (20).

IL-6 is another cytokine in the inflammatory network. Expression of IL-6 is induced in many cells, including mononuclear phagocytes and endothelial cells, after stimulation with LPS, IL-1 and TNF. IL-6 is thought to promote neutrophil activation and accumulation at sites of inflammation (46). In keeping with these properties of IL-6, increased plasma concentrations of IL-6 have been detected in patients with sepsis and are associated with increased mortality. For that reason, most authors now agree that IL-6 is a marker of the severity of the infection (45, 47, 48).

IL-8 is a recently described peptide that is secreted by a variety of cells such as alveolar macrophages, monocytes, endothelial cells in response to endotoxin, IL-1 and TNF (15, 49). IL-8 causes chemoattraction and activation of neutrophils and is believed to mediate neutrophil recruitment in host defence and disease (50-52). It was also demonstrated that IL-8 enhances binding affinity of adhesion molecules on human neutrophils (53, 54). The accumulation of activated neutrophils in lungs and other organs is felt to play a key role in the pathogenesis of SIRS and MODFS (55, 56).

An important first step in the process of neutrophil-mediated organ injury involves the binding of neutrophils to endothelial cells (57, 58). This interaction is largely regulated by complementary adherence molecules that are present on these cells and expressed in increasing numbers in response to endotoxin, TNF, IL-1, and IL-8. Endothelial and neutrophil cell activation by these cytokines is accompanied by enhanced expression of adhesin molecules such as ELAM1, VCAM1, ICAM1 (59). Anti-CD18 antibodies, in contrast to anti-CD11 antibodies, are able to reduce LPS-induced neutrophil sequestration in tissue and organ injury (59, 60). This antibody worsened endotoxaemia, acidosis and cardiovascular function in a canine model of LPS shock and in the baboon model of sepsis. In the same model, anti-ELAM1 therapy was beneficial. The contribution of the adhesion molecule and the interaction between endothelial and circulating cells play a major role in tissue damage and organ dysfunction. However, the use of antibodies to interfere in this process remains controversial (13).

Cytokine-binding proteins

Cytokine-binding proteins (CBPs), such as cytokine receptors and antibodies, mostly monoclonal, impair interactions of cytokines with their cellular receptors and so these agents can potentially provide a means for treating pathological conditions that have a significant cytokine involvement as is the case in septic shock. Indeed, the efficacy of such treatment was demonstrated for the first time a decade ago whereby experimental shock was prevented by antibodies against TNF α (61, 62). The cloning of genes encoding cytokine receptor chains, and the characterization of their soluble forms, has opened the way to new strategies in anticytokine therapy. These molecules clearly act as antagonists of their respective ligands through competition with the membrane receptors that transduce the biological signal into the target cell. As anticipated, the injection of sIL-1R modulate the allogenic response in vitro and prevent allograft rejection (63). Cerami's group and many others have confirmed the protective activities of anti-TNF antibodies in various models. Also in humans, anti-TNF antibodies were effective when combined with antibiotics (10, 14, 61). Counteracting the effect of another proinflammatory cytokine, IL-1 has been investigated with the IL-1 receptor antagonist (IL-1-ra) (64-66). This natural IL-1-like molecule binds to the same receptors as IL-1, but fails to transmit any signal (61). A second phase III study of IL-1ra was dropped after an interim analysis had failed to show any evidence of benefit (10). Another possibility for containing IL-1 is the use of IL-1 β converting enzyme (ICE) inhibitors. ICE is responsible for cleaving a biologically inactive IL-1 precursor into the mature IL-1 active form (67). The results of the INTERSEPT placebo-controlled trial of anti-TNF mAb

(Bayx1351) in 563 patients with severe sepsis shows lack of significantly altered mortality between the studied groups (68).

In contrast, the results of the Immunex study of p75 sTNFR-Fc which have now been reported indicate an increase of mortality among patients in the treated group. In patients with sepsis, plasma levels of both sTNFR p55 and p75 are markedly increased and highly correlate with simultaneously obtained APACHE II and MODFS scores. Since the degree of increased sTNFR levels correlated poorly with patient survival, elevated sTNFR levels represent a good marker for severity of sepsis and predict an outcome (69).

However, exceptions do exist: soluble IL-6 receptor and cilliary neurotrophic factor act as agonists (70). A potential advantage of soluble receptors over high affinity mAbs is that they are of human origin and, accordingly, the problem of patient immunization is obviated. There are two drawbacks of soluble receptors: (i) their usually lower affinity than that of mAb and (ii) their shorter half-life in vivo because they are molecules smaller than antibodies. To overcome these problems, for instance, immunoadhesins have been generated that comprise two soluble receptor fragments linked genetically to a human immunoglobulin constant region (i.e. sTNFR-Fc) (71-74). Additionally, this procedure decreases the therapeutic dose of soluble CBPs in vivo. However, the major drawback of using CBPs relates to the fact that they stabilize the cytokine in the form of a cytokine-CBP complex in vivo (61). The first demonstration that CBPs are capable of stabilizing cytokines was provided by treatment with anti-IL-6 mAbs. The longer in vivo half-life of IL-6-anti-IL-6 complexes (3.5 days) provides a pharmacokinetic explanation for the accumulation of cytokine (the half-life of free IL-6 is as low as 20 min) and for their potential action as agonists (62). However, when CBPs are present in excess over the cytokine-membrane receptor, they seem to act as antagonists.

Inhibition of proinflammatory cytokine production

Rather than counteracting cytokines already generated, inhibition of their production may prevent these mediators from becoming involved in the immunoinflammatory cascade. Glucocorticoids were thought to do it; however, using them in sepsis has not been successful (13). Pentoxyphylline, a phosphodiesterase inhibitor, limits the synthesis of TNF (75). Various drugs which reduced TNF and/or IL-1 production (e.g. linomide, prostacyclin analogues or chlorpromazine) also had beneficial effects in experimental models of septic shock (76-78). More recently, it has been shown that tyrosine kinase inhibitors block LPS-induced TNF and NO production (79).

Cytokines such as IL-4, IL-10, IL-13 and TGF β possess anti-inflammatory properties because they inhibit the generation of the most of proinflammatory cytokines in monocytes/macrophages. Moreover, these particular cytokines also

induce IL-1-ra (13). Among other putative beneficial cytokines is IFN α (80). Its usefulness in septic shock is still controversial. Maybe the natural balance between pro- and anti-inflammatory cytokines is not sufficient to slow down the running inflammatory cascade.

Serum HLA class I antigens

More than 20 years ago, MHC class I antigens were reported to be present in serum. The development of anti-HLA mAbs had a significant impact on the analysis of serum HLA class I antigens. Similar to their cell-membrane associated counterpart, the serum HLA class I molecular complex comprises a polymorphic heavy α -chain noncovalently associated with β_2 microglobulin. The level of serum HLA class I antigens markedly increases in the course of viral infections caused by cytomegalovirus, hepatitis B virus, hepatitis C virus, varicella-zostervirus, and human immunodeficiency-1 virus. During HIV-1 infection, the level of serum HLA class I antigens correlates with stages of disease and represents a good prognostic marker of the disease progression. An increase in the level of total serum HLA class I antigens has also been observed in recipients of heart, kidney or liver transplants. The rapid decrease in the level of serum HLA antigens observed following immunosuppressive therapy of acute episodes of graft rejection suggests that their level may be the result of immune system activation. The elevation of donor-derived serum HLA class I allospecificities precedes the clinical evidence of a graft rejection episode. It means that measurement of donor-derived serum HLA antigens may represent a test to diagnose graft rejection episodes (5, 6, 81, 82).

Lipid mediators

Many other mediators produced by activated cells contribute to the inflammatory syndrome. Some may be directly induced by LPS while others may be induced following target cell activation by proinflammatory cytokines. As a consequence, prostaglandins (PGs), thromboxane, leukotrienes and PAF are other potential targets for therapeutic approach (13, 21). Inhibition of PG formation by COX-2 rather than COX-1 inhibitors (e.g. ibuprofen) attenuate many alterations associated with LPS injection in animal models of endotoxin shock and inhibit TNF production. However, the experiments in human volunteers show that an injection of ibuprofen immediately before administration of endotoxin caused a significant increase in the level of circulating TNF. Prostaglandin E_1 (PGE₁) has several properties that could be beneficial for the treatment of severe sepsis. It is a potent vasodilator of the pulmonary and systemic circulation. Like prostacyclin, it has also important anti-inflammatory effects by blocking macrophage activation. It can influence coagulation by inhibiting platelet

aggregation and by inducing fibrynolysis. In dogs, PGE_1 almost entirely restored tissue oxygen extraction after endotoxin challenge (83). PAF antagonists have led to successful protection in various septic shock models induced by LPS. Recent study in patients with sepsis has shown that BN52021 (a PAF inhibitor) offered an improvement (10, 13). Another PAF antagonist, TCV-309, inhibited cytokine production in experimental endotoxaemia in chimpanzees (84).

Nitric oxide as a marker of septic shock

The free radical of NO is synthesized from L-arginine by a family of enzymes -NO synthases (NOS) (85). The continuous biosynthesis of NO by the constitutive endothelial isoform of NOS (eNOS) keeps the vasculature in active vasodilatation and reduces platelets and polymorphonuclear cell adhesion to the endothelium. The inducible isoform of NOS (iNOS) is expressed in response to immunological stimuli. iNOS produces NO at nanomolar amounts and then acts as a cytostatic and cytotoxic agent.

The role of NO in septic shock will be presented in a separate contribution and this is why only a very short summary of available data will be presented here. It has been suggested that the overproduction of NO is responsible for death during endotoxic shock or sepsis (86-91). In 1989, Vallance et al. showed that inhibition of NO synthesis by NG-monomethyl-L-arginine (L-NMMA) had elevated blood pressure in rats (92). Furthermore, it was demonstrated that local infusion of L-NAME into brachial artery of healthy volunteers caused a dosedependent fall in resting forearm blood flow and attenuated the dilator response to acetylcholine. The effect of L-NAME was stereospecific (D-NAME was ineffective) and reversed by supplying an excess of L-arginine (93). Ochoa et al. found elevated nitrite/nitrate levels (stable breakdown products of NO) in patients with septic shock but these levels were decreased in patients with trauma (94, 95). Recently, Wang and Chaudry pointed out the complexity of the alteration in NO production with the progression of sepsis (91). They hypothesized that NO inhibition under some septic conditions might be detrimental. We also observed ambiguous effects of NOS inhibitors in rats with LPS-induced shock. When NOS inhibitors had been administered before LPS was given, the removal of NO potentiated LPS-induced shock. Administration of NO inhibitors in the second hour after injection of LPS caused a temporary improvement. The activity of eNOS is regulated by calmodulin and changes in intracellular calcium. The enhanced formation of NO in the early stage of shock is due to the activation of eNOS. We observed the induction of iNOS mRNA in the lungs, spleen and heart of LPS-treated rats as early as 1 h after endotoxin administration. This is why we postulate that in early and late stages of shock, differential effects of nonselective NOS inhibitors might account for their action on different isoforms of NOS. So far, the role of NOS inhibitors in the treatment of sepsis remains to be defined. The availability of specific inhibitors of iNOS would help to answer the relevant questions. Such agents are in the early stages of development but they have not as yet been studied in humans (96, 97).

Programmed cell death - apoptosis

Apoptosis constitutes an efficient system in cell biology designed to eliminate superfluous, unwanted, altered, aged, or transformed cells without eliciting damage to adjacent normal cells or surrounding tissues. The mechanism of apoptosis has long been neglected in clinical research and in clinical thinking. Nevertheless, apoptosis offers understanding of a number of pathological syndromes and clinical observations which otherwise cannot be explained by well-known biological processes. Leucocytes, monocytes and macrophages are selectively eliminated from inflammatory tissues by the occurrence of programmed cell death. The therapeutic regulation of apoptosis during and after an inflammation offers a new approach for promoting rapid healing and reduction of unwanted pathological sequelae of inflammation processes (7, 8, 98, 99).

Necrosis, or accidental cell death, occurs in response to harmful insults such as physical damage, hypoxia, hyperthermia, complement attack or chemical injury. Table 2 shows the differences between apoptosis and necrosis.

Analysis of the mechanism that prevents cell death, such as activation of the bcl-2 gene, addition of growth factors and the use of protein synthesis inhibitors or calcium entry blockers, might aid in the development of new treatment strategies. Drug and therapy designs directed at the modulation of apoptotic process will offer new opportunities for the treatment and control of tissue damage in the coming years. If these goals can be accomplished, we may finally see a reduction in the morbidity and mortality associated with MODFS.

Conclusion

Comprehension of the basic mechanisms involved in the host inflammatory response is necessary for clinicians to make educated choices and decisions regarding therapies. The discovery of the sphingomyelin cycle as a target of LPS action that mimics natural mediator ceramide promoted progress in our understanding of the events during the development of septic shock. Advances in molecular biology and in the cytokine network led to the development of novel approaches to the treatment of septic shock, SIRS and MODFS. As noted in this review, therapies are directed to distinct levels: the initiating event (i.e. endotoxin), to various mediators, and the effector cells (i.e. macrophages, endothelial cells). These factors are not merely markers of sepsis and its severity,

	Apoptosis	Necrosis	
Origin	Lack of growth factors,	Anoxia, starvation,	
	hormonal factors, mild	physical and chemical	
	toxic stimuli	damage	
Occult phase	Minutes to hours	None	
First manifestation	Shrinking	Swelling	
Nuclear changes	Pyknosis, condensation,	Karyolysis	
	internucleosome		
	cleavage, DNA laddering		
Chromatin	Segmentation and	Nuclear folding	
Nuclealar shoreas	margination Intact	Granulated	
Nucleolar changes	Persists	Failure	
Membrane integrity			
Surface	Smoothing	Lysis, blebbing	
Cytoskeleton	Formation of apoptotic bodies	Fragmentation	
Mitochondria	Unaffected	Swelling	
Endoplasmic reticulum	Unaffected Dilated		
and Golgi apparatus			
Organelles	Intact	Swollen, leaky	
Gene expression	p53 ↑ , bcl-2 \downarrow , c-myc ↑	No change	
Protein synthesis	Blocked by	Not affected by	
	cycloheximide and	antibiotics	
	actinomycin D		
Cytoplasmic changes	Endonuclease activity \uparrow ,	Release of lysosome	
	transglutaminase ↑	content	
Cells affected	Dispersed cells	Diffuse degradation	
Cell elimination	Engulfment by	Inflammatory response	
	macrophages and	in adjacent tissues	
	endothelial cells		

Table 2

but play an important role in the pathophysiological mechanisms. The era of cytokine response modification in patients with severe sepsis has evolved rapidly. These therapies if they prove effective in clinical trials are in progress and it should be stressed that current interventional capacities are far ahead of our comprehension of the mechanisms involved. Moreover, we should remember that these therapies are enormously expensive. Finally, taking into account that apoptosis is a natural route of macrophage elimination, some present therapies might be ineffective because they act on cells already programmed to their death, no matter how much effort is engaged to keep them alive.

References

- 1. Barton R, Cerra FB (1989) The hypermetabolism. Multiple organ failure syndrome. Chest 96:1153-1160
- 2. Baue AE (1994) Organ dysfunction (MODS), organ failure (MOF) and therapeutic conundrums in injured and septic patients. In: Gullo A (ed) Sepsis and organ failure. Fogliazza editore, Trieste, pp 9-39
- 3. Baue AE (1992) The horror autotoxicus and multiple-organ failure. Arch Surg 127:1451-1462
- Schlossman SF, Boumsell L, Gilks W, Harlan JM, Kishimoto T, Morimoto C, Ritz J, Shaw S, Silverstein RL, Springer TA, Tedder TF, Todd RF (1993) CD antigens 1993. Immonol Today 15:98-99
- Puppo F, Scudeletti M, Indiveri F, Ferrone S (1995) Serum class I antigens: markers and modulators of an immune response? Immonol Today 16:124-127
- Ferrone S, Yamamura M, Grosse-Wilde H, Pouletty P (1992) Summary of serum-soluble HLA class I antigen component. In: Kimiyoshi Tsuji MA (ed) HLA 1991. Oxford, New York, pp 1057-1061
- 7. Haanen C, Vermes I (1995) Apoptosis and inflammation. Mediators of Inflammation 4:5-15
- 8. Vermes I, Haanen C (1994) Apoptosis and programmed cell death in health and disease. Adv Clin Chem 31:177-246
- 9. Darville T, Giroir B, Jacobs R (1993) The systemic inflammatory response syndrome (SIRS): immunology and potential immunotherapy. Infection 21:279-290
- 10. Lynn WA, Cohen J (1995) Adjunctive therapy for septic shock: a review of experimental approaches. CID 20:143-158
- 11. Manthous CA, Hall JB, Samsel RW (1993) Endotoxin in human disease. Part 2: Biologic effects and clinical evaluations of anti-endotoxin therapies. Chest 104:1872-1881
- 12. Gawaz M, Fateh-Moghadam S, Pilz G, Gurland H-J, Werdan K (1995) Severity of multiple organ failure (MOF) but not of sepsis correlates with irreversible platelet degranulation. Infection 1:16-23
- Cavaillon J-M (1995) Controversies surrounding current therapies for sepsis syndrome. Bull Inst Pasteur 93:21-41
- 14. Talan DA (1993) Recent developments in our understanding of sepsis: evaluation of antiendotoxin antibodies and biological response modifiers. Ann Emerg Med 22:1871-1890
- 15. St. John RC, Dorinsky PM (1993) Immunologic therapy for ARDS, septic shock, and multiple-organ failure. Chest 103:932-943
- 16. Bar Natan MF, Wilson MA, Spain DA, Garrison RN (1995) Platelet-activating factor and sepsis-induced small intestinal microvascular hypoperfusion. J Surg Res 58:38-45
- 17. Lin RY, Astiz ME, Saxon JC, Rackow EC (1993) Altered leukocyte immunophenotypes in septic shock. Studies of HLA-DR, CD11b, CD14, and IL-2R expression. Chest 104:847-853
- Lin RY, Astiz ME, Saxon JC, Saha DC, Rackow EC (1993) Alterations in C3, C4, factor B, and related metabolites in septic shock [see comments]. Clin Immunol Immunopathol 69: 136-142
- 19. Busund R, Lindsetmo RO, Rasmussen LT, Rokke O, Rekvig OP, Revhaug A (1991) Tumor necrosis factor and interleukin 1 appearance in experimental gram-negative septic shock. The effects of plasma exchange with albumin and plasma infusion. Arch Surg 126:591-597
- van Deuren M, van der Ven Jongekrijg J, Demacker PN, Bartelink AK, van Dalen R, Sauerwein RW, Gallati H, Vannice JL, van der Meer JW (1994) Differential expression of proinflammatory cytokines and their inhibitors during the course of meningococcal infections. J Infect Dis 169:157-161
- 21. Korbut R, Warner TD, Gryglewski RJ, Vane JR (1994) The effect of nitric oxide synthase inhibition on the plasma fibrinolytic system in septic shock in rats. Br J Pharmacol 112: 289-291
- 22. Smith MFJ, Eidlen D, Arend WP, Gutierrez Hartmann A (1994) LPS-induced expression of the human IL-1 receptor antagonist gene is controlled by multiple interacting promoter elements. J Immunol 153:3584-3593

- 23. Glauser MP, Heumann D, Baumgartner JD, Cohen J (1994) Pathogenesis and potential strategies for prevention and treatment of septic shock: an update. Clin Infect Dis 18 (Suppl 2):S205-S216
- 24. Parrillo JE (1993) Pathogenetic mechanisms of septic shock [see comments]. N Engl J Med 328:1471-1477
- 25. Schumann RR, Rietschel ET, Loppnow H (1994) The role of CD14 and lipopolysaccharidebinding protein (LBP) in the activation of different cell types by endotoxin. Med Microbiol Immunol 183:279-297
- 26. Manthous CA, Hall JB, Samsel RW (1993) Endotoxin in human disease. Part 1: Biochemistry, assay, and possible role in diverse disease states. Chest 104:1572-1581
- 27. Levin J, Tomasulo PA, Oser RJ (1970) Detection of endotoxin in blood and demonstration of an inhibitor. J Lab Clin Med 75:903-911
- 28. Danner RL, Elin RJ, Hosseini JM, Wesley RA, Reilly JM, Parillo JE (1991) Endotoxemia in human septic shock. Chest 99:169-175
- 29. Bagasra O, Wright SD, Seshamma T, Oakes JW, Pomerantz RJ (1992) CD14 is involved in control of human immunodeficiency virus type 1 expression in latently infected cells by lipopolysaccharide. Proc Natl Acad Sci USA 89:6285-6289
- Machleidt T, Wiegmann K, Henkel T, Schutze S, Baeuerle P, Kronke M (1994) Sphingomyelinase activates proteolytic I kappa B-alpha degradation in a cell-free system. J Biol Chem 269:13760-13765
- 31. Mohri M, Spriggs DR, Kufe D (1990) Effects of lipopolysaccharide on phospholipase A2 activity and tumor necrosis factor expression in HL-60 cells. J Immunol 144:2678-2682
- 32. Raines MA, Kolesnick RN, Golde DW (1993) Sphingomyelinase and ceramide activate mitogen-activated protein kinase in myeloid HL-60 cells. J Biol Chem 268:14572-14575
- Pushkavera M, Obdeid LM, Hannun YA (1995) Ceramide: an endogenous regulator of apoptosis and growth suppression. Immonol Today 16:294-297
- 34. Kawasaki S, Moriguchi R, Sekiya K, Nakai T, Ono E, Kume K, Kawahara K (1994) The cell envelope structure of the lipopolysaccharide-lacking gram-negative bacterium Sphingomonas paucimobilis. J Bacteriol 176:284-290
- Wright SD, Kolesnick RN (1995) Does endotoxin stimulate cells by mimicking ceramide? Immonol Today 16:297-302
- 36. Hampton RY, Golenbock DT, Penman M, Krieger M, Raetz CR (1991) Recognition and plasma clearance of endotoxin by scavenger receptors. Nature 352:342-344
- 37. Ziegler-Heitbrock HWL, Ulevitch RJ (1993) CD14: cell surface receptor and differentiation marker. Immonol Today 14:121-125
- Goyer DT, Ferrero E, Retting WJ, Yenamandra AK, Obata F, LeBeau MM (1988) The CD14 monocyte differentiation antigen maps to a region encoding growth factors and receptors. Science 239:497-500
- Greenman RL, Schein RM, Martin MA, Wenzel RP, MacIntyre NR, Emmanuel G, Chmel H, Kohler RB, McCarthy M, Plouffe J et al (1991) A controlled clinical trial of E5 murine monoclonal IgM antibody to endotoxin in the treatment of gram-negative sepsis. The XOMA Sepsis Study Group [see comments]. JAMA 266:1097-1102
- 40. Teng NNH, Kaplan HS, Herbert JM (1985) Protection against Gram-negative bacteremia and endotoxemia with human monoclonal IgM antibodies. Proc Natl Acad Sci USA 82:1790-1794
- Frey EA, Miller DS, Jahr TG, Sundan A, Bazil V, Espevik T, Finlay BB, Wright SD (1992) Soluble CD14 participates in the response of cells to lipopolysaccharide. J Exp Med 176: 1665-1671
- 42. Simpson SQ, Casey LC (1989) Role of tumor necrosis factor in sepsis and acute lung injury. Crit Care Clin 5:27-47
- 43. Champan PB, Lester TJ, Casper ES, Gabrolove JL, Wong GY, Kempin SJ, Gold PJ, Welt S, Warren RS, Starnes F, Sherwin SA, Old LJ, Oettgen HF (1987) Clinical pharmacology of recombinant human tumour necrosis factor in patients with advanced cancer. J Clin Oncol 5:1942-1951

- 44. Bone RC (1991) The pathogenesis of sepsis. Ann Intern Med 115:457-469
- 45. Waage A, Brandtzaeg P, Halstensen A, Kierulf P, Espevik T (1989) The complex pattern of cytokines in serum from patients with meningococcal septic shock. Association between interleukin 6, interleukin 1, and fatal outcome. J Exp Med 169:333-338
- 46. Wong GC, Clark SC (1988) Multiple actions of interleukin 6 within a cytokine network. Immonol Today 9:137-139
- Hack CE, De Groot ER, Felt Bersma RJ, Nuijens JH, Strack Van Schijndel RJ, Eerenberg Belmer AJ, Thijs LG, Aarden LA (1989) Increased plasma levels of interleukin-6 in sepsis [see comments]. Blood 74:1704-1710
- 48. van der Poll T, Levi M, Hack CE, ten Cate H, van Deventer SJ, Eerenberg AJ, De Groot ER, Jansen J, Gallati H, Buller HR et al (1994) Elimination of interleukin 6 attenuates coagulation activation in experimental endotoxemia in chimpanzees. J Exp Med 179:1253-1259
- 49. Martich GD, Danner RL, Ceska M, Suffredini AF (1991) Detection of interleukin 8 and tumor necrosis factor in normal humans after intravenous endotoxin: the effect of antiinflammatory agents. J Exp Med 173:1021-1024
- 50. Baggiolini M, Walz A, Kunkel SL (1989) Neutrophil-activating peptide-1/interleukin 8, a novel cytokine that activates neutrophils. J Clin Invest 84:1045-1049
- 51. Leonard EJ, Yoshimura T (1990) Neutrophil attractant/activation protein-1 (NAP-1 [interleukin-8]). Am J Respir Cell Mol Biol 2:479-486
- 52. Leonard EJ, Yoshimura T (1990) Human monocyte chemoattractant protein-1 (MCP-1). Immunol Today 11:97-101
- 53. Carveth HJ, Bohnsack JF, McIntyre TM, Baggiolini M, Prescott SM, Zimmerman GA (1989) Neutrophil activating factor (NAF) induces polymorphonuclear leukocyte adherence to endothelial cells and to subendothelial matrix proteins. Biochem Biophys Res Commun 162:387-393
- Detmers PA, Lo SK, Olsen Egbert E, Walz A, Baggiolini M, Cohn ZA (1990) Neutrophilactivating protein 1/interleukin 8 stimulates the binding activity of the leukocyte adhesion receptor CD11b/CD18 on human neutrophils. J Exp Med 171:1155-1162
- Mizer LA, Weisbrode SE, Dorinsky PM (1989) Neutrophil accumulation and structural changes in nonpulmonary organs after acute lung injury induced by phorbol myristate acetate. Am Rev Respir Dis 139:1017-1026
- 56. Weiss SJ (1989) Tissue destruction by neutrophils [see comments]. N Engl J Med 320: 365-376
- 57. Argenbright LW, Letts LG, Rothlein R (1991) Monoclonal antibodies to the leukocyte membrane CD18 glycoprotein complex and to intercellular adhesion molecule-1 inhibit leukocyte-endothelial adhesion in rabbits. J Leukoc Biol 49:253-257
- Chang HR, Vesin C, Grau GE, Pointaire P, Arsenijevic D, Strath M, Pechere JC, Piguet PF (1993) Respective role of polymorphonuclear leukocytes and their integrins (CD11/118) in the local or systemic toxicity of lypopolysaccharide. J Leukoc Biol 53:636-639
- 59. Morisaki T, Goya T, Toh H, Nishihara K, Torisu M (1991) The anti Mac-1 monoclonal antibody inhibits neutrophil sequestration in lung and liver in a septic murine model. Clin Immunol Immunopathol 61:365-375
- Burch RM, Noronha Blob L, Bator JM, Lowe VC, Sullivan JP (1993) Mice treated with a leumedin or antibody to Mac-1 to inhibit leukocyte sequestration survive endotoxin challenge. J Immunol 150:3397-3403
- 61. Klein B, Brailly H (1995) Cytokine-binding proteins: stimulating antagonist. Immonol Today 16:216-220
- 62. Klein B, Wijdenes J, Zhang XG, Jourdan M, Boiron JM, Brochier J, Liautard J, Merlin M, Clement C, Morel Fournier B et al (1991) Murine anti-interleukin-6 monoclonal antibody therapy for a patient with plasma cell leukemia. Blood 78:1198-1204
- 63. Franslow WC, Sims JE, Sassenfeld H, Morrissey PJ, Gillis S, Dower SK, Widmer MB (1990) Regulation of alloreactivity in vivo by a soluble form of the interleukin-1 receptor. Science 248:739-742

- 64. Fischer E, Van Zee KJ, Marano MA, Rock CS, Kenney JS, Poutsiaka DD, Dinarello CA, Lowry SF, Moldawer LL (1992) Interleukin-1 receptor antagonist circulates in experimental inflammation and in human disease. Blood 79:2196-2200
- 65. Granowitz EV, Porat R, Mier JW, Orencole SF, Callahan MV, Cannon JG, Lynch EA, Ye K, Poutsiaka DD, Vannier E et al (1993) Hematologic and immunomodulatory effects of an interleukin-1 receptor antagonist coinfusion during low-dose endotoxemia in healthy humans. Blood 82:2985-2990
- 66. Rogy MA, Moldawer LL, Oldenburg HS, Thompson WA, Montegut WJ, Stackpole SA, Kumar A, Palladino MA, Marra MN, Lowry SF (1994) Anti-endotoxin therapy in primate bacteremia with HA-1A and BPI. Ann Surg 220:77-85
- Dinarello CA (1993) Modalities for reducing interleukin-1 activity in disease. Immonol Today 14:260-264
- 68. Carlet J, Cohen J, Andersson J (1994) INTERSEPT: an international efficacy and safety study of monoclonal antibody to human TNF in patients with sepsis syndrome. In: Program and Abstracts of the 34th Interscience Conference on Antimicrobial Agents and Chemotherapy, American Society for Microbiology, Washington DC
- Ertel W, Scholl FA, Gallati H, Bonaccio M, Schildberg FW, Trentz O (1994) Increased release of soluble tumor necrosis factor receptors into blood during clinical sepsis. Arch Surg 129:1330-1336
- 70. Rose John S, Heinrich PC (1994) Soluble receptors for cytokines and growth factors: generation and biological function. Biochem J 300:281-290
- Ashkenazi A, Marsters SA, Capon DJ, Chamow SM, Figari IS, Pennica D, Goeddel DV, Palladino MA, Smith DH (1991) Protection against endotoxic shock by a tumor necrosis factor receptor immunoadhesin. Proc Natl Acad Sci USA 88:10535-10539
- 72. Kurschner C, Ozmen L, Garotta G, Dembic Z (1992) IFN-gamma receptor-Ig fusion proteins. Half-life, immunogenicity, and in vivo activity. J Immunol 149:4096-4100
- 73. Lesslauer W, Tabuchi H, Gentz R, Brockhaus M, Schlaeger EJ, Grau G, Piguet PF, Pointaire P, Vassalli P, Loetscher H (1991) Recombinant soluble tumor necrosis factor receptor proteins protect mice from lipopolysaccharide-induced lethality. Eur J Immunol 21:2883-2886
- 74. Peppel K, Crawford D, Beutler B (1991) A tumor necrosis factor (TNF) receptor-IgG heavy chain chimeric protein as a bivalent antagonist of TNF activity. J Exp Med 174:1483-1489
- 75. Schade UF (1990) Pentoxifylline increases survival in murine endotoxin shock and decreases formation of tumor necrosis factor. Circ Shock 31:171-181
- 76. Gadina M, Bertini R, Mengozzi M, Zandalasini M, Mantovani A, Ghezzi P (1991) Protective effect of chlorpromazine on endotoxin toxicity and TNF production in glucocorticoidsensitive and glucocorticoid-resistant models of endotoxic shock. J Exp Med 173:1305-1310
- 77. Gonzalo JA, Gonzalez Garcia A, Kalland T, Hedlund G, Martinez C, Kroemer G (1993) Linomide, a novel immunomodulator that prevents death in four models of septic shock. Eur J Immunol 23:2372-2374
- 78. Grundmann HJ, Hahnle U, Hegenscheid B, Sahlmuller G, Bienzle U, Blitstein Willinger E (1992) Inhibition of endotoxin-induced macrophage tumor necrosis factor expression by a prostacyclin analogue and its beneficial effect in experimental lipopolysaccharide intoxication. J Infect Dis 165:501-505
- 79. Novogrodsky A, Vanichkin A, Patya M, Gazit A, Osherov N, Levitzki A (1994) Prevention of lipopolysaccharide-induced lethal toxicity by tyrosine kinase inhibitors. Science 264: 1319-1322
- Tzung SP, Mahl TC, Lance P, Andersen V, Cohen SA (1992) Interferon-alpha prevents endotoxin-induced mortality in mice. Eur J Immunol 22:3097-3101
- Rhynes VK, McDonald JC, Gelder FB, Aultman DF, Hayes JM, McMillan RW, Mancini MC (1993) Soluble HLA class I in the serum of transplant recipients. Ann Surg 217:485-489
- 82. Zavazava N, Bottcher H, Ruchholtz WM (1993) Soluble MHC class I antigens (sHLA) and anti-HLA antibodies in heart and kidney allograft recipients. Tissue Antigens 42:20-26
- 83. Zhang H, Benlabed M, Spapen H, Nguyen DN, Vincent JL (1994) Prostaglandin E1 increases oxygen extraction capabilities in experimental sepsis. J Surg Res 57:470-479

- 84. Kuipers B, van der Poll T, Levi M, van Deventer SJ, ten Cate H, Imai Y, Hack CE, ten Cate JW (1994) Platelet-activating factor antagonist TCV-309 attenuates the induction of the cytokine network in experimental endotoxemia in chimpanzees. J Immunol 152:2438-2446
- 85. Moncada S, Higgs A (1993) The L-arginine-nitric oxide pathway. N Engl J Med 329: 2002-2012
- 86. Szabo C, Thiemermann C (1995) Invited opinion: role of nitric oxide in hemorrhagic, traumatic, and anaphylactic shock and thermal injury. Shock 2:145-155
- 87. Szabo C, Southan GJ, Thiemermann C (1994) Beneficial effects and improved survival in rodent models of septic shock with S-methylisothiourea sulfate, a potent and selective inhibitor of inducible nitric oxide synthase. Proc Natl Acad Sci USA 91:12472-12476
- Vallance P, Moncada S (1994) Nitric oxide-from mediator to medicines. J R Coll Physicians Lond 28:209-219
- 89. Vallance P, Moncada S (1994) Drugs that alter nitric oxide homeostasis: a reply [letter; comment]. Cardiovasc Res 28:284
- 90. Vallance P (1994) Nitric oxide in clinical arena. Biochemist 16:23-28
- Wang P, Ba ZF, Chaudry IH (1994) Nitric oxide. To block or enhance its production during sepsis? Arch Surg 129:1137-1142
- 92. Vallance P, Collier J, Moncada S (1989) Effects of endothelium-derived nitric oxide on peripheral arteriolar tone in man [see comments]. Lancet 2:997-1000
- Vallance P (1989) The interplay between platelet and vessel-wall mediators in coronary artery occlusion. Biomed Pharmacother 43:113-119
- Benjamin N, Vallance P (1994) Plasma nitrite as a marker of nitric oxide production [letter]. Lancet 344:960
- 95. Lin PJ, Chang C-H, Chang J-P (1994) Reversal of refractory hypotension in septic shock by inhibitor of nitric oxide synthase. Chest 106:626-629
- 96. Petros A, Lamb G, Leone A, Moncada S, Bennett D, Vallance P (1994) Effects of a nitric oxide synthase inhibitor in humans with septic shock. Cardiovasc Res 28:34-39
- 97. Southan GJ, Szabo C, Thiemermann C (1995) Isothioureas: potent inhibitors of nitric oxide synthases with variable isoform selectivity. Br J Pharmacol 114:510-516
- 98. Earnshaw WC (1995) Apoptosis: lessons from in vitro systems. Trends Cell Biol 5:217-220
- 99. Hawkins CJ, Vaux DL (1994) Analysis of the role of bcl-2 in apoptosis. Immunol Rev 142: 127-139