ERRATUM TO:

Flow Cytometry Protocols

3rd Edition

Edited by

Teresa S. Hawley

Flow Cytometry Core Facility, George Washington University Medical Center, Washington, DC, USA

Robert G. Hawley

Department of Anatomy and Regenerative Biology, George Washington University Medical Center, Washington, DC, USA

Page V: The text from line 61 to line 63 should read: "Great strides have been made toward quantitative fluorescence measurement, bead-based multiplexed analysis, semiautomated high-throughput flow cytometry, and fluorescence measurement, resonance energy transfer analysis of protein interactions."

Page XI: Joseph D. Tario's name should read "Joseph D. Tario, Jr."

Page 87: The text of the sentence on line 76 should read, "The intensity of FL3 fluorescence thus provides the "address" analyte."

page 91: On line 213, the sample should be changed from '1:2 to 1:1'

Teresa S. Hawley and Robert G. Hawley (eds.), *Flow Cytometry Protocols*, Methods in Molecular Biology, vol. 699,
DOI 10.1007/978-1-61737-950-5,

page 92: Method 13 should read: “Determine the median intensity of PE fluorescence at each dilution of the sample for each of microsphere population. The lowest dilution should exhibit the greatest signal, which decreases as the analyte is diluted. Plot the FL2 intensity versus dilution in log scale (Fig. 3b) for each sample and each microsphere population. The lowest dilution is on the right of the *x*-axis; the highest dilution is on the left. The dilution value with detectable signal (usually two standard deviations above the signal of the blank microspheres in wells 1–3 then is the titer of the sample for the analyte defined by the microsphere population.

page 158: Note 42 on beginning on line 1081 should read: “Choice of optimized labeling conditions for tracking dye(s) should have already established that stained cells will be on scale, without events accumulating in the highest channel. This condition should be established when compensation is set to 0% and voltages are adjusted to place unstained control cells in the first decade, but sufficiently above the left axis such that events do not accumulate in the lowest channel. In addition, it is critical to recognize that cells labeled with tracking dyes can be extremely bright, requiring substantially decreased detector voltage settings compared with those used for the detection of immunofluorescence. If the tracking dye signal in a secondary channel (set to a higher detector voltage) is greater than its intensity in the primary channel, it will be impossible to compensate properly for tracking dye overlap in the secondary channel (41). In this case, staining with reduced concentrations of tracking dye(s) may be required.

page 280: In ‘Table 1’ superscript ‘6’ should read ‘f’

Page 317: Replace ‘pipet’ with ‘pippet’ throughout Chapter 15

Page 347: The sentence beginning on line 365 should read: “It is important to include negative isotype controls for the selected specific Abs because there is greater nonspecific binding of Abs to MPs than intact cells and the Ab may not be removed if washing is omitted.”

Page 353: Note 5 should read: “These abs are not absolutely cell-type specific because, for example, CD4 is detected on monocytes and CD11b on granulocytes.”

Page 485: Change ‘target labeling Cy3, Cy5, Leader 423-426’ to ‘target labeling: Cy3, Cy5’