

# Chapter 1

## Introduction

Taicheng An, Huijun Zhao, and Po Keung Wong

### 1.1 Water Disinfection

The last 50 years have witnessed a growing awareness of the fragile state of most of the planets' drinking water resources. Access to freshwater will become even more important in the near future, as the world's population rises from 7 billion today to 9 billion by 2050. The World Health Organization (WHO) has estimated that 80 % of illnesses in the developing world are water related, resulting from poor water quality and lack of sanitation [1]. There are 3.3 million deaths each year from diarrheal diseases caused by bacteria such as *Escherichia coli*, *Salmonella* sp. and *Cholera* sp., parasites and viral pathogens. In the 1990s, the number of children who died of diarrhoea was greater than the sum of people killed in conflicts since World War II [2]. It is also estimated that around 4 billion people worldwide experience to have no or little access to clean and sanitized water supply, and millions of people died of severe waterborne diseases annually [3, 4].

Waterborne diseases are caused by pathogenic microorganisms that most commonly are transmitted in contaminated freshwater. The pathogenic microorganisms responsible for these diseases include a variety of helminthes, protozoa, fungi,

---

T. An

Institute of Environmental Health and Pollution Control, School of Environmental Science and Engineering, Guangdong University of Technology, Guangzhou 510006, Guangdong, China  
e-mail: [antc99@gdut.edu.cn](mailto:antc99@gdut.edu.cn); [antc99@gig.ac.cn](mailto:antc99@gig.ac.cn)

H. Zhao

Centre for Clean Environment and Energy, Griffith University, Gold Coast Campus,  
Gold Coast, QLD 4222, Australia  
e-mail: [h.zhao@griffith.edu.au](mailto:h.zhao@griffith.edu.au)

P.K. Wong (✉)

School of Life Sciences, The Chinese University of Hong Kong, Shatin, N.T.,  
Hong Kong SAR, China  
e-mail: [pkwong@cuhk.edu.hk](mailto:pkwong@cuhk.edu.hk)

bacteria, rickettsiae, viruses and prions [1, 5], many of which are intestinal parasites or invade the tissues or circulatory system through walls of the digestive tract. Water disinfection means the removal, deactivation or killing of pathogenic microorganisms, resulting in termination of growth and reproduction. Problems with waterborne diseases are expected to grow worse in the future, both in developing and industrialized nations. Therefore, effective and lower-cost methods to disinfect microorganism-contaminated waters are urgently needed, without further stressing the environment or endangering human health by the treatment itself [6].

## 1.2 Traditional Water Disinfection Methods

The existing drinking water pretreatment processes, such as coagulation, flocculation and sedimentation, can remove a maximum of 90 % of bacteria, 70 % of viruses and 90 % of protozoa [4]. Filtration for drinking water treatment (e.g. sand and membrane filtration), with proper design and adequate operation, can act as a consistent and effective barrier for microbial pathogens leading to about 90 % removal of bacteria. However, the remaining bacteria might still be able to cause disease, which makes filtration a good pretreatment, but not a completely safe disinfection technique [7]. The most commonly used drinking water disinfection techniques after pretreatment include chlorination (chlorine and derivatives), ozonation and UVC irradiation.

### 1.2.1 Chlorination

Chlorine is a very effective disinfectant for most microorganisms. It is reported that 99 % of bacterial cells can be killed with chlorine of 0.08 mg/min/L at 1–2 °C under neutral pH condition. In addition, 99 % of viruses can be killed by 12 mg/min/L chlorine at 0–5 °C under neutral pH condition. However, the protozoa including *Cryptosporidium*, *Giardia* and *Acanthamoeba* are quite resistant to chlorination and cannot be effectively inactivated [7]. Another major disadvantage of chlorination is the formation of potentially mutagenic and carcinogenic disinfection byproducts (DBPs) during water chlorination, which can lead to the problems of recontamination and salting of freshwater sources [8, 9]. The DBPs are formed from the reaction of chlorine with natural organics in water and include trihalo-methanes (THMs) and haloacetic acids (HAAs). US Environmental Protection Agency (USEPA) regulations have further limited THMs, HAAs and other DBPs (including chlorite and bromate) in drinking water [10]. As a result, many water systems now limit the use of chlorine to high-quality groundwater or reduce total organic carbon prior to disinfection.

### 1.2.2 Ozonation

The application of ozone is another widespread disinfection method for drinking water treatment throughout the world [11]. Similar to chlorination, ozone is unstable in water and undergoes reactions with some water matrix components. However, the unique feature of ozone is its decomposition into hydroxyl radicals ( $\bullet\text{OH}$ ), which are the strongest oxidants in water [12]. While disinfection occurs dominantly through ozone, oxidation processes may occur through both ozone and  $\bullet\text{OH}$  [13], making the ozonation even more effective than  $\text{Cl}_2$  in destroying bacterial cells and viruses [14, 15]. It is reported that 99 % of bacterial cells can be removed with 0.02 mg/min/L ozone at 5 °C under neutral pH condition. For the disinfection of protozoa *Cryptosporidium*, the required ozone concentration is suggested to be 40 mg/min/L at 1 °C [16]. Despite its highly efficient inactivation of all microorganisms, ozonation can also produce DBPs, such as aldehydes, carboxylic acids and ketones, in the presence of dissolved organic matter [17]. However, as ozonation is usually followed by biological filtration, some organic compounds can be mineralized microbiologically. Thus, the most important ozonation DBP regulated in drinking waters today is bromate, which is formed during ozonation of bromide-containing waters and cannot be degraded in biological filtration process [18, 19]. In addition, ozonation is a more complex technology than chlorination and is often associated with increased costs and process complexity [20].

### 1.2.3 UV Irradiation

Water disinfection utilizing germicidal UV irradiation has become more and more important in recent years, as the low-pressure UV produces almost no disinfection byproducts [21]. In addition, unlike chemical disinfectants, the biological stability of the water is not affected by low-pressure lamps. In Europe, UV has been widely applied for drinking water disinfection since the 1980s, for the control of incidental contamination of vulnerable groundwater and for the reduction of heterotrophic plate counts [22]. Depending on irradiation wavelengths, UV can be divided into UVA (315–400 nm), UVB (280–315 nm), UVC (200–280 nm) and vacuum UV (VUV) (100–200 nm). In particular, UVC is the most effective wavelength for microorganism inactivation, as UVC light will damage irradiated DNA, directly inducing pyrimidine and purine dimers and pyrimidine adducts. For water disinfection, 99 % inactivation of bacterial cells can be achieved at UVC intensity of 7 mJ/cm<sup>2</sup>. The susceptibility of protozoa to UVC damage is very similar to that of bacteria; thus, the 99 % inactivation for *Cryptosporidium* can be achieved at 5 mJ/cm<sup>2</sup> [23]. However, due to the weak penetration power, UV disinfection can only inactivate bacterial cells on the surface of the wastewater [24], and the treated cells can often regrow after removal of UV irradiation [25]. General application of UV

disinfection was further hampered because of high costs, poor equipment reliability and maintenance problems [26, 27].

Therefore, although traditional disinfection methods can be effectively applied in water disinfection, the disadvantages of these methods must be considered when selecting suitable disinfection methods for water treatment, and alternative technologies are needed.

### 1.3 Advanced Oxidation Process

Advanced oxidation processes (AOPs) are defined as the processes that generate hydroxyl radicals ( $\bullet\text{OH}$ ) in sufficient quantities to be able to oxidize the majority of the complex chemicals present in the effluent water [28]. AOPs have been receiving increasing attention to be effectively applied in the near-ambient total degradation of soluble organic contaminants from waters and soils, as the produced  $\bullet\text{OH}$  would be able to oxidize almost all organic compounds to carbon dioxide and water because of its powerful redox potential (2.8 V vs. NHE) [29]. These processes include cavitation [30, 31], photo-Fenton [32, 33], photocatalytic oxidation [34] and other combination methods, such as  $\text{H}_2\text{O}_2/\text{UV}$ ,  $\text{O}_3/\text{UV}$  and  $\text{H}_2\text{O}_2/\text{O}_3/\text{UV}$ , which utilize the photolysis of  $\text{H}_2\text{O}_2$  and  $\text{O}_3$  to produce  $\bullet\text{OH}$  [35]. In particular, heterogeneous photocatalysis based on the use of a semiconductor with suitable energy band gap ( $E_g$ ) is the most interesting and promising advanced oxidation technology that has received much attention in the past few decades for a variety of photochemical applications, including water splitting, organic compounds degradation and  $\text{CO}_2$  reduction, as well as water disinfection.

### 1.4 Photocatalysis

With respect to the generally accepted definition of thermal catalysis, photocatalysis can be defined as “acceleration of a photoreaction by the presence of a catalyst”, which indicates both light and a catalyst are necessary to bring about or to accelerate a chemical transformation [36]. As the photoreaction takes place in more than one homogeneous medium, it is usually called “heterogeneous photocatalysis” [37, 38].

Fujishima and Honda (1972) [39] discovered the photocatalytic splitting of water on  $\text{TiO}_2$  electrodes, which has marked the beginning of heterogeneous photocatalysis [40]. Since then, tremendous research efforts have been devoted into understanding the fundamental process of heterogeneous photocatalysis, thus enhancing the photocatalytic efficiencies [41–44]. Photocatalysis was initially applied in hydrogen evolution by splitting water, with intention to address the energy crisis [45–48]. Research activities were soon extended to photocatalytic oxidation of organic pollutants [49, 50],  $\text{CO}_2$  reduction [51] and the disinfection of microorganisms in contaminated water [52, 53]. Although an early study

demonstrated that there was no improved antimicrobial activity of TiO<sub>2</sub> for the disinfection of primary wastewater effluent [54], a number of subsequent studies have shown the effectiveness of TiO<sub>2</sub> photocatalysis for water disinfection [55, 56], including inactivation of bacterial cells [57] and viruses from contaminated water [58], tertiary treatment of wastewater [59], purifying drinking water [60], treatment of wash waters from vegetable preparation [61] and in bioreactor design to prevent biofilm formation [62].

### 1.4.1 Fundamental Mechanism for TiO<sub>2</sub> Photocatalysis

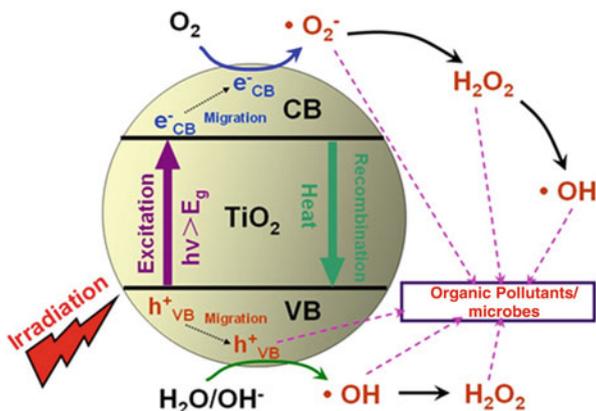
Semiconductors acting as the photocatalysts for the light-reduced redox processes, such as TiO<sub>2</sub>, ZnO, Fe<sub>2</sub>O<sub>3</sub>, CdS and ZnS, are characterized by a filled valence band and an empty conduction band [63]. When the valence band receives a photon with energy bigger than the band gap, an electron (e<sup>-</sup>) will be excited and promoted into the conduction band, leaving a hole (h<sup>+</sup>) in the valence band. The photo-generated e<sup>-</sup>-h<sup>+</sup> pairs will subsequently migrate onto the surface of photocatalyst and undergo a variety of complicated reactions to produce reactive oxidative species (ROSs), which are potentially involved in the photocatalytic oxidation process. The most widely used photocatalyst is TiO<sub>2</sub>, as it is nontoxic, low cost and highly efficient and has long-term photostability [64, 65]. The fundamental mechanism for TiO<sub>2</sub> photocatalysis under UV irradiation has been well established for photocatalytic oxidation process towards organic compounds degradation as well as microorganism inactivation (Fig. 1.1) [38, 66].

The primary photocatalytic oxidation mechanism includes the following four steps (Eqs. 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 1.10, 1.11 and 1.12):

#### 1. Irradiation

The first step is the light irradiation process for harvesting and conversion of light energy to chemical energy, thus leading to the generation of e<sup>-</sup>-h<sup>+</sup> pairs.

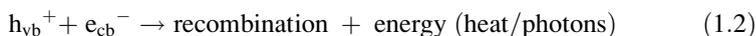
**Fig. 1.1** A schematic diagram showing the photocatalytic oxidation mechanism of TiO<sub>2</sub> photocatalysis under UV irradiation



The requirement of this step is the incoming photon should have an energy of  $h\nu$  that matches or exceeds the semiconductor band gap energy. For  $\text{TiO}_2$ , the light wavelength for fulfilment of the excitation process is restricted to the UV region because of its wide band gap (3.2 eV) [67].

## 2. Separation and recombination of $e^-$ - $h^+$ pairs

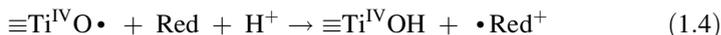
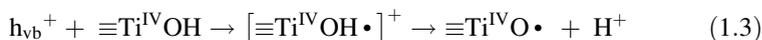
The photoexcited  $e^-$  is injected into the conduction band, leading to the separation of  $e^-$ - $h^+$  pairs. However, the photo-generated  $e^-$  and  $h^+$  can recombine in bulk or on surface of the semiconductor within extremely short time, releasing energy in the form of heat or photons (Eqs. 1.1 and 1.2) [68, 69].



The separated  $e^-$  and  $h^+$  without recombination are migrated to the surface of  $\text{TiO}_2$  and trigger photochemical reactions to produce secondary reactive species (i.e. ROSS) or directly oxidize/reduce the substrates adsorbed by the  $\text{TiO}_2$ .

## 3. $h^+$ trapping reactions

In the valence band, the separated  $h^+$  is migrated to the surface and trapped by surface-adsorbed hydroxyl groups or water to produce trapped holes ( $\equiv\text{Ti}^{\text{IV}}\text{O}\cdot$ ) (Eq. 1.3), which is usually described as a surface-bound or surface-adsorbed hydroxyl radical ( $\cdot\text{OH}_{\text{ads}}$ ) [70–72]. When electron donors (Red) (i.e. reductants) are available on the  $\text{TiO}_2$  surface, the photocatalytic oxidation process thus happens by electron transferring from Red to trapped holes (Eq. 1.4). The subsequent release of  $\cdot\text{OH}_{\text{ads}}$  to bulk solution, thus leading to the formation of bulk hydroxyl radical ( $\cdot\text{OH}_{\text{bulk}}$ ), is suggested to contribute to the oxidation process (Eqs. 1.5, 1.6 and 1.7) [73]. On the other hand,  $h^+$  can also be directly involved in oxidation of Red [74] and indirectly involved in production of  $\text{H}_2\text{O}_2$  by coupling of two  $\cdot\text{OH}$  (Eqs. 1.8 and 1.9) [75–77].



## 4. $e^-$ trapping reactions

In the conduction band,  $\text{O}_2$  often acts as the electron acceptor to trap the photoexcited  $e_{\text{cb}}^-$  in aerated systems, thus preventing the  $e^-$ - $h^+$  recombination. In this process,  $\cdot\text{O}_2^-$  is formed and undergoes a variety of reactions to produce  $\text{H}_2\text{O}_2$  (Eqs. 1.10, 1.11, 1.12 and 1.13) [78, 79]. Meanwhile, the as-generated

H<sub>2</sub>O<sub>2</sub> can also produce the highly reactive •OH by reduction or cleaving (Eqs. 1.14 and 1.15) [80–82].



During the overall photochemical process, the photo-generated e<sup>-</sup>/h<sup>+</sup> and the produced ROSs such as •OH, •O<sub>2</sub><sup>-</sup>, •OOH and H<sub>2</sub>O<sub>2</sub> are suggested to be responsible for the oxidation of organic pollutants, including synthetic dyes and pathogenic microorganisms in aqueous media. The importance of •OH as the oxidation agent was particularly attended by researchers in this typical mechanism model of photocatalytic oxidation in UV irradiation TiO<sub>2</sub> systems [38, 83, 84].

### 1.4.2 Photocatalytic Water Disinfection

Photocatalysis was first shown to be an effective disinfection process by Matsunaga et al. (1985) [53], who reported on the inactivation of *Lactobacillus acidophilus*, *Saccharomyces cerevisiae* and *Escherichia coli* by Pt-loaded TiO<sub>2</sub>. Since then, a concerted range of research has been conducted on the development of photocatalysis for water disinfection. Photocatalytic disinfection of a wide range of bacteria and yeasts including *Escherichia coli* [85, 86], *Candida albicans* [87], *Enterococcus faecium*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* [24], *Streptococcus faecalis* [88], *Streptococcus mutans* [89], *Salmonella choleraesuis*, *Vibrio parahaemolyticus* and *Listeria monocytogenes* [90] as well as poliovirus [91] has been reported. The inactivation of the protozoan of *Cryptosporidium* and *Giardia*, known for their resistance to many chemical disinfectants, including chlorine, was also reported in recent years [92–94].

As the archetypical photocatalyst for water splitting and organic compounds degradation, TiO<sub>2</sub> also holds the preponderant position in water disinfection for destruction of various microorganism including bacteria (both Gram-negative and Gram-positive), fungi, algae, protozoa and viruses as well as microbial toxins [56]. Table 1.1 shows the typical examples of TiO<sub>2</sub> photocatalysis for microorganism inactivation. For all the inactivation of microorganism reported so far, only *Acanthamoeba* cysts and *Trichoderma asperellum* conidiospores were found to be resistant to photocatalysis [95, 96]. There are three crystal phases of TiO<sub>2</sub>: anatase, rutile and brookite, in which anatase shows the highest photocatalytic activity [97].

**Table 1.1** Typical examples of microorganism inactivation caused by TiO<sub>2</sub> photocatalysis [56]

Microorganism	Photocatalysts	References
<b>Bacteria (Gram-negative)</b>		
<i>Escherichia coli</i>	Degussa P25 suspension	[98]
<i>Escherichia coli</i>	TiO <sub>2</sub> -impregnated cloth filter	[99]
<i>Enterobacter aerogenes</i>	Degussa P25 suspension	[100]
<i>Flavobacterium</i> sp.	TiO <sub>2</sub> -coated glass beads	[101]
<i>Fusobacterium nucleatum</i>	Anatase TiO <sub>2</sub> thin film	[102]
<i>Pseudomonas aeruginosa</i>	TiO <sub>2</sub> -coated soda lime glass and silica tubing	[103, 104]
<i>Legionella pneumophila</i>	Degussa P25 suspension	[105]
<i>Porphyromonas gingivalis</i>	TiO <sub>2</sub> sol/gel-coated orthodontic wires	[106]
<i>Vibrio vulnificus</i>	TiO <sub>2</sub> -impregnated steel fibres	[107]
<b>Bacteria (Gram-positive)</b>		
<i>Actinobacillus actinomycetemcomitans</i>	TiO <sub>2</sub> coated on Ti substrates	[102]
<i>Bacillus cereus</i>	TiO <sub>2</sub> suspension	[108]
<i>Streptococcus cricetus</i>	Kobe Steel TiO <sub>2</sub>	[109]
<i>Streptococcus mutans</i>	TiO <sub>2</sub> thin film	[110]
<i>Clostridium difficile</i>	Evonik Aeroxide P25 thin film	[111]
<i>Clostridium perfringens</i> spores	Degussa P25 suspension	[112]
<i>Bacillus subtilis</i> endospore	TiO <sub>2</sub> coated on Al foil	[113]
<b>Fungi</b>		
<i>Aspergillus niger</i>	TiO <sub>2</sub> coated on wood	[114]
<i>Aspergillus niger</i> spores	Degussa P25 film on quartz discs	[62]
<i>Candida famata</i>	TiO <sub>2</sub> -coated catheters	[115]
<i>Candida albicans</i>	TiO <sub>2</sub> thin film	[24]
<i>Penicillium citrinum</i>	TiO <sub>2</sub> -coated air filter	[116]
<i>Trichoderma asperellum</i>	TiO <sub>2</sub> -coated concrete	[96]
<b>Protozoa</b>		
<i>Cryptosporidium parvum</i>	Nanostructured TiO <sub>2</sub> films	[117]
<i>Giardia</i> sp.	Fibrous ceramic TiO <sub>2</sub> filter	[94]
<i>Giardia lamblia</i>	TiO <sub>2</sub> thin film	[118]
<i>Acanthamoeba castellanii</i>	Degussa P25 suspension	[95]
<b>Algae</b>		
<i>Cladophora</i> sp.	TiO <sub>2</sub> -coated glass	[119]
<i>Chroococcus</i> sp.	Anatase TiO <sub>2</sub>	[120]
<i>Oedogonium</i> sp.	TiO <sub>2</sub> -coated concrete	[121]
<i>Melosira</i> sp.	TiO <sub>2</sub> -coated glass	[122]
<b>Virus</b>		
Influenza A/H5N2	Degussa P25/TiO <sub>2</sub> Millennium PC500	[123]
<i>E. coli</i> coliphage	Degussa P25 suspension	[112]
<i>E. coli</i> MS2	TiO <sub>2</sub> suspension	[124]
<i>E. coli</i> λ vi	Degussa P25 suspension	[125]
Influenza A/H1N1	TiO <sub>2</sub> suspension	[126]

(continued)

**Table 1.1** (continued)

Microorganism	Photocatalysts	References
Influenza A/H3N2	TiO <sub>2</sub> /Pt-TiO <sub>2</sub>	[127]
SARS coronavirus	Titanium apatite filter	[128]
<b>Toxins</b>		
Brevetoxins	Degussa P25 suspension	[129]
Microcystins LR, YR and YA	Degussa P25 suspension	[130]
Nodularin	Degussa P25 suspension	[131]

However, the most active and commercially available TiO<sub>2</sub> is P25 (Degussa Ltd., Germany), consisting of 80 % anatase and 20 % rutile. The improved activity of mixed crystal phases is generally ascribed to interactions between the two forms, thus preventing bulk recombination. For catalyst immobilization, TiO<sub>2</sub> is often coated on various supports, including glass plate, cloth filter, steel substrates, silica, wood, catheter, concrete, etc.

Although exciting progress has been made in TiO<sub>2</sub> photocatalysis for microorganism disinfection, challenges still pose in achieving photocatalytic water disinfection utilizing solar energy. Unfortunately, the most widely used TiO<sub>2</sub> is only active under UV irradiation which accounts for only 4 % of the sunlight spectrum, while 45 % of the sunlight spectrum is visible light. TiO<sub>2</sub> modification techniques have been attempted to shift its light absorption capacity towards visible wavelengths, while considerable scientific interests have been devoted to the development of new types of photocatalyst that is active under visible light irradiation. This opens avenue for designing and fabricating nanostructured materials that can be used in photocatalytic water disinfection by employing material science and nanotechnology [132–134].

### ***1.4.3 Advances in Photocatalytic Disinfection***

In this book, some of the key development of photocatalytic disinfection in the last decade will be presented and discussed. The use of naturally occurring minerals or novel synthetic catalysts for effective microbial disinfection will be compiled. In addition, the mechanism, catalysts and performance of microbial disinfection by photoelectrocatalytic process will be presented and discussed. Finally, how to apply modelling approaching to study the kinetics of the photocatalytic disinfection will be included in this book. With all these updated information, the useful information and data will be provided to the people in academic, engineering and technical sectors.

## References

1. World Health Organization (2003) Emerging issues in water and infectious disease 1–22. World Health Organization, Geneva
2. Smith A (2009) Nanotechnology: an answer to the world's water crisis. *Chem Int* 31:12–14
3. Montgomery MA, Elimelech M (2007) Water and sanitation in developing countries: including health in the equation. *Environ Sci Technol* 41:17–24
4. Malato S, Fernandez-Ibanez P, Maldonado MI, Blanco J, Gernjak W (2009) Decontamination and disinfection of water by solar photocatalysis: recent overview and trends. *Catal Today* 147:1–59
5. Pitman GK (2002) Bridging troubled waters – assessing the World Bank water resources strategy. World Bank Publications, Washington, DC
6. Shannon MA, Bohn PW, Elimelech M, Georgiadis JG, Marinas BJ, Mayes AM (2008) Science and technology for water purification in the coming decades. *Nature* 452:301–310
7. World Health Organization (2006) Guidelines for drinking-water quality first addendum to third edition 1 recommendations. World Health Organization, Library Cataloguing-in-Publication Data
8. Bryant EA, Fulton GP, Budd GC (1992) Disinfection alternatives for safe drinking water. van Nostrand Reinhold, New York
9. Nieuwenhuijsen MJ, Toledano MB, Eaton NE, Fawell J, Elliott P (2000) Chlorination disinfection byproducts in water and their association with adverse reproductive outcomes: a review. *Occup Environ Med* 57:73–85
10. Public Law (1996) Safe drinking water act amendments of 1996, 104–182, 1620–1621
11. Camel V, Bermond A (1998) The use of ozone and associated oxidation processes in drinking water treatment. *Water Res* 32:3208–3222
12. Staehelin J, Hoigné J (1985) Decomposition of ozone in water in the presence of organic solutes acting as promoters and inhibitors of radical chain reactions. *Environ Sci Technol* 19:1206–1213
13. Hoigné J (1998) Chemistry of aqueous ozone, and transformation of pollutants by ozonation and advanced oxidation processes. In: Hubrec J (ed) The handbook of environmental chemistry quality and treatment of drinking water. Springer, Berlin, pp 341–368
14. United States Environmental Protection Agency (1991) Guidance manual for compliance with the filtration and disinfection requirements for public water systems using surface water sources. Office of Drinking Water, United States Environmental Protection Agency, Washington, DC
15. United States Environmental Protection Agency (2003) Long term 2 enhanced surface water treatment rule toolbox guidance manual (DRAFT). Office of Drinking Water, United States Environmental Protection Agency, Washington, DC
16. World Health Organization, International Programme on Chemical Safety (IPCS) (1999) Disinfectants and disinfectant by-products, international program on chemical safety (Environmental Health Criteria 216), Geneva
17. Huang WJ, Fang GC, Wang CC (2005) The determination and fate of disinfection by-products from ozonation of polluted raw water. *Sci Total Environ* 345:261–272
18. Richardson SD, Thruston AD, Caughran TV, Chen PH, Collette TW, Floyd TL, Schenck KM, Lykins BW, Sun GR, Majetich G (1999) Identification of new ozone disinfection byproducts in drinking water. *Environ Sci Technol* 33:3368–3377
19. von Gunten U (2003) Ozonation of drinking water: Part I. Oxidation kinetics and product formation. *Water Res* 37:1443–1467
20. Sichel C, Blanco J, Malato S, Fernández-Ibáñez P (2007) Effects of experimental conditions on *E. coli* survival during solar photocatalytic water disinfection. *J Photochem Photobiol A Chem* 189:239–246
21. Hijnen WAM, Beerendonk EF, Medema GJ (2006) Inactivation credit of UV radiation for viruses, bacteria and protozoan (oo)cysts in water: a review. *Water Res* 40:3–22

22. Kruithof JC, Van der Leer RC, Hijnen WAM (1992) Practical experiences with UV disinfection in The Netherlands. *J Water Supply Res Technol Aqua* 41:88–94
23. Masschelin WJ, Rice RG (2002) *Ultraviolet light in water and wastewater sanitation*. Lewis Publishers, Boca Raton
24. Kühn KP, Chaberny IF, Massholder K, Stickler M, Benz VW, Sonntag HG, Erdinger L (2003) Disinfection of surfaces by photocatalytic oxidation with titanium dioxide and UVA light. *Chemosphere* 53:71–77
25. Hancock GG, Davis EM (1999) Regrowth potential of coliforms after UV disinfection of municipal wastewater. *J Environ Sci Health, Part A: Tox Hazard Subst Environ Eng* 34:1737–1743
26. Wolfe RL (1990) Ultraviolet disinfection of potable water – current technology and research needs. *Environ Sci Technol* 24:768–772
27. Hoyer O (2004) Water disinfection with UV radiation – requirements and realization. In: *Proceedings of the European conference UV Karlsruhe, UV radiation. Effects and technologies, September 2003, Karlsruhe*
28. Gogate PR, Pandit AB (2004) A review of imperative technologies for wastewater treatment I: oxidation technologies at ambient conditions. *Adv Environ Res* 8:501–551
29. Pera-Titus M, Garcia-Molina V, Banos MA, Gimenez J, Esplugas S (2004) Degradation of chlorophenols by means of advanced oxidation processes: a general review. *Appl Catal B Environ* 47:219–256
30. Adewuyi YG (2001) *Sonochemistry: environmental science and engineering applications*. *Ind Eng Chem Res* 40:4681–4715
31. Gogate PR (2002) Cavitation: an auxiliary technique in wastewater treatment schemes. *Adv Environ Res* 6:335–358
32. Venkatadri R, Peters RW (1993) Chemical oxidation technologies – ultraviolet-light hydrogen-peroxide, fenton reagent, and titanium dioxide-assisted photocatalysis. *Hazard Waste Hazard Mater* 10:107–149
33. Nesheiwat FK, Swanson AG (2000) Clean contaminated sites using Fenton’s reagent. *Chem Eng Prog* 96:61–66
34. Bhatkhande DS, Pangarkar VG, Beenackers A (2002) Photocatalytic degradation for environmental applications – a review. *J Chem Technol Biotechnol* 77:102–116
35. Trapido M, Hirvonen A, Veressinina Y, Hentunen J, Munter R (1997) Ozonation, ozone/UV and UV/H<sub>2</sub>O<sub>2</sub> degradation of chlorophenols. *Ozone Sci Eng* 19:75–96
36. Kisch H (1989) What is photocatalysis? In: Serpone N, Pelizzetti E (eds) *Photocatalysis: fundamentals and applications*. Wiley, New York
37. Fox MA, Dulay MT (1993) Heterogeneous photocatalysis. *Chem Rev* 93:341–357
38. Hoffmann MR, Martin ST, Choi WY, Bahnemann DW (1995) Environmental applications of semiconductor photocatalysis. *Chem Rev* 95:69–96
39. Fujishima A, Honda K (1972) Electrochemical photolysis of water at a semiconductor electrode. *Nature* 37:238
40. Linsebigler AL, Lu GQ, Yates JT (1995) Photocatalysis on TiO<sub>2</sub> surfaces – principles, mechanisms, and selected results. *Chem Rev* 95:735–758
41. Mills A, LeHunte S (1997) An overview of semiconductor photocatalysis. *J Photochem Photobiol A Chem* 108:1–35
42. Sakthivel S, Kisch H (2003) Daylight photocatalysis by carbon-modified titanium dioxide. *Angew Chem Int Ed* 42:4908–4911
43. Kamat PV (2007) Meeting the clean energy demand: nanostructure architectures for solar energy conversion. *J Phys Chem C* 111:2834–2860
44. Chen XB, Liu L, Yu PY, Mao SS (2011) Increasing solar absorption for photocatalysis with black hydrogenated titanium dioxide nanocrystals. *Science* 331:746–750
45. Bard AJ (1979) Photoelectrochemistry and heterogeneous photocatalysis at semiconductors. *J Photochem* 10:59–75
46. Bard AJ (1980) Photoelectrochemistry. *Science* 207:139–144

47. Bard AJ (1982) Design of semiconductor photo-electrochemical systems for solar-energy conversion. *J Phys Chem* 86:172–177
48. Kalyanasundaram K, Gratzel M, Pelizzetti E (1986) Interfacial electron-transfer in colloidal metal and semiconductor dispersions and photodecomposition of water. *Coord Chem Rev* 69:57–125
49. Carey JH, Lawrence J, Tosine HM (1976) Photo-dechlorination of PCBs in presence of titanium-dioxide in aqueous suspensions. *Bull Environ Contam Toxicol* 16:697–701
50. Frank SN, Bard AJ (1977) Heterogeneous photocatalytic oxidation of cyanide ion in aqueous-solutions at TiO<sub>2</sub> powder. *J Am Chem Soc* 99:303–304
51. Inoue T, Fujishima A, Konishi S, Honda K (1979) Photoelectrocatalytic reduction of carbon-dioxide in aqueous suspensions of semiconductor powders. *Nature* 277:637–638
52. Matusunga T (1985) Sterilization with particulate photosemiconductor. *J Antibact Antifung Agents* 13:211–220
53. Matsunaga T, Tomoda R, Nakajima T, Wake H (1985) Photoelectrochemical sterilization of microbial-cells by semiconductor powders. *FEMS Microbiol Lett* 29:211–214
54. Carey JH, Oliver BG (1980) The photochemical treatment of waste water by ultraviolet irradiation of semiconductors. *Water Pollut Res J Can* 15:157–185
55. Baram N, Starosvetsky D, Starosvetsky J, Epshtein M, Armon R, Ein-Eli Y (2011) Photocatalytic inactivation of microorganisms using nanotubular TiO<sub>2</sub>. *Appl Catal B Environ* 101:212–219
56. Foster HA, Ditta IB, Varghese S, Steele A (2011) Photocatalytic disinfection using titanium dioxide: spectrum and mechanism of antimicrobial activity. *Appl Microbiol Biotechnol* 90:1847–1868
57. Chung CJ, Lin HI, Chou CM, Hsieh PY, Hsiao CH, Shi ZY, He JL (2009) Inactivation of *Staphylococcus aureus* and *Escherichia coli* under various light sources on photocatalytic titanium dioxide thin film. *Surf Coat Technol* 203:1081–1085
58. Li QL, Mahendra S, Lyon DY, Brunet L, Liga MV, Li D, Alvarez PJJ (2008) Antimicrobial nanomaterials for water disinfection and microbial control: potential applications and implications. *Water Res* 42:4591–4602
59. Arana J, Melian JAH, Rodriguez JMD, Diaz OG, Viera A, Pena JP, Sosa PMM, Jimenez VE (2002) TiO<sub>2</sub>-photocatalysis as a tertiary treatment of naturally treated wastewater. *Catal Today* 76:279–289
60. Lonnen J, Kilvington S, Kehoe SC, Al-Touati F, McGuigan KG (2005) Solar and photocatalytic disinfection of protozoan, fungal and bacterial microbes in drinking water. *Water Res* 39:877–883
61. Selma MV, Allende A, Lopez-Galvez F, Conesa MA, Gil MI (2008) Heterogeneous photocatalytic disinfection of wash waters from the fresh-cut vegetable industry. *J Food Prot* 71:286–292
62. Wolfrum EJ, Huang J, Blake DM, Maness PC, Huang Z, Fiest J, Jacoby WA (2002) Photocatalytic oxidation of bacteria, bacterial and fungal spores, and model biofilm components to carbon dioxide on titanium dioxide-coated surfaces. *Environ Sci Technol* 36:3412–3419
63. Boer KW (1990) Survey of semiconductor physics. van Nostrand Reinhold, New York
64. McLoughlin OA, Ibanez PF, Gernjak W, Rodriguez SM, Gill LW (2004) Photocatalytic disinfection of water using low cost compound parabolic collectors. *Sol Energy* 77:625–633
65. Chong MN, Jin B, Chow CWK, Saint C (2010) Recent developments in photocatalytic water treatment technology: a review. *Water Res* 44:2997–3027
66. Brillas E, Mur E, Sauleda R, Sanchez L, Peral J, Domenech X, Casado J (1998) Aniline mineralization by AOP's: anodic oxidation, photocatalysis, electro-Fenton and photoelectro-Fenton processes. *Appl Catal B Environ* 16:31–42
67. Serpone N (2006) Is the band gap of pristine TiO<sub>2</sub> narrowed by anion- and cation-doping of titanium dioxide in second-generation photocatalysts? *J Phys Chem B* 110:24287–24293

68. Li FB, Li XZ (2002) The enhancement of photodegradation efficiency using Pt-TiO<sub>2</sub> catalyst. *Chemosphere* 48:1103–1111
69. Ni M, Leung MKH, Leung DYC, Sumathy K (2007) A review and recent developments in photocatalytic water-splitting using TiO<sub>2</sub> for hydrogen production. *Renew Sustain Energy Rev* 11:401–425
70. Sun YF, Pignatello JJ (1995) Evidence for a surface dual hole – radical mechanism in the TiO<sub>2</sub> photocatalytic oxidation of 2,4-dichlorophenoxyacetic acid. *Environ Sci Technol* 29:2065–2072
71. Rabani J, Yamashita K, Ushida K, Stark J, Kira A (1998) Fundamental reactions in illuminated titanium dioxide nanocrystallite layers studied by pulsed laser. *J Phys Chem B* 102:1689–1695
72. Chen YX, Yang SY, Wang K, Lou LP (2005) Role of primary active species and TiO<sub>2</sub> surface characteristic in UV-illuminated photodegradation of acid orange 7. *J Photochem Photobiol A Chem* 172:47–54
73. Turchi CS, Ollis DF (1990) Photocatalytic degradation of organic-water contaminants – mechanisms involving hydroxyl radical attack. *J Catal* 122:178–192
74. Palominos R, Freer J, Mondaca MA, Mansilla HD (2008) Evidence for hole participation during the photocatalytic oxidation of the antibiotic flumequine. *J Photochem Photobiol A Chem* 193:139–145
75. Sakai H, Baba R, Hashimoto K, Fujishima A, Heller A (1995) Local detection of photoelectrochemically produced H<sub>2</sub>O<sub>2</sub> with a wired horseradish-peroxidase microsensor. *J Phys Chem* 99:11896–11900
76. Kikuchi Y, Sunada K, Iyoda T, Hashimoto K, Fujishima A (1997) Photocatalytic bactericidal effect of TiO<sub>2</sub> thin films: Dynamic view of the active oxygen species responsible for the effect. *J Photochem Photobiol A Chem* 106:51–56
77. Ranjit KT, Willner I, Bossmann SH, Braun AM (2001) Lanthanide oxide-doped titanium dioxide photocatalysts: novel photocatalysts for the enhanced degradation of p-chlorophenoxyacetic acid. *Environ Sci Technol* 35:1544–1549
78. Cho M, Chung H, Choi W, Yoon J (2004) Linear correlation between inactivation of *E. coli* and •OH radical concentration in TiO<sub>2</sub> photocatalytic disinfection. *Water Res* 38:1069–1077
79. Rincón AG, Pulgarin C (2004) Effect of pH, inorganic ions, organic matter and H<sub>2</sub>O<sub>2</sub> on *E. coli* K12 photocatalytic inactivation by TiO<sub>2</sub> – implications in solar water disinfection. *Appl Catal B Environ* 51:283–302
80. Wang YB, Hong CS (1999) Effect of hydrogen peroxide, periodate and persulfate on photocatalysis of 2-chlorobiphenyl in aqueous TiO<sub>2</sub> suspensions. *Water Res* 33:2031–2036
81. Rincón AG, Pulgarin C (2003) Photocatalytic inactivation of *E. coli*: effect of (continuous-intermittent) light intensity and of (suspended-fixed) TiO<sub>2</sub> concentration. *Appl Catal B Environ* 44:263–284
82. Kositzki M, Poullos I, Malato S, Caceres J, Campos A (2004) Solar photocatalytic treatment of synthetic municipal wastewater. *Water Res* 38:1147–1154
83. Bahnemann D (2004) Photocatalytic water treatment: solar energy applications. *Sol Energy* 77:445–459
84. Kilic M, Cinar Z (2009) A quantum mechanical approach to TiO<sub>2</sub> photocatalysis. *J Adv Oxidation Technol* 12:37–46
85. Christensen PA, Curtis TP, Egerton TA, Kosa SAM, Tinlin JR (2003) Photoelectrocatalytic and photocatalytic disinfection of *E. coli* suspensions by titanium dioxide. *Appl Catal B Environ* 41:371–386
86. Dunlop PSM, Ciavola M, Rizzo L, Byrne JA (2011) Inactivation and injury assessment of *Escherichia coli* during solar and photocatalytic disinfection in LDPE bags. *Chemosphere* 85:1160–1166
87. Tatlidil I, Sokmen M, Breen C, Clegg F, Buruk CK, Bacaksiz E (2011) Degradation of candida albicans on TiO<sub>2</sub> and Ag-TiO<sub>2</sub> thin films prepared by sol-gel and nanosuspensions. *J Sol-Gel Sci Technol* 60:23–32

88. Melián JAH, Rodríguez JMD, Suárez AV, Rendón ET, do Campo CV, Arana J, Peña JP (2000) The photocatalytic disinfection of urban waste waters. *Chemosphere* 41:323–327
89. Saito T, Iwase T, Horie J, Morioka T (1992) Mode of photocatalytic bactericidal action of powdered semiconductor TiO<sub>2</sub> on *mutans streptococci*. *J Photochem Photobiol B* 14:369–379
90. Kim B, Kim D, Cho D, Cho S (2003) Bactericidal effect of TiO<sub>2</sub> photocatalyst on selected food-borne pathogenic bacteria. *Chemosphere* 52:277–281
91. Watts RJ, Kong SH, Orr MP, Miller GC, Henry BE (1995) Photocatalytic inactivation of coliform bacteria and viruses in secondary waste-water effluent. *Water Res* 29:95–100
92. Cho M, Yoon J (2008) Measurement of •OH radical CT for inactivating *Cryptosporidium parvum* using photo/ferrioxalate and photo/TiO<sub>2</sub> systems. *J Appl Microbiol* 104:759–766
93. Ryu H, Gerrity D, Crittenden JC, Abbaszadegan M (2008) Photocatalytic inactivation of *Cryptosporidium parvum* with TiO<sub>2</sub> and low-pressure ultraviolet irradiation. *Water Res* 42:1523–1530
94. Navalon S, Alvaro M, Garcia H, Escrig D, Costa V (2009) Photocatalytic water disinfection of *Cryptosporidium parvum* and *Giardia lamblia* using a fibrous ceramic TiO<sub>2</sub> photocatalyst. *Water Sci Technol* 59(4):639–645
95. Sökmen M, Degerli S, Aslan A (2008) Photocatalytic disinfection of *Giardia intestinalis* and *Acanthamoeba castellanii* cysts in water. *Exp Parasitol* 119:44–48
96. Giannantonio DJ, Kurth JC, Kurtis KE, Sobczyk PA (2009) Effects of concrete properties and nutrients on fungal colonization and fouling. *Int Biodeterior Biodegrad* 63:252–259
97. Wang H, Wu Y, Xu BQ (2005) Preparation and characterization of nanosized anatase TiO<sub>2</sub> cuboids for photocatalysis. *Appl Catal B Environ* 59:139–146
98. Benabbou AK, Derriche Z, Felix C, Lejeune P, Guillard C (2007) Photocatalytic inactivation of *Escherichia coli* – effect of concentration of TiO<sub>2</sub> and microorganism, nature, and intensity of UV irradiation. *Appl Catal B Environ* 76:257–263
99. Vohra A, Goswami DY, Deshpande DA, Block SS (2006) Enhanced photocatalytic disinfection of indoor air. *Appl Catal B Environ* 64:57–65
100. Ibáñez JA, Litter MI, Pizarro RA (2003) Photocatalytic bactericidal effect of TiO<sub>2</sub> on enterobacter cloacae: comparative study with other Gram(–) bacteria. *J Photochem Photobiol A Chem* 157:81–85
101. Cohen-Yaniv V, Narkis N, Armon R (2008) Photocatalytic inactivation of flavobacterium and *E. coli* in water by a continuous stirred tank reactor (CSTR) fed with suspended/immobilised TiO<sub>2</sub> medium. *Water Sci Technol* 58(1):247–252
102. Suketa N, Sawase T, Kitaura H, Naito M, Baba K, Nakayama K, Wennerberg A, Atsuta M (2005) An antibacterial surface on dental implants, based on the photocatalytic bactericidal effect. *Clin Implant Dent Relat Res* 7:105–111
103. Amezaga-Madrid P, Nevarez-Moorillon GV, Orrantia-Borunda E, Miki-Yoshida M (2002) Photoinduced bactericidal activity against *pseudomonas aeruginosa* by TiO<sub>2</sub> based thin films. *FEMS Microbiol Lett* 211:183–188
104. Amezaga-Madrid P, Silveyra-Morales R, Cordoba-Fierro L, Nevarez-Moorillon GV, Miki-Yoshida M, Orrantia-Borunda E, Solis FJ (2003) TEM evidence of ultrastructural alteration on *pseudomonas aeruginosa* by photocatalytic TiO<sub>2</sub> thin films. *J Photochem Photobiol B Biol* 70:45–50
105. Cheng YW, Chan RCY, Wong PK (2007) Disinfection of *Legionella pneumophila* by photocatalytic oxidation. *Water Res* 41:842–852
106. Chun MJ, Shim E, Kho EH, Park KJ, Jung J, Kim JM, Kim B, Lee KH, Cho DL, Bai DH, Lee SI, Hwang HS, Ohk SH (2007) Surface modification of orthodontic wires with photocatalytic titanium oxide for its antiadherent and antibacterial properties. *Angle Orthod* 77:483–488
107. Song SJ, Kim KS, Kim KH, Li HJ, Cho DL, Kim JB, Park HJ, Shon H, Kim JH (2008) Fabrication of TiO<sub>2</sub> impregnated stainless steel fiber photocatalysts and evaluation of photocatalytic activity. *J Kor Ind Eng Chem* 19:674–679
108. Cho M, Choi Y, Park H, Kim K, Woo GJ, Park J (2007) Titanium dioxide/UV photocatalytic disinfection in fresh carrots. *J Food Prot* 70:97–101

109. Nagame S, Oku T, Kambara M, Konishi K (1989) Antibacterial effect of the powdered semiconductor TiO<sub>2</sub> on the viability of oral microorganisms. *J Dent Res* 68:1696–1697
110. Kim BH, Kim D, Cho DL, Lim SH, Yoo SY, Kook JK, Cho YI, Ohk SH, Ko YM (2007) Sterilization effects of a TiO<sub>2</sub> photocatalytic film against a *streptococcus mutans* culture. *Biotechnol Bioprocess Eng* 12:136–139
111. Dunlop PSM, Sheeran CP, Byrne JA, McMahan MAS, Boyle MA, McGuigan KG (2010) Inactivation of clinically relevant pathogens by photocatalytic coatings. *J Photochem Photobiol A Chem* 216:303–310
112. Guimarães JR, Barretto AS (2003) Photocatalytic inactivation of *Clostridium perfringens* and coliphages in water. *Braz J Chem Eng* 20:403–411
113. Greist HT, Hingorani SK, Kelly K, Goswami DY (2002) Using scanning electron microscopy to visualize photocatalytic mineralization of airborne microorganisms. In: Proceedings of the 9th international conference on indoor air quality and climate, July 2002, Monterey, California., pp 712–717
114. Chen FN, Yang XD, Wu Q (2009) Antifungal capability of TiO<sub>2</sub> coated film on moist wood. *Build Environ* 44:1088–1093
115. Yao Y, Ohko Y, Sekiguchi Y, Fujishima A, Kubota Y (2008) Self-sterilization using silicone catheters coated with Ag and TiO<sub>2</sub> nanocomposite thin film. *J Biomed Mater Res B Appl Biomater* 85B:453–460
116. Lin CY, Li CS (2003) Effectiveness of titanium dioxide photocatalyst filters for controlling bioaerosols. *Aerosol Sci Technol* 37:162–170
117. Sunnotel O, Verdoold R, Dunlop PSM, Snelling WJ, Lowery CJ, Dooley JSG, Moore JE, Byrne JA (2010) Photocatalytic inactivation of *Cryptosporidium parvum* on nanostructured titanium dioxide films. *J Water Health* 8:83–91
118. Lee JH, Kang M, Choung SJ, Ogino K, Miyata S, Kim MS, Park JY, Kim JB (2004) The preparation of TiO<sub>2</sub> nanometer photocatalyst film by a hydrothermal method and its sterilization performance for *Giardia lamblia*. *Water Res* 38:713–719
119. Peller JR, Whitman RL, Griffith S, Harris P, Peller C, Scalzitti J (2007) TiO<sub>2</sub> as a photocatalyst for control of the aquatic invasive alga, cladophora, under natural and artificial light. *J Photochem Photobiol A Chem* 186:212–217
120. Hong JL, Ma H, Otaki M (2005) Controlling algal growth in photo-dependent decolorant sludge by photocatalysis. *J Biosci Bioeng* 99:592–597
121. Linkous CA, Carter GJ, Locuson DB, Ouellette AJ, Slattery DK, Smith LA (2000) Photocatalytic inhibition of algae growth using TiO<sub>2</sub>, WO<sub>3</sub>, and cocatalyst modifications. *Environ Sci Technol* 34:4754–4758
122. Kim SC, Lee DK (2005) Inactivation of algal blooms in eutrophic water of drinking water supplies with the photocatalysis of TiO<sub>2</sub> thin film on hollow glass beads. *Water Sci Technol* 52(9):145–152
123. Guillard C, Bui TH, Felix C, Moules V, Lina B, Lejeune P (2008) Microbiological disinfection of water and air by photocatalysis. *C R Chim* 11:107–113
124. Cho M, Chung HM, Choi WY, Yoon JY (2005) Different inactivation behaviors of MS-2 phage and *Escherichia coli* in TiO<sub>2</sub> photocatalytic disinfection. *Appl Environ Microbiol* 71:270–275
125. Yu KP, Lee GWM, Lin ZY, Huang CP (2008) Removal of bioaerosols by the combination of a photocatalytic filter and negative air ions. *J Aerosol Sci* 39:377–392
126. Lin ZX, Li ZH, Wang XX, Fu XZ, Yang GQ, Lin HX, Meng C (2006) Inactivation efficiency of TiO<sub>2</sub> on H1N1 influenza virus. *Chem J Chin Univ* 27:721–725
127. Kozlova EA, Safatov AS, Kiselev SA, Marchenko VY, Sergeev AA, Skarnovich MO, Emelyanova EK, Smetannikova MA, Buryak GA, Vorontsov AV (2010) Inactivation and mineralization of aerosol deposited model pathogenic microorganisms over TiO<sub>2</sub> and Pt/TiO<sub>2</sub>. *Environ Sci Technol* 44:5121–5126

128. Han W, Zhang PH, Cao WC, Yang DL, Taira S, Okamoto Y, Ara JI, Yan XY (2004) The inactivation effect of photocatalytic titanium apatite filter on SARS virus. *Prog Biochem Biophys* 31:982–985
129. Khan U, Benabderrazik N, Bourdelais AJ, Baden DG, Rein K, Gardinali PR, Arroyo L, O'Shea KE (2010) UV and solar TiO<sub>2</sub> photocatalysis of brevetoxins (PbTx<sub>s</sub>). *Toxicon* 55:1008–1016
130. Shephard GS, Stockenstrom S, De Villiers D, Engelbrecht WJ, Sydenham EW, Wessels GFS (1998) Photocatalytic degradation of cyanobacterial microcystin toxins in water. *Toxicon* 36:1895–1901
131. Liu I, Lawton LA, Bahnmann DW, Robertson PKJ (2005) The photocatalytic destruction of the cyanotoxin, nodularin using TiO<sub>2</sub>. *Appl Catal B Environ* 60:245–252
132. Theron J, Walker JA, Cloete TE (2008) Nanotechnology and water treatment: applications and emerging opportunities. *Crit Rev Microbiol* 34:43–69
133. Likodimos V, Dionysiou DD, Falaras P (2010) Clean water: water detoxification using innovative photocatalysts. *Rev Environ Sci Bio-Technol* 9:87–94
134. Zhang DQ, Li GS, Yu JC (2010) Inorganic materials for photocatalytic water disinfection. *J Mater Chem* 20:4529–4536