



Characterization of phosphorus in algae from a eutrophic lake by solution ^{31}P nuclear magnetic resonance spectroscopy

Weiying Feng¹ · Cuicui Li^{1,2,3} · Chen Zhang¹ · Shasha Liu¹ · Fanhao Song¹ · Wenjing Guo¹ · Zhongqi He⁴ · Tingting Li¹ · Haiyan Chen¹

Received: 4 July 2018 / Accepted: 12 September 2018 / Published online: 4 October 2018
© The Author(s) 2018

Abstract

The identification and quantification of phosphorus (P) compounds derived from algal biomass are crucial for a better understanding of algal P dynamics in lake ecosystems. Quantity and species of P in algae collected from Chao Lake (a typical ultra-eutrophic lake) in China were analyzed by chemical analysis and ^{31}P NMR. Total P (TP) in algae biomass ranged from 2671 to 5385 mg kg⁻¹ of dry matter. Proportion of organic P (P_o) accounted for 78.3 ± 2.6% in algae biomass collected from the western part of Chao Lake, which was higher than that (64.7 ± 1.4%) in the eastern part of the lake. Eight P species including inorganic P species (orthophosphate and pyrophosphate) and P_o species (five monoesters P and diesters P) were identified in NaOH–EDTA extracts of algal samples. Monoesters P accounted for 48.4% in extracted TP, which was the main component of P_o. β -glycerophosphates were the largest component of monoesters P, which accounted for 22.6% in extracted TP. This study improved knowledge on the mechanism of the cycling of endogenous P in the aquatic system and would be helpful in developing a strategy for control of repeated algae blooms in eutrophic Chao Lake.

Keywords ^{31}P NMR · Algae · Organic phosphorus · Lake · Eutrophication

Introduction

Algae blooms are serious environmental problems around the world, especially in developing countries (Kagaloua et al. 2008; Pernet-Coudrier et al. 2012). Algae blooms occur in an aquatic environment if too much phosphorus

(P) enters the system (Giles et al. 2015). For a eutrophic lake, release of P from dead algae is an important nutrient source that will support continuous algal blooms in lakes (Li et al. 2009; Feng et al. 2016a). Algae are not usually collected by humans because they are not valuable, so that the debris is allowed to decompose in situ. The decomposition of algal residues affects the bio-cycling and release of P, increasing the risk of resurgence of algal blooms (Feng et al. 2016b; Lehman et al. 2017). Decaying algal debris releases both inorganic P (P_i) and P_o. The P_o constituents need to be hydrolyzed to bioavailable P_o by various enzymes (Feng et al. 2016b). Therefore, the forms and concentrations of P in algae shall be evaluated while algae debris is decomposing. However, until now, studies of the species, concentrations, and effects of P_o in algae of eutrophic lakes have been limited because of the complexity and limitations of analytical methods (Turner et al. 2005; Bell et al. 2017).

Typical analytical approaches, such as enzymatic reactivity, high performance liquid chromatography (HPLC), and mass spectrometry are based on operational definitions so that they cannot discern P classes at a molecular level (Suzumura 2005; Baldwin 2013; Karl 2014). Phosphorus-31 nuclear magnetic resonance spectroscopy (^{31}P

Handling Editor: Richard Sheibley.

Weiying Feng and Cuicui Li are co-lead authors.

✉ Chen Zhang
zhangchen0353@yeah.net

✉ Haiyan Chen
chenhy@craes.org.cn

¹ State Key Laboratory of Environmental Criteria and Risk Assessment, Chinese Research Academy of Environmental Sciences, Beijing 100012, China

² Guangzhou Institute of Geochemistry, Chinese Academy of Sciences, Guangzhou 510640, China

³ University of Chinese Academy of Sciences, Beijing 100049, China

⁴ USDA-ARS, Southern Regional Research Center, New Orleans, LA 70124, USA

NMR) is a non-destructive, non-invasive technique for identifying chemical forms in various environmental samples. Solid-state ^{31}P NMR is focused on inorganic P compounds, solution ^{31}P NMR is mainly used to determine organic P compounds (Turner et al. 2005; He et al. 2011; Abdi et al. 2014; Sørensen et al. 2014). Several P compounds have been detected by solution ^{31}P NMR, including phosphonates, orthophosphate, monoesters P, diesters P, pyrophosphates and polyphosphates (He et al. 2007; Turner et al. 2012; Zhu et al. 2013). Generally, monoesters P represent a wide range of important P_o compounds, such as inositol phosphate and sugar phosphates (He et al. 2007; Doolette et al. 2009; Jarosch et al. 2015). Therefore, it is an ideal technique for analyzing P_o species in algae of eutrophic lakes, as it would not only provide important information pertaining to P biogeochemical cycling in lake ecosystems, but also yield abundant insight into identities of specific P compounds.

Chao Lake ($31^\circ 25' 28'' - 31^\circ 43' 28'' \text{N}$, $117^\circ 16' 54'' - 117^\circ 51' 46'' \text{E}$) is one of the five largest freshwater lakes in China. It is situated on the flood plains between the Yangtze River and Huai River in the central Anhui Province of eastern China (Zan et al. 2010; Tang et al. 2015). Chao Lake is a typical shallow lake with a mean depth of 3 m and a surface area of 780 km^2 and drainage area of

$13,350 \text{ km}^2$ (Wang et al. 2013). Due to the rapid increase in anthropogenic activities in the lake's watershed over recent decades, the lake has suffered from serious pollution, eutrophication and algae blooms (Xu et al. 2005). As a matter of fact, since the mid 1980s, algae blooms have occurred each year in Chao Lake (Chen and Liu 2014). In order to assess the environmental risks of algae on the eutrophic lake, this study analyzed the P species in algae by solution ^{31}P NMR, and based on the knowledge obtained, predicted the P bioavailability of the algal biomass in aquatic environments.

Materials and methods

Study sites and sample collection

Samples of algae were collected from six sites in Chao Lake in September 2015 (Fig. 1). These sampling sites were located in different eutrophic areas. Generally, Chao Lake is divided into two parts: the western part (samples C_1 and C_2) and the eastern part (samples C_3 , C_4 , C_5 and C_6) along the line of Zhongmaio–Mushan–Qitouzui, as shown in Fig. 1. The quality of water was worst in the western part of the lake and gradually became better from west to east (Zhu et al.

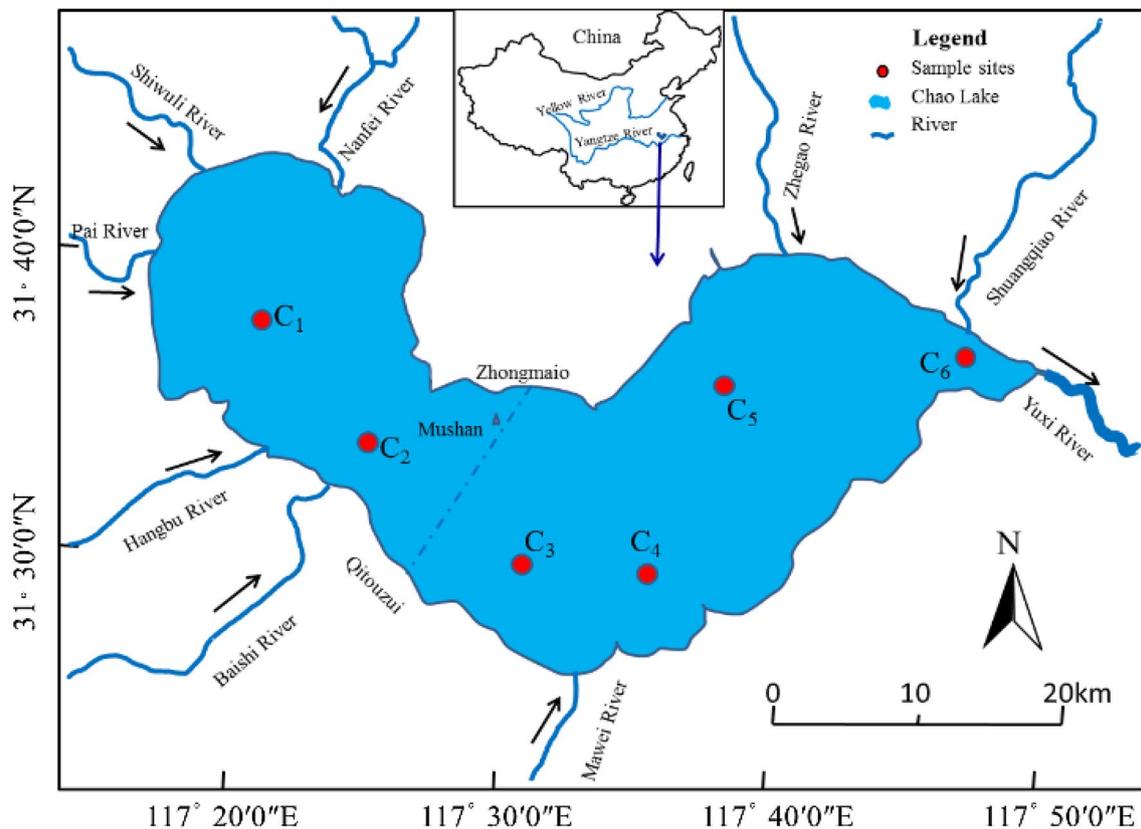


Fig. 1 Major inlets and outlets of Chao Lake with algal sampling sites

2006; Tang et al. 2015). The lake's annual mean concentrations of total nitrogen (TN) and TP approached 2.85 and 0.26 mg l⁻¹, respectively, and the annual mean chlorophyll-*a* reached up to 25.6 µg l⁻¹ (Li et al. 2015). Algae were collected by use of a plankton collector (HB403-BWS). Samples were placed in sealed bags and put in ice boxes immediately. These algal samples were freeze-dried, ground, then passed through a 2-mm sieve before being stored at -20 °C (Feng et al. 2016a). The dominant species in Chao Lake was *Microcystic aeruginosa* with an appearance frequency of 90.9% (Yang et al. 2011).

Extraction of P and chemical analysis

Samples of algae were extracted by use of optimized NaOH-EDTA extracting agent (mixtures of 0.5 mol l⁻¹NaOH and 25 mmol l⁻¹EDTA) with a ratio of 150:1 (ml g⁻¹), and the mixtures were shaken for 18 h at room temperature (Cade-Menun and Preston 1996; Feng et al. 2016b). The extracting solutions were centrifuged (8000 × g) for 30 min, and filtered through 0.45-µm glass-fiber filters (Whatman GF/C). Extractable total phosphorus (ETP) after digestion and free molybdate reactive phosphorus (MRP) were measured using the molybdenum blue method (He and Honeycutt 2005). Extractable organic phosphorus (EOP) was calculated by the difference between ETP and MRP. The remaining extracts were freeze-dried for solution ³¹P NRM spectroscopy analysis.

Percentages of carbon (C) and nitrogen (N) in algae were determined by use of an elemental analyzer (Elementarvario macro EL, Germany). Total phosphorus (TP) and inorganic phosphorus (P_i) were determined by the SMT method described by Ruban et al. (2001). Organic phosphorus (P_o) in algae samples was calculated by the difference between TP and P_i. There were three replications for extraction of P and chemical analysis. Data were checked for deviations from normality and homogeneity of variance before performing statistical analyses.

³¹P NMR analysis

A 100-mg sample of freeze-dried algal extracts was ground, and then redissolved in 1 ml 1 mol l⁻¹ NaOH + 0.1 mol l⁻¹ EDTA and 0.2 ml D₂O. After ultrasonication for 30 min and equilibration for 5 min, 2% (v/v) of bicarbonate buffered dithionite (0.11 mol l⁻¹ NaHCO₃ + 0.11 mol l⁻¹ Na₂S₂O₄) was added to the extract to reduce interference from paramagnetic ions, such as Fe and Mn (He et al. 2009; Giles and Cade-Menun 2014). The pH of the supernatant solution was adjusted using 10 mol l⁻¹ NaOH to ensure a pH > 12. The supernatant solution was centrifuged (8000 × g) for 30 min and transferred to a 5-mm NMR tube. Solution ³¹P NMR spectra were acquired at 24 °C on a Bruker AVANCE

400 MHz spectrometer at a ³¹P frequency of 161.98 MHz, using a 90° pulse, a 5 s relaxation delay and a 0.21 s acquisition time, similar to the parameters used in Feng et al. (2016b). The scan time for each sample was more than 15 h. Peak areas were calculated by integration and completed using MestrelabMnova v.10.

Spiking experiments

The peak of specific monoesters P forms (i.e., glucose 6-phosphate, ribonucleotides, α-glycerophosphate, β-glycerophosphate, myo-inositol hexaphosphate) needs to be confirmed with spiking experiments (Fig. 4). Methods for identifying specific P forms in NMR spectra of soil and other environmental samples are well-established and have been used for many years (Smernik and Dougherty 2007; He et al. 2011; McDowell and Hill 2015), combined with P compound libraries developed by Turner et al. (2003); Doolette et al. (2009) and Cade-Menun (2015). Standard samples of monoesters P were purchased from Sigma-Aldrich. Spiked samples were analyzed by ³¹P NMR as described above. Monoesters P compounds were identified by their chemical shifts, with the orthophosphate peak in all spectra standardized to 6.0 ppm. Spectral processing was done using MestReNove software version 9.0.1 (MestReNove Research SL).

Results and analysis

Nutrients (C, N and P) in debris of algae

Composition of C, N and their ratios in debris of algae are shown in Table 1. Percentages of C ranged from 31.5 to 50.3% with a mean value of 41.9% in the algae samples from Chao Lake. Content of N was 5.2–7.8% with a mean value of 6.4%. Both contents of C and N of algae was greater than those of aquatic macrophytes, which has been widely reported previously (Zhong et al. 2012; Qu et al. 2013, Feng et al. 2016a). The ratio of C:N was a good predictor of degradation in algae and aquatic macrophytes with a lower ratio of C:N for material more readily degradation (Reitzel et al. 2006). In this study, the ratios of C:N in algae from the western lake (7.1 ± 0.3) were higher than those of the eastern lake (6.4 ± 0.3), suggesting higher lability of the algal debris of the eastern lake. The ratio of C:N of aquatic macrophytes (average 12.9) (Feng et al. 2016a) was greater than that of algae in Chao Lake (average 6.7) (Table 1). Therefore, the algae decomposed more easily than aquatic macrophytes in the same lake. This was also consistent with the results of previous studies (Liu et al. 2016).

Contents of TP in debris of algae ranged from 2671 to 5385 mg kg⁻¹ dry mass (dm) with a mean value of

Table 1 Contents of C, N, and P in algae and their NaOH–EDTA extraction efficiency in Chao Lake

Samples	Coordinates	Original algae powders						NaOH–EDTA extract algae samples	
		C (%)	N (%)	C:N	TP(mg kg ⁻¹)	P _o (mg kg ⁻¹)	P _o /TP(%)	ETP(mg kg ⁻¹)	EOP(mg kg ⁻¹)
C ₁	31°38'3.52"E, 117°21'16.74"N	44.8 ± 2.5 ^a	6.7 ± 0.5	6.7 ± 0.3	4173 ± 516	3372 ± 126	80.8 ± 1.6	1864 ± 126(44.7) ^b	785 ± 124(23.3) ^b
C ₂	31°34'3.50"E, 117°24'51.80"N	39.5 ± 0.6	5.3 ± 0.8	7.5 ± 0.2	4059 ± 125	3072 ± 432	75.7 ± 3.5	1757 ± 214(43.3)	464 ± 52(15.1)
C ₃	31°30'9.64"E, 117°28'56.52"N	38.3 ± 1.8	5.4 ± 0.2	7.1 ± 0.4	3787 ± 256	2536 ± 59	67.0 ± 2.5	1572 ± 56(41.5)	351 ± 98(13.8)
C ₄	31°27'56.84"E, 117°34'50.00"N	31.5 ± 3.5	5.2 ± 0.1	6.1 ± 0.1	2671 ± 198	1476 ± 123	55.3 ± 1.4	1292 ± 89(48.4)	567 ± 15(38.4)
C ₅	31°33'27.70"E, 117°36'46.18"N	50.3 ± 2.9	7.8 ± 0.8	6.5 ± 0.4	3543 ± 112	2107 ± 78	59.5 ± 0.9	2774 ± 21(78.3)	1141 ± 215(54.2)
C ₆	31°36'10.25"E, 117°47'38.55"N	46.9 ± 4.6	7.8 ± 0.7	6.0 ± 0.2	5385 ± 290	4136 ± 19	76.8 ± 0.8	2183 ± 51(40.5)	1066 ± 164(25.8)

ETP extractable total phosphorus, EOP extractable organic phosphorus

^aMean ± standard deviation ($n = 3$)

^bValues in brackets show the percentage of ETP, EOP in NaOH–EDTA extracts to TP and P_o in the unextracted algae powders, respectively

3936 mg kg⁻¹. The greatest concentration of TP in debris of algae was observed in sample C₆. This may be because of two major inflowing rivers (e.g., the Zhe gao river and Shuang qiao river) with heavy nutrition inputs (Tang et al. 2015). Contents of TP in the surface sediments from Chao Lake only ranged from 420 to 1090 mg kg⁻¹ with a mean value of 687 mg kg⁻¹ (Zhang and Xing 2013). Content of TP in debris of algae from Chao Lake was approximately 5 times higher than TP in surface sediments. Therefore, dead algae-derived P might be an important source of bioavailable P for repeated algal blooming in eutrophic lakes such as Chao Lake. Concentration of P_o in these algal samples ranged from 2107 to 4136 mg kg⁻¹ with a mean value of 2783 mg kg⁻¹. The greatest concentration of P_o was also observed in the heavily polluted region (i.e., sample C₆). The proportion of P_o in algae of the western lake (78.3% ± 2.6%) was higher than that in the eastern lake (64.7% ± 1.4%). The mean ratio of P_o/TP of the six algal samples was 69.2% in Chao Lake (Table 1). This value was greater than that of algae from Tai Lake (mean ratio of P_o/TP 57.8%) (Feng et al. 2016b). The previous studies reported that P_o could be converted to bioavailable P (e.g., HPO₄²⁻) for algae blooming through a series of redox-driven solubilization reactions and phosphatase-mediated hydrolytic processes (Wang and Pant 2010; Zhu et al. 2015). Thus, we believed that P_o in debris of algae from Chao Lake possessed larger bioavailability potential than algae from Tai Lake per their difference in P_o/TP ratios.

NaOH–EDTA extractable P from algae

In this study, contents of NaOH–EDTA extractable TP ranged from 1292 to 2774 mg kg⁻¹, with an extraction

efficiency of 40.5–78.3%, and contents of extractable P_o from algae ranged from 351 to 1141 mg kg⁻¹, with an extraction efficiency of 13.8–54.2% (Table 1). The recoveries of P_o in debris of algae from Chao Lake were lower than those with pure algae such as *Microcystis*, *Chlorella vulgaris*, and *Spirulina platensis* (Feng et al. 2016a). However, the extraction efficiencies of P_o of algae from Chao Lake were similar to those of particulate P from Tai Lake (23–56%) (Bai et al. 2017) and of sediments from Haihe River (30–73%) (Zhang et al. 2017). The extraction efficiency of TP from algae from Chao Lake was similar to that of soils and sediments (49–83%) (Xu et al. 2005; Tang et al. 2015). Multiple-step extractions (e.g., additional or sequential HCl extraction) (He et al. 2008; Cade-Menun 2015; Zhu et al. 2016; Liu et al. 2017) seem needed to increase the P recovery from these algal samples.

Solution ³¹P NMR spectra of NaOH–EDTA extracts of algae

Eight main P species including inorganic P species (orthophosphate and pyrophosphate) and P_o species (five monoesters P and diesters P) were identified in the NaOH–EDTA extracts of the six algal samples by solution ³¹P NMR (Fig. 2; Table 2). The peak of orthophosphate was at 6.00 ppm in the ³¹P NMR spectra, monoesters P was at 3.33–5.49 ppm, diesters P was at -0.69 to -0.31 ppm, and pyrophosphate was at -4.21 to -4.12 ppm (Fig. 2). The sum of orthophosphate and monoesters P in ETP accounted for more than 93% of ETP (Table 1; Fig. 3b). With the peak of orthophosphate as the largest signal in these ³¹P NMR spectra. The content of orthophosphate was between 615.1 and 1331.7 mg kg⁻¹, and accounted

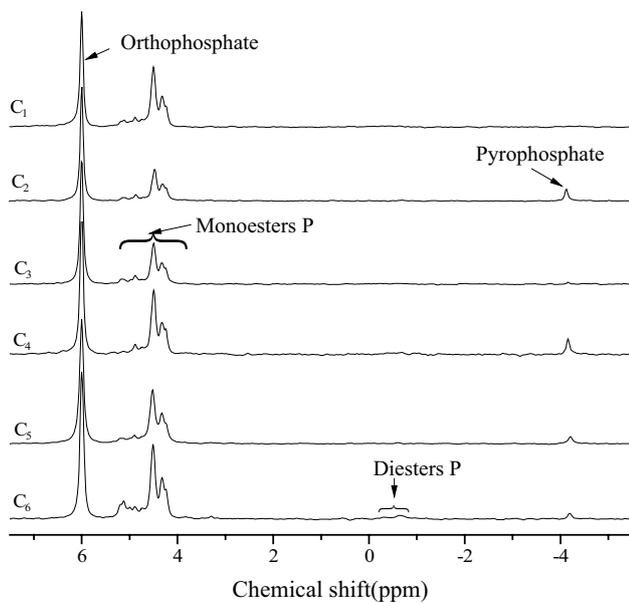


Fig. 2 Solution ³¹P NMR spectra of NaOH-EDTA extracts of algae in Chao Lake

for 41.5–54.0% of ETP (Table 1; Fig. 3). Polyphosphates were not detected in any algae of Chao Lake.

Monoesters P comprised the largest P_o fraction with NaOH-EDTA extracts, and accounted for 48.4% (average) in ETP (Table 2; Fig. 3b). Through the spiking experiments (Fig. 4), the peak at 4.88 ± 0.02 ppm was assigned to α-glycerophosphates and the peak at 4.50 ± 0.02 ppm was assigned to β-glycerophosphates, based on Turner and Richardson (2004) and Doolette et al. (2009); the percentages of α- and β-glycerophosphates in ETP were 3.4% (65.5 mg kg⁻¹) and 22.6% (432.7 mg kg⁻¹), respectively. β-glycerophosphates were the largest component of monoesters P. The peak at 4.32 ± 0.01 ppm was assigned to ribonucleotides (He et al. 2011) and the percentage of ribonucleotides in ETP was 10.6% (218.2 mg kg⁻¹). The peak at 5.12 ± 0.01 ppm was assigned to glucose 6-phosphate (Cade-Menun 2015), the percentage of which in ETP was 1.9% (38.7 mg kg⁻¹). In addition, a few of the peaks in the monoesters P region were unidentified, because chemical shifts were strongly influenced by subtle differences among samples for viscosity, pH, salts and paramagnetic ions (Young et al. 2013; Abdi et al. 2014; He et al. 2011; Giles et al. 2015). Unidentified monoesters P are defined as ‘other monoesters P’ in this study. These other monoesters P accounted for 9.9% of ETP in algae of Chao Lake (Table 2).

Pyrophosphate was detected in most samples except the sample C₁, which is consistent with a number of other studies (Bedrock et al. 1995; Mahieu et al. 2000; He et al. 2011). In other literature, polyphosphates and pyrophosphate were

Table 2 Concentrations of P compounds in NaOH-EDTA extracts of the algae determined by solution ³¹P NMR

Algae	P _i (mg kg ⁻¹)		P _o (mg kg ⁻¹)						Total P _o
	Orthophosphate	Pyrophosphate	Monoesters P			Other monoesters P			
			Glucose 6-phosphate	Ribonucleotides	α-glycerophosphate	β-glycerophosphate			
C ₁	818.2(43.9) ^a	n.d.	32.7(1.8)	204.6(11.0)	60.6(3.2)	526.9(28.3)	220.9(11.9)	n.d.	1045.7(56.1)
C ₂	938.1(53.4)	115.4(6.6)	42.2(2.4)	144.5(8.2)	63.8(3.6)	322.7(18.4)	130.4(7.4)	n.d.	703.5(40.0)
C ₃	849.1(54.0)	15.8(1.0)	n.d.	137.6(8.7)	49.2(3.1)	325.2(20.5)	201.5(12.7)	n.d.	714.1(45.0)
C ₄	615.1(47.6)	65.8(5.1)	15.4(1.2)	131.2(8.8)	44.9(3.5)	297.7(23.0)	131.0(10.1)	9.2(0.7)	611.4(47.3)
C ₅	1331.7(48.0)	87.9(3.2)	49.3(1.8)	487.4(17.6)	103.9(3.7)	636.6(22.9)	67.9(2.4)	9.3(0.3)	1354.4(48.8)
C ₆	905.6(41.5)	32.6(1.5)	92.4(4.2)	203.8(9.3)	70.6(3.2)	487.2(22.3)	329.6(15.1)	61.6(2.8)	1245.2(57.0)

P_i inorganic P, P_o organic P, Total P_o the sum of monoesters P and diesters P, n.d not detected

^aValues in parentheses are percentages of individual P compounds in NaOH-EDTA extracts TP

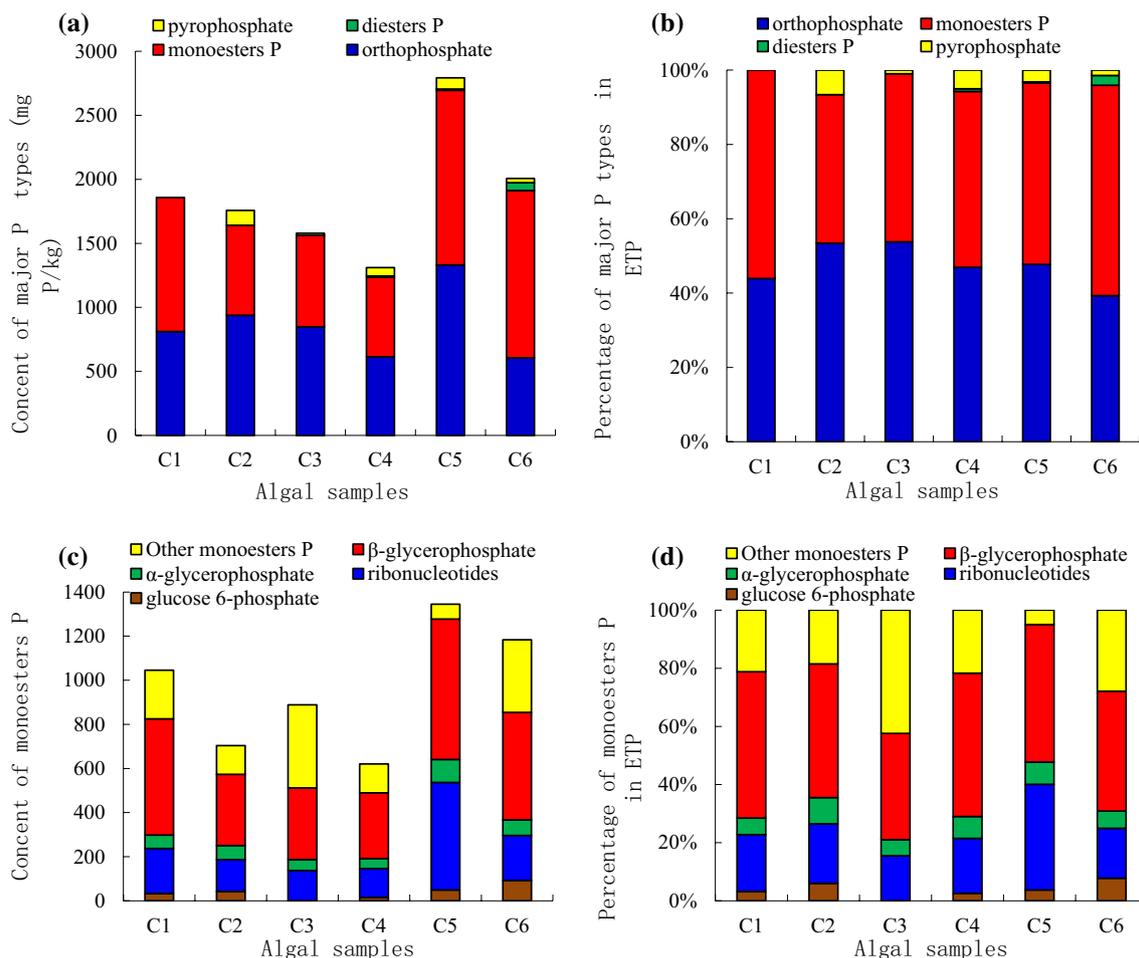


Fig. 3 Content and percentage of major P types (a, b) and monoesters P compound forms (c, d) in algae collected from six sites (C₁–C₆) of Chao Lake

detected in some samples, but not necessarily (Busato et al. 2005; Feng et al. 2016b).

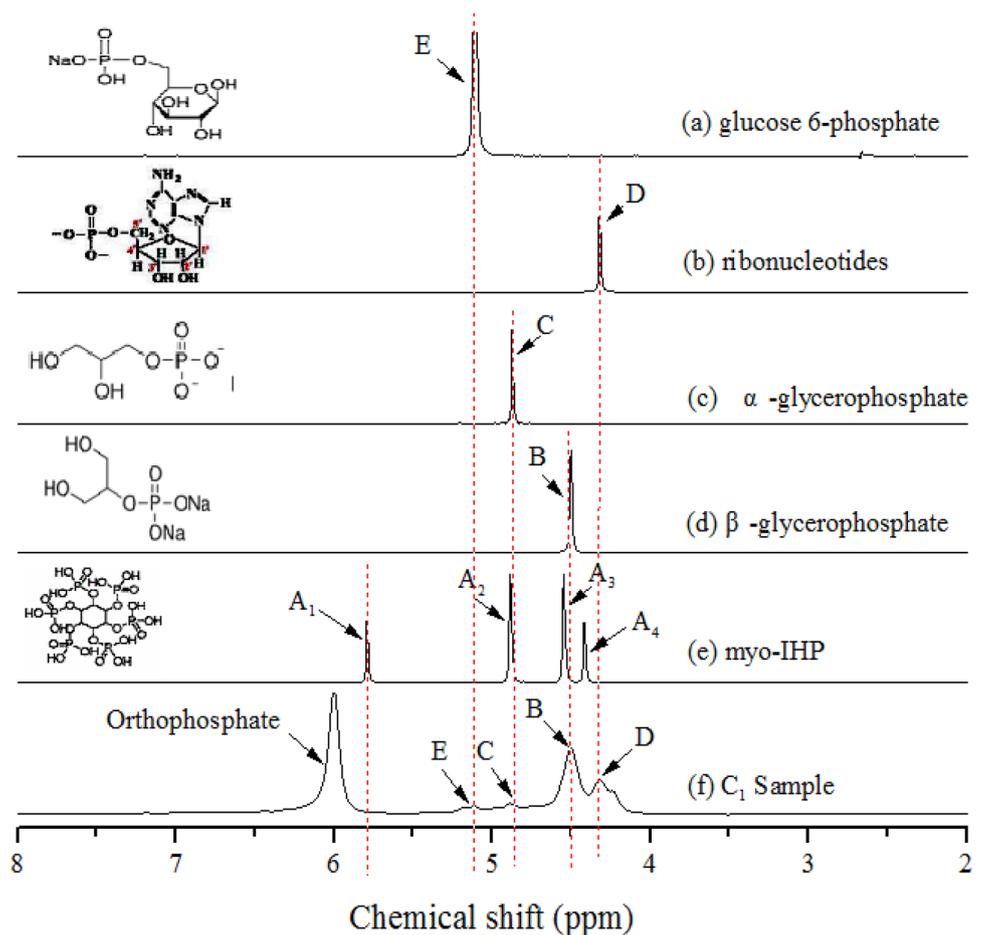
Discussion

Identification of inositol hexaphosphate (IHP) in algae of Chao Lake

The inositol hexaphosphate (IHP) stereoisomers (scyllo-, myo-, chiro-, neo-IHP) were important monoesters P components in many environmental samples (Turner et al. 2012; Cade-Menun 2015). Each of these compounds contains six phosphates, and the conformation of those phosphate groups causes them to have multiple peaks in a single spectrum, in an arrangement specific to each compound. The only exception is scyllo-IHP, which has one peak for the six phosphates; the peak at 3.55 ± 0.02 ppm was assigned to scyllo-IHP, based on Turner and Richardson (2004) and Doolette et al. (2009). None of the spectra

shown in Fig. 2 have a peak at 3.55 ppm, so the scyllo-IHP was not present in algae of Chao Lake. Myo-IHP has four peaks in a 1:2:2:1 arrangement (with respect to peak areas), with peaks at 5.79 ± 0.01 , 4.88 ± 0.01 , 4.55 ± 0.01 and 4.42 ± 0.01 ppm (in Fig. 4e). Three of these peaks were not present in algae samples in Chao Lake (Fig. 2), so myo-IHP was not present in algae of Chao Lake. The fact that myo-IHP had not been observed in algae in a previous study is consistent with the observation of this study (Feng et al. 2016a). For chiro-IHP [in either the 4 equatorial/2 axial (4e/2a) conformation or the 2e/4a conformation], three peaks must be clearly visible, in a 2:2:2 arrangement, and the diagnostic peaks for each are at 6.2–6.5 ppm (Cade-Menun 2015); however, none of the spectra shown in Fig. 2 have any peaks between 6.0 and 7.0 ppm, so these compounds were not present in algae of Chao Lake. In addition, neo-IHP requires two peaks to be present, in a 4:2 arrangement, so that the peak at 6.4 ± 0.01 ppm is twice as large as the one at 4.3 ± 0.01 ppm. Given that there were no peaks between 6.0 and 7.0 ppm, it can be

Fig. 4 Solution ^{31}P NMR spectra of monoesters P standard compounds (a–e) and C_1 algal sample in Chao Lake (f)



assumed that neo-IHP was not present in algae of Chao Lake.

Degradation behaviors of diesters P in algae of Chao Lake

Apart from monoesters P, other important P_0 compounds, diesters P, were detected in some algae samples. The concentration of diesters P was generally low (mean 0.63% of ETP), compared to other P_0 fractions, and only detected in samples C_4 , C_5 and C_6 ($9.2\text{--}61.6 \text{ mg kg}^{-1}$).

It is well established that some diesters P such as phospholipids and RNA can degrade to monoesters P [e.g., α - β -glycerophosphates (phospholipids) and various monophosphates (e.g., nucleotides) when analyzed at the high pH required for good peak separation in ^{31}P NMR spectra] (Turner et al. 2003; Doolette et al. 2009; He et al. 2011; Schneider et al. 2016). The degree of degradation will vary depending on the length of NMR experiment and other factors (Cade-Menun and Liu 2014; Cade-Menun 2015; Feng et al. 2018). It was essential that these degradation peaks were identified and quantified, in order to determine the correct concentrations of monoesters P and diesters

P (Young et al. 2013; Vincent et al. 2013). Therefore, the corrected total monoesters P and corrected total diesters P were those corrected by moving the percentages of α - and β -glycerophosphates and nucleotides from monoesters P to diesters P. When uncorrected, the total monoesters P were significantly higher than the total diesters P, but the reverse was true for the corrected values.

Eutrophication and algal blooming versus biogeochemical cycling of algae-derived P in Chao Lake

Based on the results in this study, we were able to think further about the biogeochemical cycling of P driven by algal blooming in Chao Lake. The TP of algae-derived biomass loading was $10.94 \times 10^3 \text{ kg}$ in Chao Lake (Li et al. 2015). With the P_0 content in algae determined in this study (Table 1), we estimated the P_0 biomass of algae to be approximately $7.57 \times 10^3 \text{ kg}$ in Chao Lake. In previous research (Feng et al. 2016b) with Tai Lake samples, we estimated that approximately 32.7–41.3% of extractable P_0 from algae has the potential for phosphatase hydrolysis to soluble orthophosphate which can be released into the water body.

Thus, in the case of Chao Lake, this bioavailable P would be 2475–3126 kg in algae and would be released into the water and promote repeated algal blooms in Chao Lake if not appropriately removed naturally or artificially. This conclusion indicated that decomposition of algal debris would be a key factor in regeneration of bioavailable P for life in eutrophic lakes, even when external P is excluded. It is therefore necessary to remove algae debris from eutrophic lakes to control the release of P from internal P cycling and the phenomenon of eutrophication of lakes.

Conclusion

This research used solution ^{31}P NMR to provide insights into the P species and distribution of P in algae of the heavily polluted Chao Lake. Data derived from this study showed that the eutrophic lake algae have accumulated remarkable amounts of P_i and P_o . The proportion of P_o in algae ranged from 55.3 to 80.8% with a mean of 69.2% in Chao Lake.

Eight compounds P were detected in the NaOH–EDTA extracts of algal samples by ^{31}P NMR. The sum of orthophosphate and monoesters P in ETP was greater than 93% of algal P. Our observations implied that the release of P induced by the decomposition of algae debris could be a potential source of bioavailable P in aquatic systems of Chao Lake even without any more external P input. Thus, recycling of the potential bioavailable P in algae might be the mechanism of repeated algae blooming in eutrophic Chao Lake. Remediation of the lake requires a strategy to remove the algal biomass P effectively.

Acknowledgements This research was jointly supported by the National Natural Science Foundation of China (41703115, 41521003, 41630645, 41807372) and Postdoctoral Science Foundation of China (2017M610967).

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

- Abdi D, Cade-Menun BJ, Ziadi N, Parent LE (2014) Long-term impact of tillage practices and P fertilization on soil P forms as determined by ^{31}P -NMR spectroscopy. *J Environ Qual* 43:1432–1441
- Bai X, Sun J, Zhou Y, Gu L, Zhao H, Wang J (2017) Variations of different dissolved and particulate phosphorus classes during an algae bloom in a eutrophic lake by ^{31}P NMR spectroscopy. *Chemosphere* 169:577–585
- Baldwin DS (2013) Organic phosphorus in the aquatic environment. *Environ Chem* 10:439–454
- Bedrock CN, Cheshire MV, Chudek JA, Fraser AR, Goodman BA, Shand CA (1995) Effect of pH on precipitation of humic acid from peat and mineral soils on the distribution of phosphorus forms in humic and fulvic acid fractions. *Commun Soil Sci Plant Anal* 26:1411–1425
- Bell DW, Pellechia P, Chambers LR, Longo AF, McCabe KM, Ingall ED, Benitez-Nelson CR (2017) Isolation and molecular characterization of dissolved organic phosphorus using electro dialysis-reverse osmosis and solution ^{31}P -NMR. *Limnol Oceanogr Methods*. <https://doi.org/10.1002/lom3.10171>
- Busato JG, Canellas LP, Rumjanek VM, Velloso ACX (2005) Phosphorus in an inceptisol under long-term sugarcane: II. Humic acid analysis by NMR P-31. *Rev Bras Cienc Solo* 29:945–953
- Cade-Menun BJ (2015) Improved peak identification in ^{31}P -NMR spectra of environmental samples with a standardized method and peak library. *Geoderma* 257:102–114
- Cade-Menun BJ, Liu CW (2014) Solution phosphorus-31 nuclear magnetic resonance spectroscopy of soils from 2005 to 2013: a review of sample preparation and experimental parameters. *Soil Sci Soc Am J* 78:19–37
- Cade-Menun BJ, Preston CM (1996) A comparison of soil extraction procedures for ^{31}P NMR spectroscopy. *Soil Sci* 161:770–785
- Chen Y, Liu Q (2014) On the horizontal distribution of algal-bloom in Chaohu Lake and its formation process. *Acta Mech Sinica* 30:656–666
- Doolette A, Smernik R, Dougherty W (2009) Spiking improved solution phosphorus-31 nuclear magnetic resonance identification of soil phosphorus compounds. *Soil Sci Soc Am J* 73:919–927
- Feng W, Zhu Y, Wu F, Meng W, Giesy JP, He Z, Song L, Fan M (2016a) Characterization of phosphorus forms in lake macrophytes and algae by solution ^{31}P nuclear magnetic resonance spectroscopy. *Environ Sci Pollut Res* 23:7288–7297
- Feng W, Zhu Y, Wu F, He Z, Zhang C, Giesy JP (2016b) Forms and lability of phosphorus in algae and aquatic macrophytes characterized by solution ^{31}P NMR coupled with enzymatic hydrolysis. *Sci Rep* 6:37164
- Feng W, Wu F, He Z, Song F, Zhu Y, Giesy JP, Wang Y, Qin N, Zhang C, Chen H, Sun F (2018) Simulated bioavailability of phosphorus from aquatic macrophytes and phytoplankton by aqueous suspension and incubation with alkaline phosphatase. *Sci Total Environ* 616:1431–1439
- Giles CD, Cade-Menun BJ (2014) Applied manure and nutrient chemistry for sustainable agriculture and environment. In: Zhang H (ed) *Phytate in animal manure and soils: abundance, cycling and bioavailability*. Springer, The Netherlands, pp 163–190
- Giles CD, Lee LG, Cade-Menun BJ, Hill JE, Isles PD, Schroth AW, Druschel GK (2015) Characterization of organic phosphorus form and bioavailability in lake sediments using P nuclear magnetic resonance and enzymatic hydrolysis. *J Environ Qual* 44:1–13
- He Z, Honeycutt CW (2005) A modified molybdate blue method for orthophosphate determination suitable for investigating enzymatic hydrolysis of organic phosphates. *Commun Soil Sci Plant Anal* 36:1373–1383
- He Z, Cade-Menun BJ, Toor GS, Sim JT (2007) Comparison of phosphorus forms in wet and dried animal manures by solution phosphorus-31 nuclear magnetic resonance spectroscopy and enzymatic hydrolysis. *J Environ Qual* 36:1086–1095
- He Z, Honeycutt CW, Cade-Menun BJ, Senwo ZN, Tazisong IA (2008) Phosphorus in poultry litter and soil: enzymatic and nuclear magnetic resonance characterization. *Soil Sci Soc Am J* 72:1425–1433
- He Z, Honeycutt CW, Griffin TS, Cade-Menun BJ, Pellechia PJ, Dou Z (2009) Phosphorus forms in conventional and organic dairy

- manure identified by solution and solid state P-31 NMR spectroscopy. *J Environ Qual* 38:1909–1918
- He Z, Olk DC, Cade-Menun BJ (2011) Forms and lability of phosphorus in humic acid fractions of Hord silt loam soil. *Soil Sci Soc Am J* 75:1712–1722
- Jarosch KA, Doolette AL, Smernik RJ, Tamburini F, Frossard E, Büne-mann EK (2015) Characterisation of soil organic phosphorus in NaOH–EDTA extracts: a comparison of ³¹P NMR spectroscopy and enzyme addition assays. *Soil Biol Biochem* 91:298–309
- Kagalou I, Papastergiadoub E, Leonardasa I (2008) Long-term changes in the eutrophication process in a shallow Mediterranean lake ecosystem of W Greece: response after the reduction of external load. *J Environ Manag* 87:497–506
- Karl DM (2014) Microbially mediated transformations of phosphorus in the sea: new views of an old cycle. *Ann Rev Mar Sci* 6:279–337
- Lehman PW, Kurobe T, Lesmeister S, Baxa D, Tung A, Teh SJ (2017) Impacts of the 2014 severe drought on the *Microcystis* bloom in San Francisco estuary. *Harmful Algae* 63:94–108
- Li M, Wu Y, Yu Z, Sheng G, Yu H (2009) Enhanced nitrogen and phosphorus removal from eutrophic lake water by *Ipomoea aquatica* with low-energy ion implantation. *Water Res* 43:1247–1256
- Li J, Cui K, Lu W, Cheng Y, Jiang Y (2015) Community dynamics of spring-summer plankton in Lake Chaohu. *Acta Hydrobiol Sinica* 39:185–194 (in Chinese)
- Liu S, Zhu Y, Meng W, He Z, Feng W, Zhang C, Giesy JP (2016) Characteristics and degradation of carbon and phosphorus from aquatic macrophytes in lakes: insights from solid-state ¹³C NMR and solution ³¹P NMR spectroscopy. *Sci Total Environ* 543:746–756
- Liu S, Zhu Y, Wu F, Meng W, Wang H, He Z, Guo W, Song F, Giesy JP (2017) Using solid ¹³C NMR coupled with solution ³¹P NMR spectroscopy to investigate molecular species and lability of organic carbon and phosphorus from aquatic plants in Tai Lake. *China Environ Sci Pollut Res* 24:1880–1889
- Mahieu N, Olk DC, Randall EW (2000) Analysis of phosphorus in two humic acid fractions of intensively cropped lowland rice soils by ³¹P NMR. *Eur J Soil Sci* 51:391–402
- McDowell RW, Hill SJ (2015) Speciation and distribution of organic phosphorus in river sediments: a national survey. *J Soil Sediment* 15:2369–2379
- Pernet-Coudrier B, Qi WX, Liu HJ, Nüller B, Berg M (2012) Sources and pathway of nutrients in the semi-arid region of Beijing–Tianjin. *China Environ Sci Technol* 46:5294–5301
- Qu X, Xie L, Lin Y, Bai Y, Zhu Y, Xie F, Geisy JP, Wu F (2013) Quantitative and qualitative characteristics of dissolved organic matter from eight dominant aquatic macrophytes in Lake Dianchi. *China Environ Sci Pollut Res* 20:7413–7423
- Reitzel K, Ahlgren J, Gogoll A (2006) Effects of aluminum treatment on phosphorus, carbon, and nitrogen distribution in lake sediment: a ³¹P NMR study. *Water Res* 40:647–654
- Ruban V, López-Sánchez JF, Pardo P (2001) Harmonized protocol and certified reference material for the determination of extractable contents of phosphorus in freshwater sediments—a synthesis of recent works. *Fresen J Anal Chem* 370:224–228
- Schneider K, Resl P, Spribille T (2016) Escape from the cryptic species trap: lichen evolution on both sides of a cyanobacterial acquisition event. *Mol Ecol* 25:3453–3468
- Smernik RJ, Dougherty WJ (2007) Identification of phytate in phosphorus-31 nuclear magnetic resonance spectra: the need for spiking. *Soil Sci Soc Am J* 71:1045–1050
- Sørensen DR, Nielsen UG, Skou EM (2014) Solid state ³¹P MAS NMR spectroscopy and conductivity measurements on NbOPO₄ and H₃PO₄ composite materials. *J Solid State Chem* 219:80–86
- Suzumura M (2005) Phospholipids in marine environments: a review. *Talanta* 66:422–434
- Tang J, Shi T, Wu X, Cao H, Li X, Hua R, Tang F, Yue Y (2015) The occurrence and distribution of antibiotics in Lake Chaohu, China: seasonal variation, potential source and risk assessment. *Chemosphere* 122:154–161
- Turner BL, Richardson AE (2004) Identification of -inositol phosphates in soil by solution phosphorus-31 nuclear magnetic resonance spectroscopy. *Soil Sci Soc Am J* 68:802–808
- Turner BL, Mahieu N, Condrón LM (2003) Phosphorus-31 nuclear magnetic resonance spectral assignments of phosphorus compounds in soil NaOH–EDTA extracts. *Soil Sci Soc Am J* 67:497–510
- Turner BL, Cade-Menun BJ, Condrón LM, Newman S (2005) Extraction of soil organic phosphorus. *Talanta* 66:294–306
- Turner BL, Cheesman AW, Godage HY, Riley AM, Potter BVL (2012) Determination of neo- and D-chiro-inositol hexakisphosphate in soils by solution ³¹P NMR spectroscopy. *Environ Sci Technol* 46:4994–5002
- Vincent AG, Vestergren J, Gröbner G, Persson P, Schleucher J, Biesler R (2013) Soil organic phosphorus transformations in a boreal forest chronosequence. *Plant Soil* 367:149–162
- Wang J, Pant HK (2010) Enzymatic hydrolysis of organic phosphorus in river bed sediments. *Eco Eng* 36:963–968
- Wang X, Xi B, Huo S, Deng L, Pan H, Xia X, Zhang J, Ren Y, Liu H (2013) Polybrominateddiphenyl ethers occurrence in major inflowing rivers of Lake Chahu (China): characteristics, potential sources and inputs to lake. *Chemosphere* 93:1624–1631
- Xu M, Cao H, Xie P, Deng D, Feng W, Xu J (2005) The temporal and spatial distribution, composition and abundance of protozoa in Chaohu Lake, China: relationship with eutrophication. *Europ J Protistol* 41:183–192
- Yang L, Han X, Sun P, Yan W, Li Y (2011) Canonical correspondence analysis of algae community and its environmental factors in Lake Chaohu. *China J Agro-Environ Sci* 30:952–958
- Young EO, Ross DS, Cade-Menun BJ, Liu CW (2013) Phosphorus speciation in riparian soils: a phosphorus-31 nuclear magnetic resonance spectroscopy and enzyme hydrolysis study. *Soil Sci Soc Am J* 77:1636–1647
- Zan F, Huo S, Xi B, Li Q, Liao H, Zhang J (2010) Phosphorus distribution in the sediments of a shallow eutrophic lake. *Lake Chaohu, China. Environ Earth Sci* 62:1643–1653
- Zhang WQ, Xing BS (2013) Detection of phosphorus species in the sediments of Chaohu Lake by ³¹P nuclear magnetic resonance spectroscopy (31P-NMR). *Acta Sci Circum* 33:1967–1973 (in Chinese)
- Zhang WQ, Zhu XL, Jin X, Meng X, Tang WZ, Shan BQ (2017) Evidence for organic phosphorus activation and transformation at the sediment-water interface during plant debris decomposition. *Sci Total Environ* 583:458–465
- Zhong W, Zhang Z, Luo Y (2012) Biogas productivity by co-digesting Taihu blue algae with corn straw as an external carbon source. *Bioresour Technol* 114:281–286
- Zhu G, Qin B, Zhang L, Luo L (2006) Geochemical forms of phosphorus in sediments of three large, shallow lakes of China. *Pedosphere* 16:726–734
- Zhu Y, Wu F, He Z, Guo J, Qu X, Xie F, Giesy JP, Liao H, Guo F (2013) Characterization of organic phosphorus in lake sediments by sequential fractionation and enzymatic hydrolysis. *Environ Sci Technol* 47:7679–7687
- Zhu Y, Wu F, He Z (2015) Bioavailability and preservation of organic phosphorus in freshwater sediments and its role in lake eutrophication. In: He Z, Wu F (eds) *Labile organic matter—chemical compositions, function, and significance in soil and the environment*. SSSA, Madison, pp 275–294
- Zhu Y, Wu F, Feng W, Liu S, Giesy JP (2016) Interaction of alkaline phosphatase with minerals and sediments: activities, kinetics and hydrolysis of organic phosphorus. *Colloids surf A: Physicochem Eng Aspects* 495:46–53