



Quinolines and Quinolones as Antibacterial, Antifungal, Anti-virulence, Antiviral and Anti-parasitic Agents

Lidija Senerovic, Dejan Opsenica, Ivana Moric, Ivana Aleksic, Marta Spasić, and Branka Vasiljevic

Abstract

Infective diseases have become health threat of a global proportion due to appearance and spread of microorganisms resistant to majority of therapeutics currently used for their treatment. Therefore, there is a constant need for development of new antimicrobial agents, as well as novel therapeutic strategies. Quinolines and quinolones, isolated from plants, animals, and microorganisms, have demonstrated numerous biological activities such as antimicrobial, insecticidal, anti-inflammatory, antiplatelet, and antitumor. For more than two centuries quinoline/quinolone moiety has been used as a scaffold for drug

development and even today it represents an inexhaustible inspiration for design and development of novel semi-synthetic or synthetic agents exhibiting broad spectrum of bioactivities. The structural diversity of synthesized compounds provides high and selective activity attained through different mechanisms of action, as well as low toxicity on human cells. This review describes quinoline and quinolone derivatives with antibacterial, antifungal, anti-virulent, antiviral, and anti-parasitic activities with the focus on the last 10 years literature.

Keywords

Antibiotics · Antifungals · Anti-parasitics · Antivirals · Anti-virulence activity · Quinoline/quinolone derivatives

L. Senerovic (✉), I. Moric, I. Aleksic, and B. Vasiljevic
Institute of Molecular Genetics and Genetic Engineering,
University of Belgrade, Belgrade, Serbia
e-mail: seneroviclidija@imgge.bg.ac.rs;
ivanamoric@imgge.bg.ac.rs;
ivana_aleksic@imgge.bg.ac.rs; brankav@imgge.bg.ac.rs

D. Opsenica
Institute of Chemistry, Technology and Metallurgy,
University of Belgrade, Belgrade, Serbia

Center of excellence in Environmental Chemistry and
Engineering, ICTM - University of Belgrade,
Belgrade, Serbia
e-mail: dopsen@chem.bg.ac.rs

M. Spasić
Faculty of Chemistry, University of Belgrade,
Belgrade, Serbia

1 Introduction

Antimicrobial drugs, structurally diverse molecules, can be natural products, semi-synthetic derivatives of natural compounds, or chemically synthesized compounds. The development of antimicrobials in general, first of all antibiotics, but also antivirals and antimalarials, revolutionized medicine in many ways, and as such is one of the greatest successes of modern

medicine. Unfortunately, time with these drugs is rapidly running out. Occurrence and global spread of resistance of bacteria, fungi, viruses, and protozoan parasites to available antimicrobial medicines threaten to send humanity back to pre-antimicrobial era. Therefore, there is an urgent need not just to develop novel antimicrobials but also to introduce into practice novel therapeutic options to fight against both drug-sensitive and drug-resistant pathogens.

A promising alternative to the classic antibiotic approach has recently been established and is known as anti-virulence therapy. Instead of targeting microbial viability, this alternative strategy aims to target pathogens' virulence machinery required to cause host damage and disease. Microbial virulence machinery includes plethora of virulence factors, which are diverse in structure, function, and localization. One of the most important virulent characteristics of both bacteria and fungi is their ability to form biofilms. Biofilms are multicellular communities enclosed in self-synthesized polymeric matrices attached to biotic or abiotic surfaces (Hall-Stoodley et al. 2004; Costa-Orlandi et al. 2017). According to the National Institutes of Health of the United States, more than 75% of microbial infections that occur in the human body are promoted by the formation and persistence of biofilms (Miquel et al. 2016). Biofilm confers an extreme capacity for persistence against phagocytosis, oxidative stress, nutrient/oxygen restriction, metabolic waste accumulation, interspecies competition, and most importantly, conventional antimicrobial agents (Moradali et al. 2017). Quorum sensing (QS), a cell-to-cell communication system, is a global regulatory system of virulence factor production and biofilm formation in both bacteria and fungi (Albuquerque and Casadevall 2012; Defoirdt 2018), with no homologous components in humans, thus its inhibition is considered the most attractive strategy for the development of anti-virulence agents.

Quinine (Fig. 1), quinoline alkaloid isolated from the bark of the *Cinchona* tree in 1820, used in the treatment of malaria played a historical role in the development of quinoline alkaloids as therapeutics. These quinoline based compounds have

been isolated and identified from natural sources (plants, animals, and microorganisms), and many studies have documented their antitumor, antimalarial, antibacterial, antifungal, antiviral, anti-parasitic and insecticidal, anti-inflammatory, antiplatelet and other activities (Shang et al. 2018). Quinoline and 4-quinolone (Fig. 1) moieties were used as a scaffold for drug development for more than two centuries (Heeb et al. 2011). The most successful drug based on quinoline scaffold is chloroquine (Fig. 1), which was specifically developed as antimalarial agent. Until today, numerous quinoline-based compounds and drugs were developed as antimalarial agents, designed to target all stages of parasite life-cycle.

In fact quinolines still serve as inexhaustible models for design and development of new semi-synthetic or synthetic quinoline/quinolone antimicrobial agents, which are the focus of this review article.

2 Antibacterial Activity

Important group of antibacterial agents are synthetic antibiotics with 4-quinolone as core structure, which are used in the treatment of urinary tract and respiratory infections (Anderson et al. 2012). Today antibacterial 4-quinolones in clinical use include nalidixic acid, that is introduced to medical practice in 1964 (Bisacchi 2015), followed by ciprofloxacin, levofloxacin, norfloxacin, besifloxacin, and moxifloxacin (Fig. 2). Majority of them belong to fluoroquinolone chemotype. The quinolone antibiotics are very potent towards a wide range of Gram negative bacteria, with minimal inhibitory concentrations (MICs) in the ng/ml range, and are reasonably active against many Gram positive bacteria (MICs in the mg/ml range) (Anderson et al. 2012). Their antibacterial activity is based on inhibition of DNA replication through inhibition of DNA gyrase and topoisomerase IV activities to varying extents depending on the pathogen. In order to maintain antibacterial activity, positions C(3) (unsubstituted carboxyl group) and C(4) (keto group) in 4-quinolone ring should not be altered (Gualerzi et al. 2013).

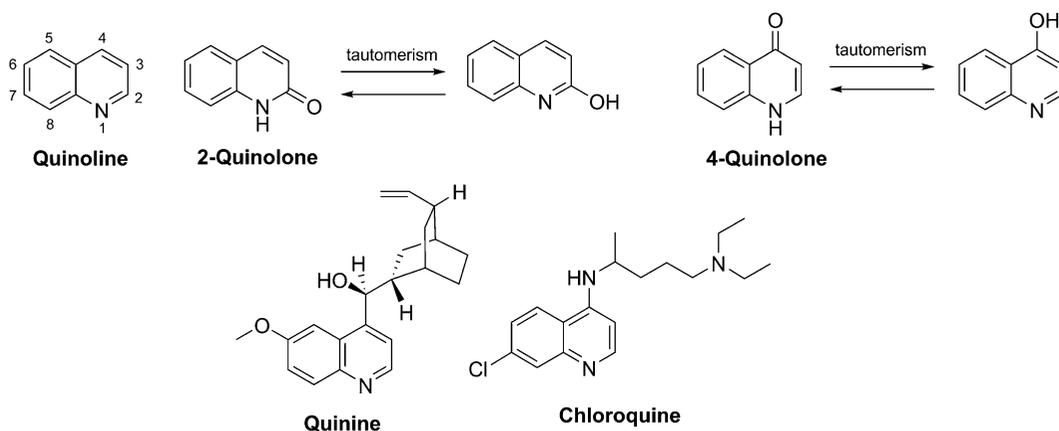


Fig. 1 Quinoline and quinolone scaffolds and their best-known drugs

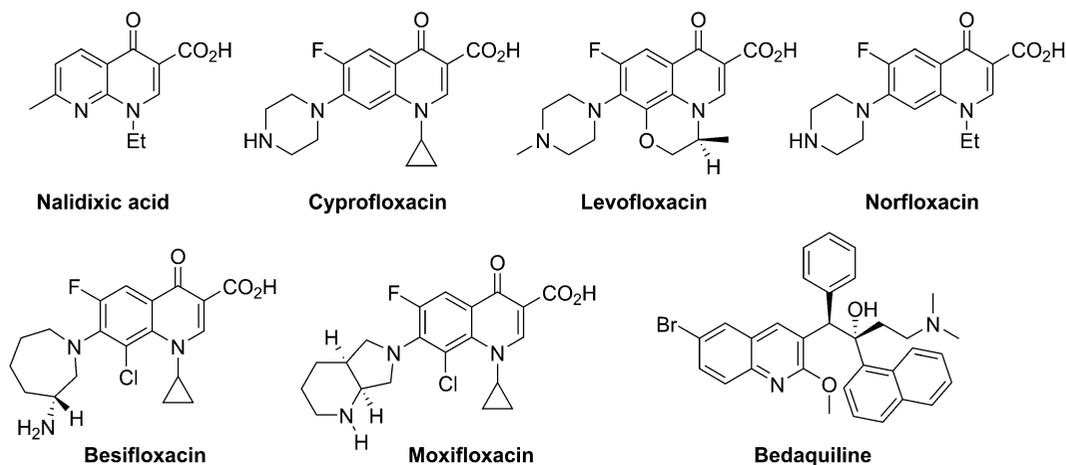


Fig. 2 Quinoline and quinolone antibiotics

After more than 40 years, in 2012, a new antibiotic with quinoline as a core pharmacophore developed to fight multidrug-resistant (MDR) mycobacteria, was granted accelerated approval by the United States Food and Drug Administration, process applicable only to the therapeutics that should fill an unmet medical needs (World Health Organization 2013). Unlike other members of quinolone antibiotics, new antibiotic bedaquiline (Fig. 2) imposes its antimycobacterial activity (MIC 0.06 $\mu\text{g/ml}$), through interaction with proton pump of the ATP synthase of *Mycobacterium tuberculosis* (Andries et al. 2005).

Using building-block approach in quest for novel antibacterial agents Dolan and colleagues

(Dolan et al. 2016) combined structural moieties, such as functional groups bearing fluorine atoms, quinoline bicycles, saturated N-heterocycles, and thioureas, all of them recognized as motifs in antimicrobial agents, and synthesized a series of thiourea-containing compounds. Derivative **1** (Fig. 3) was the most active compound with MIC₉₀ values of 7.90–10.52 μM , 10.52–15.78 μM , and 17.74 μM for *Escherichia coli*, *Staphylococcus aureus*, and methicillin resistant *S. aureus* (MRSA), respectively, but without any effect on *Pseudomonas aeruginosa*. Its antibacterial activities were in the line with MIC₉₀ values obtained for vancomycin, well-known antibiotic of the last resort often used to

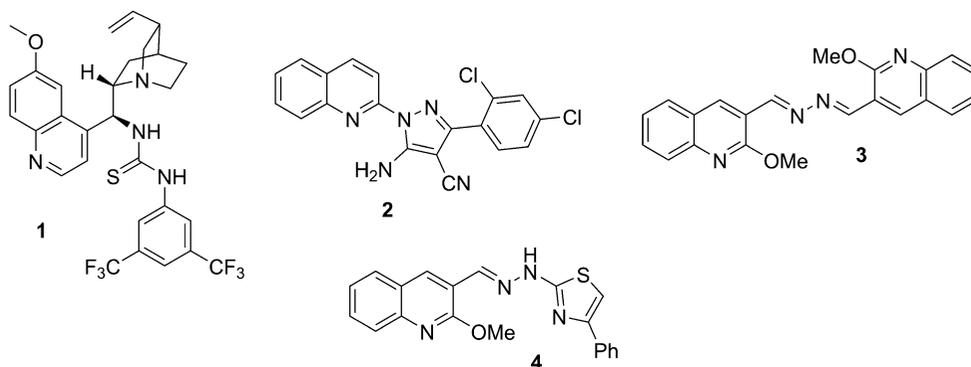


Fig. 3 Quinoline derivatives with antibacterial activity

treat drug-resistant infections. The derivative **1** was found to be non-toxic to *Galleria mellonella* larvae at concentrations of up to 1000 $\mu\text{g/ml}$. Hence, the quinoline-thiourea structure, as found in compound **1**, has potential as a new class of non-toxic, anti-MRSA agent.

Synergistic effect of quinoline and pyrazole derivatives (also known for numerous bioactivities including antituberculosis, antiviral, and anti-inflammatory ones) on antimicrobial activity of three diverse series was assessed (El Shehry et al. 2018). Majority of derivatives exhibited antibacterial and antifungal activities, and the most promising derivative proved to be compound **2** (Fig. 3). Its MICs ranged from 0.12 to 0.98 $\mu\text{g/ml}$ for *Shigella flexneri*, *Klebsiella pneumoniae*, *Staphylococcus epidermidis*, and *Proteus vulgaris*, were in the range of ampicillin and gentamycin MICs, thus demonstrating that quinoline derivative bearing pyrazole motifs are interesting scaffolds for development of antimicrobial agents.

Similarly, antibacterial and antifungal activities of a novel quinoline series of Schiff bases and hydrazide derivatives containing moiety, synthesized *via* condensation of aromatic amines or hydrazines with 2-substituted quinoline-3-carbaldehydes was assessed (Hamama et al. 2018). Antimicrobial activities of quinoline derivatives **3** and **4** (Fig. 3; MIC values 30.6–93.7 $\mu\text{g/ml}$ and 62.5–125 $\mu\text{g/ml}$, respectively) were even better than activity of ampicillin (MIC values 125–187.5 $\mu\text{g/ml}$) against *E. coli*, *P. aeruginosa*, *S. aureus* and *B. subtilis*. The

same derivative exhibited similar antifungal activity (MIC values 4.5–7.8 $\mu\text{g/ml}$ and 15.6–23 $\mu\text{g/ml}$) as clotrimazole (5.6–5.8 $\mu\text{g/ml}$). The activities of quinoline derivatives **3** and **4** are most likely due to their combination with another quinoline and thiazole moieties, respectively.

Several studies were focused on synthesis of different halogenated quinolines (HQ) and the assessment of bioactivities of the obtained derivatives, namely antibacterial and biofilm eradicating ability. In the first study, the HQ scaffolds or esters of halogenated 8-hydroxyquinoline derivatives were synthesized (Abouelhasan et al. 2014). Five brominated quinolines demonstrated more potent antibacterial activity (MICs 0.2–1.56 μM) against *S. aureus* and *S. epidermidis* strains in comparison to nitroxoline (MIC = 12.5–25 μM). In biofilm dispersion assay against MRSA isolate seven quinoline derivatives were very efficient (concentration at which 50% of pre-formed biofilms is disrupted; $\text{BDIC}_{50} \leq 5 \mu\text{M}$), with derivative **5** as the most potent (Fig. 4; $\text{BDIC}_{50} = 2.06 \mu\text{M}$). HQ compounds were less potent against *S. epidermidis*, although derivatives **6** and **7** exhibited excellent biofilm eradication potential (Fig. 4; $\text{BDIC}_{50} = 3.26 \mu\text{M}$ and $\text{BDIC}_{50} = 5.56 \mu\text{M}$, respectively). Nitroxoline BDIC_{50} values were 10.5 μM and 14.2 μM for MRSA and *S. epidermidis* biofilms, respectively. In the abovementioned study it has been revealed that the C(2)-position of the HQs is of crucial importance for their antibacterial profile against different bacterial species and their potential to

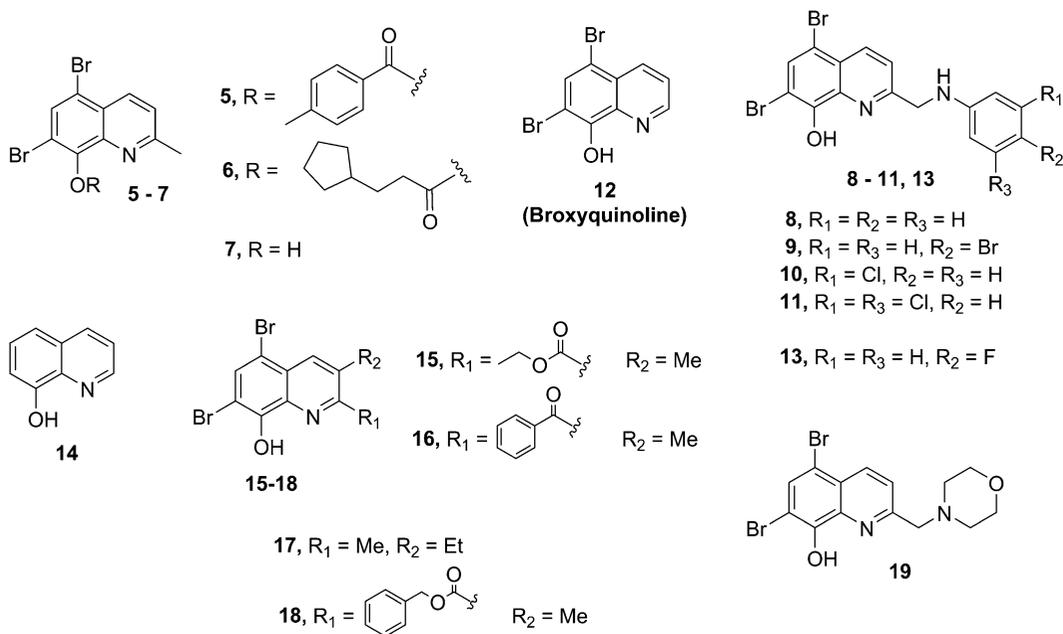


Fig. 4 Halogenated quinoline derivatives with antibacterial and antibiofilm activities

eradicate preformed biofilms. Therefore a series of C(2)-substituted analogues of the HQs were synthesized through reductive amination and tested for bioactivity (Basak et al. 2015). These derivatives demonstrated equipotent or slightly reduced antibacterial activities as parent compound **7** (Fig. 4) against clinical isolate MRSA, while products **8** (aniline derived) and **9** (4-bromoaniline derived) were the most potent analogues against methicillin-resistant *S. epidermidis* (MRSE), fivefold more potent than vancomycin (MIC = 0.78 μM) (Fig. 4). Derivatives **8** through **11** proved to be three- to six-fold more potent (MIC = 0.39–0.78 μM) against vancomycin-resistant *Enterococcus faecium* (VRE) than **7** (MIC = 2.35 μM) or linezolid (MIC = 3.13 μM). The best potency for MRSA biofilm eradication activities demonstrated **9** with minimum biofilm eradication concentration (MBEC) of 125 μM , being at least 16-fold more potent than vancomycin, daptamycin, or linezolid antibiotics (MBECs > 2000 μM). Derivatives **8** (MBEC = 3.0 μM) and **10** (MBEC = 5.9 μM) were the most potent MRSE biofilm eradicators in this series, while **7** and **8** through **11**

demonstrated potent biofilm eradication activities against VRE biofilms (MBEC = 1.0–1.5 μM) and as such were equipotent to linezolid, which is used to treat VRE infections. Haemolytic activity of the compounds has varied between 21.3% and less than 4%, as for **8** and **9**, indicating that analogues do not eradicate biofilms through the destruction of bacterial membranes, but through other mechanism. In addition, the authors have concluded that the HQs antibacterial activity is realised through a metal(II)-dependent mode of action possibly through the targeting of a metalloprotein critical to bacterial biofilm viability (Basak et al. 2015).

In the following study Basak and colleagues (Basak et al. 2016) derivatised broxyquinoline (**12**, Fig. 4) through multi-step synthetic routes to achieve highly diverse HQ analogues alkylated and aminated at the C(2)-position. Among 39 synthesized derivatives, **13** (Fig. 4) emerged as a highly potent anti-planktonic compound and eradicating agent against MRSA (MIC = 0.39 μM , MBEC = 7.8–93.8 μM), MRSE (MIC = 0.39 μM , MBEC = 5.9 μM) and VRE (MIC = 0.78 μM , MBEC = 1 μM) biofilms, when compared to vancomycin (MRSA: MIC = 0.59 μM ,

MBEC > 2000 μM ; MRSE: MIC = 0.78 μM , MBEC > 2000 μM ; VRE: MIC > 100 μM , MBEC = 150 μM). With haemolytic activity < 5% and low cytotoxicity, this compound certainly represent a promising lead for further development of useful treatments against persistent infection caused by Gram positive bacteria (Basak et al. 2016).

Abouelhasan and colleagues (Abouelhasan et al. 2015) tested whether phytochemicals, typically considered as a safe, could potentiate antibacterial activity of halogenated derivatives of 8-hydroxyquinoline (**14**, Fig. 4). They have shown that gallic acid, which itself does not act as antibacterial agent, potentiate several derivatives from authors' libraries described in previous studies against different *S. aureus* strains, by lowering their MICs at least fourfold and, in some cases, up to 11,800-fold. On the other hand, gallic acid in combination with antibiotics of different classes, including quinolone, has not potentiated their antibacterial activity, thus indicating that antibacterial mechanisms of the HQs differs from those of conventional antibiotics. It was also demonstrated that gallic acid in combination with compound **7** potentiate MRSA biofilm eradication fourfold.

For further development of 8-hydroxyquinolines derivatives microwave-enhanced Friedländer synthesis protocol was used in order to synthesize the HQ compounds with C(2)- or C(2)- and C(3)- substituted positions (Garrison et al. 2017). Several derivatives exhibited significant antibacterial activity. Compounds **15** and **16** (Fig. 4) with MICs against MRSA isolate of 0.39 μM and 0.59 μM , respectively, were similar to vancomycin (MIC = 0.59 μM) or better than daptamycin (MIC = 4.69 μM) and linezolid (MIC = 3.13 μM). Compounds **15**, **17**, and **18** exhibited MICs of 0.10 μM , 0.15 μM , and 0.10 μM , respectively, against MRSE, and as such were more efficient than vancomycin (MIC = 0.78 μM), daptamycin (MIC = 12.5 μM) and linezolid (MIC = 3.13 μM). Among all, derivative **18** demonstrated the highest potency against VRE (MIC = 0.3 μM). When tested for eradication potency, compound

16 proved to be one of the most potent biofilm eradicators ever reported against both MRSA (MBEC = 3.9–23.5 μM) and MRSE (MBEC = 1.0 μM), while compounds **15** and **18** exhibited promising biofilm eradication activities against MRSA (MBEC = 31.3 μM) and VRE (MBEC = 1.5 μM), respectively. Since negligible haemolytic activity was observed for those most promising derivatives (< 8% at 200 μM) it seems that the HQs could be a promising class of compounds capable of treating biofilm associated infections.

These studies demonstrated great potentials of numerous HQ derivatives as antibacterial agents and biofilm eradicators, so further investigations have been focused on development of the HQ analogues with improved water solubility while maintaining potent biofilm eradication properties against major human pathogens (Basak et al. 2018; Huigens 2018). Conducting diverse synthetic modification at the C(2)-position of the HQ scaffold in order to enhanced water solubility, a new compound that had lower ClogP value (3.44) than parent compound **7**; (ClogP = 4.19) was developed. This derivative, with morpholine moiety at the C(2)-position **19** (Fig. 4), was the most effective eradicator of MRSE biofilm (MBEC = 2.35 μM) but showed no activity on MRSA biofilms. Therefore, although HQ could play a critical role in the development of next-generation antibacterial therapeutics, at the moment it is still work in progress.

Massoud and colleagues (Massoud et al. 2013) described antibacterial activity of six new Ag (I) compounds with quinoline-derived ligands tested against 15 different MDR bacteria isolated from diabetic foot ulcers and compared them to antibacterial activity of silver sulfadiazine used clinically to prevent infections in burns and wounds. Compound $[\text{Ag}(\text{8-nitroquinoline})_2] \text{NO}_3 \times \text{H}_2\text{O}$ showed activity similar to topical antibiotic against clinical isolates, being active against all strains and having slightly better average silver efficiency than silver sulfadiazine (5 vs. 6 $\mu\text{g Ag/ml}$).

Promising candidates for development of new drug are dimeric molecules since they exhibit

some unique properties in comparison to corresponding monomer, such as enhanced biological activity. In the last three decades, numerous quinoline and quinolone dimers were assessed for their biological activities, including antimicrobial, and quite recently very detailed review article has been published on the subject (Chu et al. 2019); hence, quinoline/quinolone dimeric molecules are not reviewed here.

It is worth noting that bacterial responses to antibiotics (not just 4-quinolone) are concentration-dependent. At high concentrations, antibiotics exhibit their antimicrobial activities on susceptible cells, but at subinhibitory concentrations they can induce diverse biological responses in bacteria. At these non-lethal concentrations, bacteria may perceive antibiotics as extracellular signals and thus trigger different cellular responses, which may include an altered antibiotic resistance/tolerance profile (Bernier and Surette 2013). Bacterial responses to subinhibitory concentrations of antibiotics and mechanisms of their responses vary depending on antibiotic and species, thus we advise readers for literature search on this topic.

3 Antifungal Activity

Fungal infections have become an everyday problem, but also a serious threat to human health due to the development of resistant strains causing weak and unsatisfactory therapeutic response to known antifungals. Although the collection of antifungal drugs is broad, the most commonly used agents have major drawbacks such as side effects and high level of toxicity. Together with the emerging resistance, these drawbacks restrict the number of medicines which can be used to treat such infections. Thus, there is a clear need for development of novel more effective antifungal agents with a broad-spectrum activity, better pharmacokinetic profile and low toxicity. As eukaryotic organisms, fungi share numerous conserved pathways with their human hosts, therefore only a few drug targets can be exploited to selectively kill these pathogens. Enzymes

involved in the synthesis of cell wall polysaccharides are one of the most popular targets for development of antifungal drugs.

A quinoline scaffold can be found in many classes of biologically active compounds which are used as antifungals (Musiol et al. 2010). The unmodified quinoline exhibits relatively high activity against some fungal strains at nontoxic concentrations, which is a clear advantage in the context of designing of novel antifungal drugs. The fungistatic activity of 8-hydroxyquinoline (14, Fig. 4) and its metal complexes has been known since the early 1920s and these compounds are still broadly used in healthcare. Relatively simple quinoline modifications have been widely investigated in order to obtain better antifungal activity and some of these compounds are still in use. Previous efforts in the development of quinoline-based antifungals have been extensively reviewed earlier (Musiol et al. 2010), thus here we focus on the achievements from last 10 years.

A series of small HQ molecules exhibiting a potent antifungal activity against *Candida albicans* and *Cryptococcus neoformans* through intracellular mode of action have been synthesized (Zuo et al. 2016). HQ analogues with bromine and chlorine halogens (7, 20) (Figs. 4 and 5) inhibited *C. albicans* growth with MIC of 100 nM which is four to eight times more potent than the best-known antifungal agents amphotericin B and itraconazole, while several analogues inhibited *C. neoformans* at MICs of 50–780 nM. Importantly, the HQ analogues could eradicate mature *C. albicans* and *C. neoformans* biofilms with MBEC of 6.25–62.5 μ M. The same HQ analogues also showed a range of antibacterial activities inhibiting their planktonic and biofilm forms, suggesting that these compounds could be a promising scaffold for the development of therapeutics against mixed-species and/or biofilm-associated infections. The biological activity of the HQs depended on the nature of the groups attached to the C(5)- and C(7)-positions of quinoline ring. The presence of chlorine, bromine or iodine atoms in the C(5)- or C(7)- positions of the

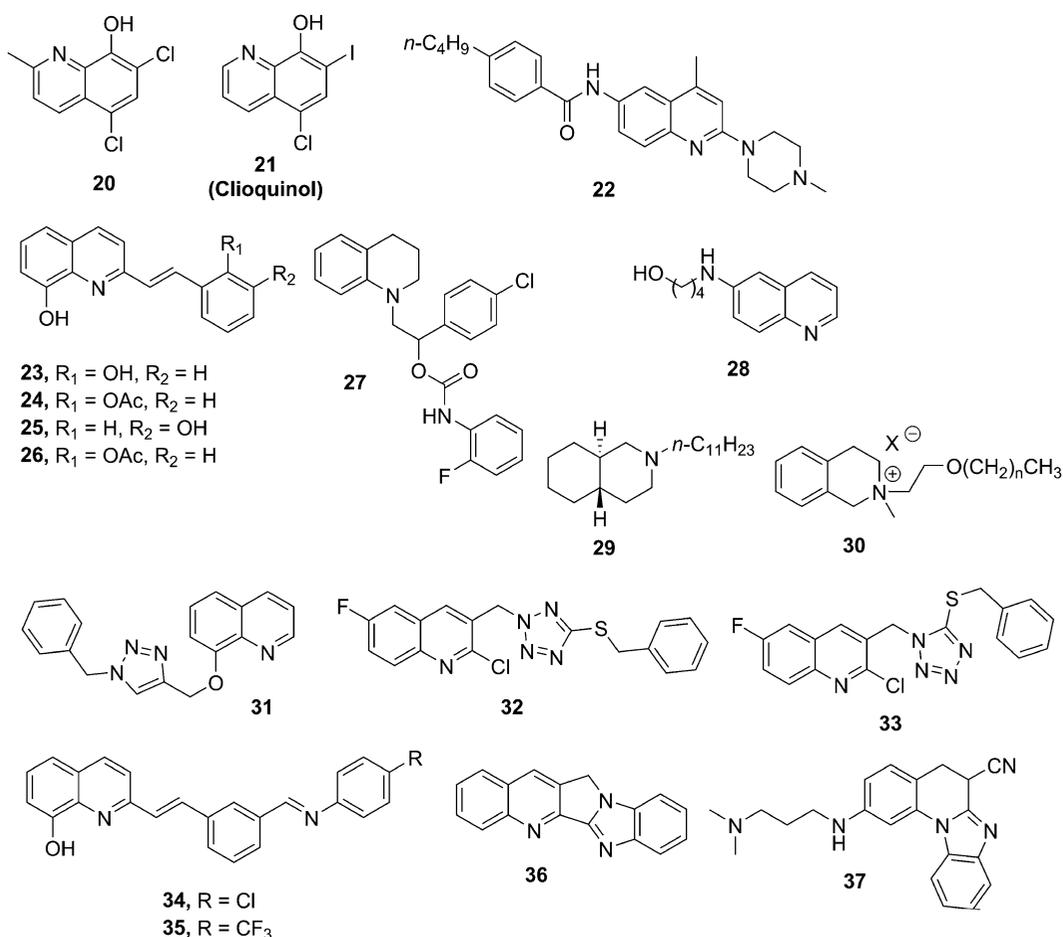


Fig. 5 Quinoline derivatives with antifungal activity

compounds was related to strong antifungal activity, which was not observed with the fluorinated compounds. Although the observed antifungal activity can be attributed to the presence of halogens, those substituents can also cause higher toxicity of the compounds. Bromoquinol (broxyquinoline; **12**, Fig. 4) is identified as one of the most potent compounds with antifungal activity during screening of 40,000 drug-like molecules from diverse chemical compound libraries against *Aspergillus fumigatus* (Ben Yaakov et al. 2017). Its activity was blocked by iron, copper or zinc supplementation, which suggested that it interferes with the utilization of these metals. It was demonstrated that bromoquinol induces oxidative stress and apoptosis in *A. fumigatus*. Bromoquinol significantly

reduced mortality rates of *Galleria mellonella* infected with *A. fumigatus*, but was ineffective in a murine model of infection.

Clioquinol (**21**, Fig. 5) is a dihalogenated 8-hydroxyquinoline and antiseptic drug effective against multidrug resistant *Candida* (Pippi et al. 2018). It blocks hyphal development thus preventing biofilm formation. Clioquinol also reduces the metabolic activity of sessile *Candida* but the susceptibility was lower compared to planktonic cells (0.031–0.5 µg/ml required to inhibit 50% planktonic cells and 4–16 µg/ml to inhibit 50% preformed biofilms). Clioquinol, as well as other 8-hydroxyquinoline derivatives such as 8-hydroxy-5-quinolinesulfonic acid and 8-hydroxy-7-iodo-5-quinolinesulfonic acid effectively inhibited growth of *Candida* spp.,

Microsporium spp. and *Trichophyton* spp. (Pippi et al. 2017) with MIC values ranging from 0.031–2 µg/ml, 1–512 µg/ml, and 2–1024 µg/ml, respectively.

Most of the known quinoline drugs have a side chain on the C(4)- or C(8)-position of the quinoline scaffold. However, other positions also provide opportunities for the design of novel bioactive compounds. Two subseries of 2,6-disubstituted quinolines, consisting of 6-amide and 6-urea derivatives exhibit fungicidal activity against *C. albicans* with minimal fungicidal concentration (MFC) values lower than 15 µM (Delattin et al. 2012). The 6-amide derivatives displayed the highest fungicidal activity against *C. albicans* and a bit lower activity against *C. glabrata* (MFC < 50 µM) (**22** as the most potent, Fig. 5). Some of the 6-amide derivatives and the 6-urea derivatives could also eradicate *C. albicans* biofilms by inducing accumulation of endogenous reactive oxygen species.

Styrylquinolines (**23–26**, Fig. 5) are a novel group of quinoline drugs that have p53-independent antiproliferative activity and antiviral properties. They were also found to have antifungal activity and to decrease the activity of ABC multidrug transporters in *C. albicans*. They also show synergistic activity with first-line drug fluconazole (Szczepaniak et al. 2017).

Novel phenylcarbamate derivatives with tetrahydroquinoline were designed as inhibitors of HIV-1 reverse transcriptase (RT) also showed antifungal activity against *C. albicans* and *A. niger* (Chander et al. 2016). The most active derivative (**27**, Fig. 5) displayed antifungal activity against *C. albicans* almost comparable to fluconazole with MIC value of 8 µg/ml.

The 7-chloroquinoline moiety was extensively studied mainly because of its antimalarial properties. Duval and co-workers synthesized 7-chloroquinolin-4-yl arylhydrazone derivatives and addressed their antifungal activity against several *Candida* species and two yeasts species of *Rhodotorula* (Duval et al. 2011). The active derivatives showed MIC and MFC values in the range of 25 µg/ml and 50 µg/ml, which was the activity comparable with the fluconazole. Four of these hydrazones potently inhibited enzymatic

processes in *C. albicans* at sub-antifungal concentrations, showing enzymatic repression of phospholipase and aspartyl proteases, which are the most frequent enzymes produced by *C. albicans* (de Azambuja Carvalho et al. 2016). Notably, the compounds exhibited low cytotoxicity against mouse fibroblasts (NIH/3 T3 cell line) at sub-antifungal concentrations.

On the way to N-functionalized 3-, 5-, 6- and 8-aminoquinolines, were obtained and their antifungal activity was assessed against *C. albicans*, *Rhodotorula bogoriensis* and *A. flavus* (Vandekerckhove et al. 2015). Several compounds displayed antifungal activity against all three microorganisms tested, while derivative **28** (Fig. 5) was selected as the most potent one, exhibiting activity toward the *A. flavus* strain comparable to amphotericin B.

Evaluation of antifungal activity of *N*-alkyl tetra and decahydroisoquinolines showed that the activity of these compounds depends on the length of the alkyl chain, with an optimum of about 10–12 carbon atoms, whereas longer or shorter alkyl chains lead to a decrease or complete loss of activity (Krauss et al. 2014). The *trans*-decahydroisoquinolines showed high antifungal activity, comparable to the reference antifungal drug clotrimazole with MIC values against *C. glabrata* between 2.4 and 25 µg/ml. The decahydroisoquinoline **29** (Fig. 5) was found to inhibit the enzyme D14-reductase in *C. glabrata*, while an additional inhibition of the downstream enzyme D8,7-isomerase was not excluded. The same group of researchers synthesized a series of (±)-*trans*-*N*-alkylperhydroquinolines, which also showed high antifungal activity with MIC values against *C. glabrata* between 5 and 50 µg/ml (Krauss et al. 2015). The activity of the most potent derivative was comparable to drug clotrimazole. The maximum of activity was found with the derivatives having C10 or C12 alkyl chains, while shorter alkyl chains led to a decrease in activity, as already found for other *N*-alkyl heterocycles. In comparison to *N*-alkyl perhydroisoquinolines, perhydroquinoline compounds showed similar antifungal activity, but higher cytotoxicity against a human cell line. Both perhydroquinolines and perhydroisoquinolines

targeted the same enzyme in ergosterol biosynthesis ($\Delta^{8,7}$ -isomerase), but the latter chemotype showed higher selectivity.

The lipid-like choline and colamine analogues based on 1,2,3,4-tetrahydro(iso)quinoline have been developed as a part of compound libraries of lipid-like systems that combine two fragments: hydrophilic – 1,2,3,4-tetrahydro(iso)quinolinium pharmacophore system, and lipophilic – long chain alkyl substituent (**30**, Fig. 5), that helps the molecule in its passive transport across lipophilic barriers and plasma membranes *in vivo* (Zablotskaya et al. 2017). Tetrahydroisoquinoline derivatives and compounds possessing substituents with chain length of 10 or 11 carbon atoms showed the strongest antifungal activities with MIC values between 8 and 64 $\mu\text{g/ml}$. Tetrahydroisoquinolinium heptyl and decyl derivatives **30** exhibited high antifungal action as compared with corresponding tetrahydroquinolinium analogues. These compounds exhibited stronger antifungal activity than fluconazole.

Quinoline based 1,2,3 triazoles emerged as a group of very potent antifungal compounds with IC_{50} values (half maximal inhibitory concentration) for the most active derivative **31** (Fig. 5) of 0.044 $\mu\text{g/ml}$ against *C. albicans*, 12.02 $\mu\text{g/ml}$ against *C. glabrata*, and 3.60 $\mu\text{g/ml}$ against *C. tropicalis* (Irfan et al. 2015; Irfan et al. 2017). Moreover, these antifungal concentrations were not cytotoxic. The presence of these compounds affected the secretion of extracellular hydrolytic enzymes (proteinases and phospholipases). The antifungal target for these triazoles was plasma membrane as suggested by altered cell membrane, reduced plasma membrane H^+ ATPase activity, and significant inhibition in ergosterol biosynthesis in the presence of these compounds.

A series of 2,5 and 1,5-regioisomers of tetrazole with quinoline and benzylthio substituents, were synthesized (Shaikh et al. 2017). The designed compounds consist of a quinoline nucleus, which serves as pharmacophore and the non-heterocyclic part such as phenyl group separated by a tetrazole bridge. The antifungal activity of synthesized compounds was tested against *A. fumigatus* and *C. albicans* strains and the obtained MIC values

for the most active compounds **32** and **33** (the 2,5- and 1,5-isomer with fluoro substituent at the C(6)-position) (Fig. 5) were between 2.5 and 25 $\mu\text{g/ml}$. All the derivatives having halogens as substituents showed better activity than other derivatives. The active compounds came to be inhibitors of dihydrofolate reductase and N-myristoyl transferase, and exhibited no cytotoxic effects.

New hybrid analogues containing 7-chloro-4-aminoquinoline and 2-pyrazoline N-heterocyclic fragments were synthesized and evaluated for antifungal activity against *C. albicans* and *C. neoformans* (Montoya et al. 2016). The compounds displayed stronger activity against *C. neoformans* with the most active derivative showing MIC_{50} values of 15.5 and < 3.9 $\mu\text{g/ml}$ against *C. albicans* and *C. neoformans*, respectively.

In an attempt for development of novel antimicrobial agents, three series of quinoline derivatives bearing pyrazole moiety were synthesized (El Shehry et al. 2018). The most active compound **2** (Fig. 3) showed fourfold potency of amphotericin B in inhibiting the growth of *A. clavatus* (MIC 0.49 $\mu\text{g/ml}$) and *C. albicans* (MIC 0.12 $\mu\text{g/ml}$), respectively. The same compound showed equipotent activity to amphotericin B in inhibiting the growth of *A. fumigatus* (MIC 0.98 $\mu\text{g/ml}$).

New quinoline derivatives (Khan et al. 2019) bearing vinyl benzylidene imine with substituted aniline at the C(2)-position showed antibiofilm and antifungal activity. The antibiofilm activity of compounds **34** and **35** (Fig. 5) against *C. albicans* (IC_{50} values 66.2 and 51.2 μM , respectively) were similar to activity of fluconazole (IC_{50} = 40.0 μM). Compound **34** exhibited slightly lower antifungal activity (MIC = 94.2 $\mu\text{g/ml}$) than fluconazole (MIC = 50.0 $\mu\text{g/ml}$).

Benzimidazole fused pyrrolo[3,4-b]quinoline (Villa et al. 2019) compound **36** (Fig. 5) exhibited potent antifungal activity against *C. albicans*, *C. parapsilosis*, *C. tropicalis*, *C. glabrata*, and *C. neoformans* with the MIC values ranging from 0.0625 to 2 $\mu\text{g/ml}$. Fungal strains that are resistant to fluconazole were also inhibited by **36** with the MIC ranging from 0.25 to 0.5 $\mu\text{g/ml}$.

These compounds also showed antibiofilm activities without apparent toxicity to mammalian cells. A fluorescent benzimidazo[1,2- α] quinoline was identified within a set of nine benzimidazole derivatives as bifunctional *Candida* spp. biofilm detector and eradicator (de Souza et al. 2016). Compound **37** (Fig. 5) was the most active derivative showing antifungal activity against different *Candida* strains including *C. tropicalis*, *C. albicans*, and *C. parapsilosis* with MIC values in the range of 4 $\mu\text{g/ml}$ against *C. albicans* and 32 $\mu\text{g/ml}$ against the other isolates. Spraying of **37** over *Candida* sp. biofilm contaminated surface enabled detection of the biofilms under UV light. At the same time, **37** showed the potential to eradicate the detected biofilms. Benzimidazole **37** thus has a good potential to enable the usage of disinfected medical and surgical instruments in clinical and surgical procedures contributing to increased safety for patients.

As described, quinoline scaffold can be considered a valuable pharmacophore to be used for a simple synthesis and development of novel antifungal agents with the superior efficacy to commonly used antifungal drugs. The structural diversity of synthesized compounds can provide high and selective activity achieved through different modes of action, as well as low cytotoxicity on human cells.

4 Anti-virulence Activity

P. aeruginosa produces more than 50 different alkylquinolines (Deziel et al. 2004) of which 2-heptyl-3-hydroxy-4-quinolone (PQS) and its direct precursor 2-heptyl-4-hydroxyquinoline (HHQ) are PQS pathway autoinducers regulating production of several virulence factors such as pyocyanin and hydrogen cyanide, as well as biofilm formation. Alkylquinolines signalling has so far been detected in *P. aeruginosa* and certain *Burkholderia* and *Alteromonas* species (Diggle et al. 2006; Vial et al. 2008).

The majority of bacterial QS inhibitors (QSIs) have been designed based on the core structures of autoinducers. Quinolone-based compounds are expected to exhibit species-specific QSI

activity against *Pseudomonas*, some *Burkholderia* and *Alteromonas* species.

Following a ligand-based drug design approach, a set of HHQ and PQS analogs were synthesized by varying the side chain and introducing substituents into the benzene moiety of the quinolone molecule. PQS analogues with hydroxy group showed PqsR agonistic activity in *P. aeruginosa*, and this effect was circumvented by synthesizing a series of HHQ analogs (Lu et al. 2012). Highly potent PQS inhibitors have been synthesized by introducing modifications in benzene ring and 3-alkyl substituents achieving almost complete PqsR inhibition at 5–10 μM concentrations (**38**, Fig. 6) and reduction of virulence factor pyocyanin production by 74% at 3 μM . Introduction of polar groups at the C(3)-position of derivative **39** resulted in novel compounds with enhanced anti-virulence activity, with the most active derivative **40** inhibiting pyocyanin production at IC_{50} of 3.8 μM (Lu et al. 2014a). The QSI activities of these compounds were confirmed in *in vivo* experiments (Lu et al. 2014b).

A 4-quinolone isostere quinazoline was designed as highly potent competitive inhibitor of PqsR with derivatives **41–43** being the most active ones (Fig. 6) (Ilangoan et al. 2013). A simple isostere replacement (OH for NH_2) changed quinolone isostere quinazoline activity from potent agonistic to potent antagonistic. Derivative **41** inhibited pyocyanin production, virulence genes expression, aminoquinoline biosynthesis, and biofilm development.

A series of 4-aminoquinoline derivatives have been described as dual *P. falciparum* and botulinum neurotoxin inhibitors (Opsenica et al. 2012; Videnovic et al. 2014; Solaja et al. 2008) as well as ligands in complexes with anticancer activities (Nikolić et al. 2015). Following the drug repurposing approach several 4-aminoquinoline-based molecules with QSI activities have been further developed. Within a series of 31 derivatives of antiprotozoal 4-aminoquinolines compounds with PqsR antagonistic activity were identified inhibiting receptor activity by 70–85% at 10 μM (Soukarieh et al. 2018). Derivatives **44** and **45** inhibited pyocyanin production, while compound **44**, inhibited also

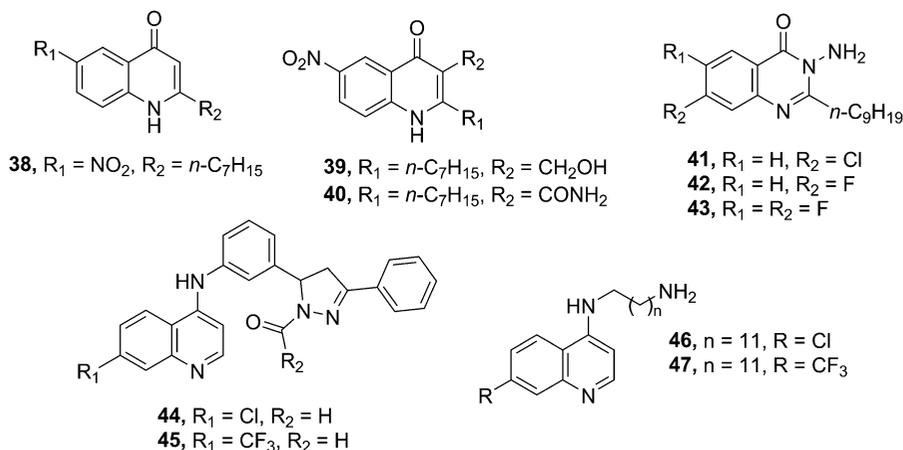


Fig. 6 Quinoline and quinolone derivatives with anti-virulence activity

HHQ, PQS, and 2-heptyl-4-hydroxyquinoline N-oxide (HQNO) production, and biofilm formation (Fig. 6). The efficacy of these compounds depended on the strains of *P. aeruginosa* (*P. aeruginosa* PA14 vs. *P. aeruginosa* PAO1).

Recently, PQS inhibitory activity of long-chain 4-aminoquinoline derivatives (Fig. 6) has been reported (Aleksić et al. 2017). Within this series, derivatives **46** and **47** with C12 alkyl chain at C(4)-position and chlorine at C(7)-position were the most active anti-virulence compounds inhibiting biofilm formation in *P. aeruginosa* with IC_{50} values 63 μM and 69 μM , respectively, and pyocyanin production with IC_{50} values of 40 and 2.5 μM , respectively. The same compounds also inhibited biofilm formation in *Serratia marcescens* with the same efficacy.

PQS, HHQ and some of the 4-quinolone derivatives can modulate interspecies and interkingdom interactions (Fernández-Piñar et al. 2011; Reen et al. 2011, 2012, 2015), which makes this chemotype even more attractive for further development of QSI.

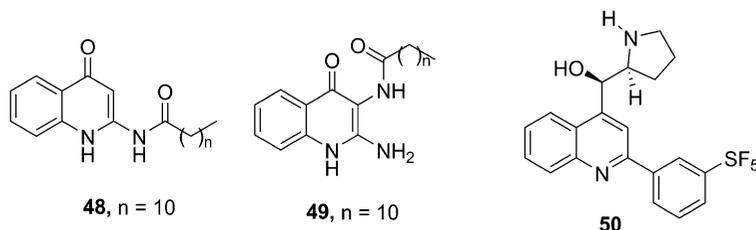
Long chain amide derivatives of 2-amino-4-quinolone showed ability to inhibit biofilm formation in *P. aeruginosa* and *S. aureus* (Espinosa-Valdes et al. 2019). The most active compounds were those with an alkyl chain of more than 12 carbon atoms and in general they were more active against biofilm formation in *P. aeruginosa* than in *S. aureus*. The most active compounds **48**

and **49** (Fig. 7) also inhibited pyocyanin production between 62.6 and 68.2% at concentration of 20 μM . None of the tested compounds affected bacterial growth suggesting that observed activity of the derivatives could be through modulation of QS.

A library of quinoline amino alcohol derivatives have been evaluated as biofilm inhibitors against the Gram-negative pathogen *Vibrio cholerae* (Leon et al. 2015). The most potent compound was *meta*-substituted pentafluorosulfonyl derivative **50** (Fig. 7) with BIFC_{50} of 4.4 μM and BDIC_{50} value of 7.4 μM . The antibiofilm activity of this compound was observed at concentration that did not affect bacterial growth ($\text{MIC} = 78.1 \mu\text{M}$) suggesting that mode of antibiofilm action was different from its bactericidal activity. This compound also exhibited cytotoxicity to HeLa cells with IC_{50} value of 7.27 μM , thus it can be considered as active component for functionalization of materials for medical use rather than for systemic application.

Chloroquine (CQ, Fig. 1) has been tested in combination with antifungal drugs fluconazole, voriconazole, amphotericin B, and caspofungin for ability to inhibit *C. albicans* planktonic forms and biofilms (Shinde et al. 2013). No synergistic activity of CQ and antifungals has been observed against planktonic growth. Alone fluconazole and voriconazole didn't affect formation

Fig. 7 Quinoline and quinolone derivatives with antibiofilm activity



of *C. albicans* biofilms even at high concentrations, but in a combination with 250 $\mu\text{g/ml}$ CQ, antibiofilm activity was observed at fluconazole concentration of 4 $\mu\text{g/ml}$ and voriconazole concentration of 0.25 $\mu\text{g/ml}$. Combinations of CQ with amphotericin B, or CQ with caspofungin showed no synergistic activities against biofilm formation. Antifungal drugs fluconazole and voriconazole showed no effect on mature *C. albicans* biofilms, while amphotericin B and caspofungin inhibited mature biofilms but high concentrations were required (0.5 and 1 $\mu\text{g/ml}$, respectively). In the presence of 250 $\mu\text{g/ml}$ CQ, mature biofilms were disrupted as a result of synergistic activity with fluconazole and voriconazole, while combination with amphotericin B and caspofungin showed low effect against mature biofilms. CQ can be developed as a promising partner molecule of antifungal drugs for combination therapy against *C. albicans* biofilm.

5 Antiviral Activity

Viral infections have concerning effects on the health of human populations worldwide. The successful treatments of these infections require highly effective antiviral drugs, which together with the growing spread of resistance require new therapeutic agents. A lot of efforts has been done on antiviral properties of quinolines and quinolones and their structural analogues against human immunodeficiency virus (HIV), but their antiviral activity was also demonstrated against human cytomegalovirus (HCMV), SARS corona virus, Zika virus, Chikungunya virus, hepatitis C virus (HCV), and Ebola virus (Luthra et al. 2018;

Plantone and Koudriavtseva 2018; Al-Bari 2015; Delvecchio et al. 2016; Barbosa-Lima et al. 2017; Loregian et al. 2010). Chloroquine shows antiviral effects by inhibiting pH-dependent steps of the replication of several viruses including members of the flaviviruses, retroviruses, and coronaviruses (Savarino et al. 2003).

Human immunodeficiency virus type 1 (HIV-1) integrase (IN), HIV-1 RT and HIV-1 protease are the essential enzymes for retroviral replication, and represent the important targets for interrupting the viral replication cycle and thus development of novel antiviral therapeutics. Reverse transcriptase and protease inhibitors, which have been used in retroviral therapy, cannot achieve complete suppression and there is a risk for resistance development. The best-studied effects of CQ are those against HIV replication, and its analogues have been used in clinical trials as investigational anti-retroviral agents in humans with HIV-1/AIDS (Al-Bari 2015). Approved 51 (Fig. 8), later called elvitegravir, is the first quinolone-based anti-HIV drug, exhibiting potent inhibitory activity against integrase-catalyzed DNA strand transfer (Sato et al. 2006). Recently, a series of quinolone-3-carboxylic acids have been synthesized as HIV-1 integrase inhibitors featuring a fluorine atom at C(5) position (He et al. 2013). The most active compound 52 (Fig. 8) exhibited activity against both wild-type and the mutant virus, with an EC_{50} value of 0.032 and 0.082 μM , respectively. Another series of quinolone-3-carboxylic acids have been synthesized by introducing different hydrophobic groups at N(1), C(2), C(7), and C(8) positions (Hajimahdi et al. 2016). Most of the compounds of this group showed anti-HIV activity without cytotoxicity at concentration of 100 μM . The

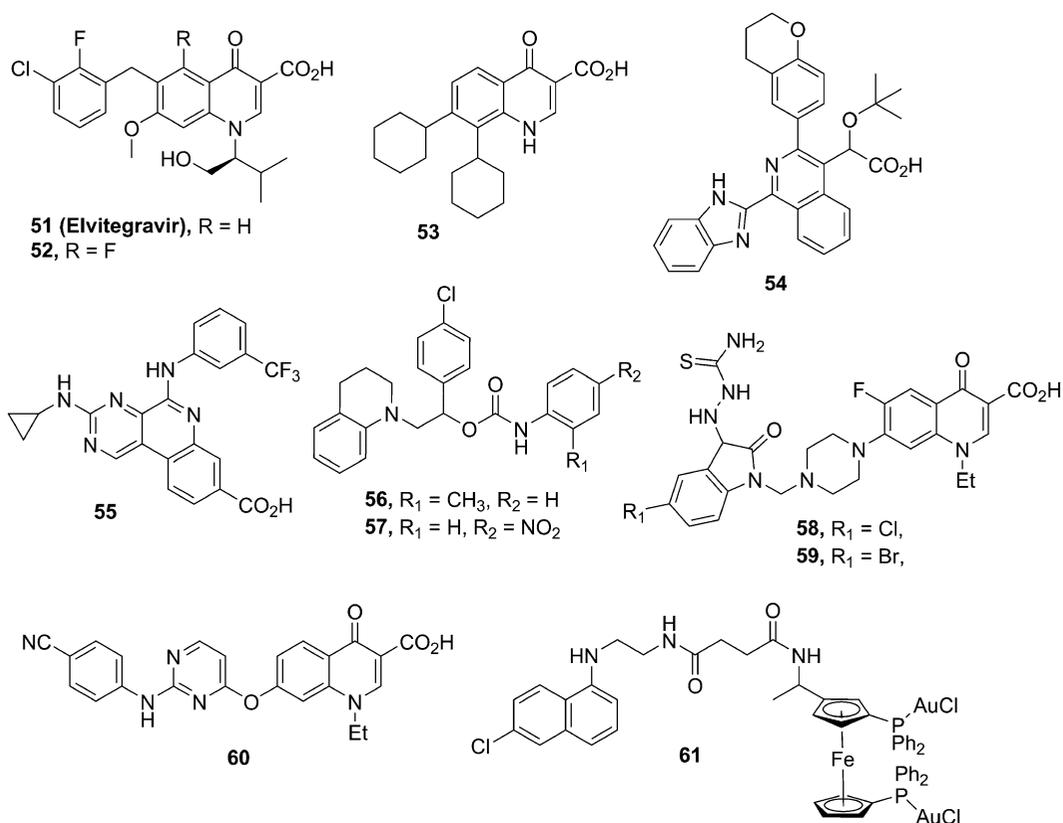


Fig. 8 Quinoline and quinolone derivatives with anti-HIV activities

most active compound **53** (Fig. 8) showed anti-HIV activity with an inhibition rate of 84%. Docking study revealed that the anti-HIV activity of this group of compounds might involve a metal chelating mechanism. Several other quinoline analogues have been synthesized, which exhibit low micromolar inhibitory potency against HIV-1 IN as recently reviewed by Wadhwa et al. (Wadhwa et al. 2018). A series of multi-substituted quinolines, with the focus on the substitution pattern of the 4-phenyl moiety and incorporation of heteroaromatic or polycyclic substituents at the C(4)-position, were prepared and examined for their ability to trigger HIV-1 IN multimerisation *via* binding to an allosteric site (Jentsch et al. 2018). 4-(4-Chlorophenyl)quinoline and 2,3-benzo[b][1,4]dioxine showed highest potency with EC₅₀ values (half maximal eradication concentration) of 0.1 and 0.08 μM , respectively. A series of isoquinoline analogues with

allosteric IN inhibitory activity has been recently reported (Wilson et al. 2019). Compound **54** (Fig. 8) was highly potent with EC₅₀ of 1.1 μM .

Protein kinase CK2 plays a role in the stimulation of RT and protease. Substituted pyrimido [4,5-c]quinoline ATP competitive inhibitors of protein kinase CK2 showed a good antiviral activity with the compound **55** (Fig. 8) having IC₅₀ values as low as 80 nM against HIV-1 viruses with an excellent therapeutic index (Pierre et al. 2011).

Using the ligand based drug design approach the 1-(4-chlorophenyl)-2-(3,4-dihydroquinolin-1(2H)-yl)ethyl phenylcarbamate derivatives were designed as inhibitors of HIV-1 RT (Chander et al. 2016). The most active compounds **56** and **57** (Fig. 8) inhibited the RT activity with IC₅₀ 8.12 and 5.42 μM , respectively. Cytotoxicity and anti-HIV activity of these compounds were evaluated on T

lymphocytes. Both compounds exhibited potent anti-HIV activity with EC_{50} values 11.10 and 2.76 $\mu\text{g/ml}$, for **56** and **57** respectively, while showing a very good safety index.

A molecular-hybrid approach is a powerful tool in the design of new molecules with improved affinity and efficacy and as such was applied for development of novel antiviral agents involving quinoline scaffold. The fluoroquinolone-isatin hybrids **58** and **59** (Fig. 8) showed inhibition on the replication of HIV-1 in human cells with EC_{50} of 11.3 and 13.9 $\mu\text{g/ml}$, and the selectivity index values were > 5 . The promising antiviral properties of isatin and fluoroquinolone hybrids have been extensively discussed recently (Xu et al. 2019). Further, a series of diarylpyrimidine–quinolone hybrids was synthesized and evaluated against both wt HIV-1 and mutant viral strains (Mao et al. 2015). The most active hybrid **60** (Fig. 8) displayed an EC_{50} value of $0.28 \pm 0.07 \mu\text{M}$ against HIV-1 by targeting RT.

HIV infects multiple cells in human body, but its replication starts after infecting CD4 lymphocytes (T-cell or CD4- cell). The treatment of proliferating CD4 T-cells with antiproliferative activity agents reduces the ability of these cells to support HIV replication. Combination therapy involving cytostatic compounds such as hydroxychloroquine or hydroxyurea with proven anti-retroviral drugs as didanosine decreases viral replication and increases the CD4 count in antiretroviral-naive HIV patients thus helping to control the infection. A novel di-gold(I) complex of ferrocene–quinoline (**61**, Fig. 8) was investigated for cytostatic behaviour as well as antiviral activity (Gama et al. 2016). Di-gold quinoline derivative showed inhibition of virus infectivity by 83% at concentration of 10 $\mu\text{g/ml}$ and cytostatic activity with significant S and G2/M phase cell arrest, which make it a good candidate for use in HIV-1 infection as a virostatic agent.

Although quinoline/quinolone derivatives showed promising antiviral effects in *in vitro* studies, published clinical studies evaluating the effects of chloroquine/hydroxychloroquine

administration, alone or in combination with other drugs, in HIV infected subjects reported different effects in terms of immune activation, viral load, and CD4 counts (Savarino and Shytaj 2015). The outcome of the studies depended on applied doses, duration of the treatments or the drug exposure, or on the individual differences in drug metabolism and distribution.

6-Aminoquinolones with amino group at the C (6) position of the bicyclic quinolone ring system, specifically inhibit HIV replication (Cecchetti et al. 2000). On the other side, **62** (Fig. 9), which is characterized by a cyclopropyl group at the N(1) position and a 4-(2-pyridyl)-1-piperazine moiety at the C(7) position showed specific activity against HCMV by inhibiting transactivation activity of immediate-early 2 (IE2), a multifunctional factor essential for viral replication (Loregian et al. 2010; Mercorelli et al. 2014). The target of this compound is different from that of the anti-HCMV drugs currently approved for clinical use. **62** demonstrated activity against laboratory strains of HCMV but also against clinical isolates and virus strains resistant to clinically relevant anti-HCMV agents. Importantly, this compound does not show cross-resistance with ganciclovir, acyclovir, cidofovir, and foscarnet, which is important advantage for its potential clinical use, since drug-resistant HCMV strains often emerge after long-term treatment with antiherpetic drugs.

Most of the current clinical compounds and approved drugs against HCV target non-structural (NS) proteins NS3, NS5A, and NS5B. 3-Heterocyclyl quinolones have been described as a series of allosteric-site (NNI-2) inhibitors of the HCV NS5B polymerase (Kumar et al. 2012). The most potent compounds in this series were **63** and **64** (Fig. 9), with EC_{50} below 250 nM. Derivatives of 1H -quinazolin-4-one were reported as allosteric HCV NS5B thumb pocket 2 (TP-2) inhibitors (Hucke et al. 2014) with picomolar antiviral potency in genotype (gt) 1a and 1b ($EC_{50} = 120$ and 110 pM, respectively) and with $EC_{50} \leq 80$ nM against gt 2–6.

A group of 6-aminoquinolone derivatives demonstrated inhibitory activity against NS5B

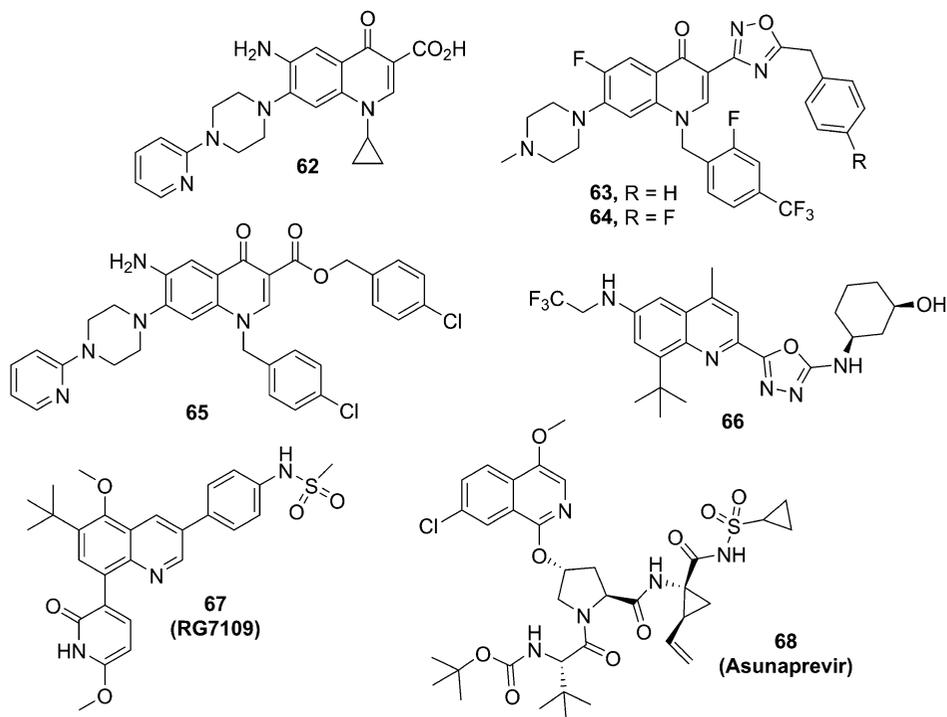


Fig. 9 Quinoline and quinolone derivatives with anti-HCMV and anti-HCV activities

polymerase (Manfroni et al. 2014). The most active compound of this group was 6-amino-7-[4-(2-pyridinyl)-1-piperazinyl]quinolone derivative **65** (Fig. 9) with an IC_{50} value of 0.069 μ M. Its selective antiviral effect was EC_{50} 3.03 μ M, while cytotoxicity was not observed.

Derivatives of 2-oxadiazoloquinoline were designed to inhibit HCV NS4B, which is integral membrane protein believed to act primarily as an endoplasmic reticulum-localized scaffold for the assembly of the replicase complexes needed for HCV RNA replication (Phillips et al. 2014). Compound **66** (Fig. 9) from this series showed exceptional activity against different replicons with EC_{50} values ranging between 0.08 nM and 3.1 nM.

The compound **67** (**RG7109**, Fig. 9), was synthesized as a result of an effort to develop a combination therapy against HCV consisting of only direct-antiviral agents. The standard therapy of HCV included pegylated interferon α (Peg-IFN) and ribavirin (RBV), however this therapy was effective against gt 2 or 3, but gt

1 did not respond well, with SVR (absence of detectable HCV RNA in blood serum for 24 weeks after treatment withdrawal) rates of <50% even after 48 weeks long therapy. Upon FDA approval of HCV protease inhibitors the treatment of gt 1 included Peg-IFN and RBV and a protease inhibitor, which improved the SVR rates up to 90% with gt 1 naïve patients. Interferon-associated side effects limit treatment success in a large number of patients. With the aim to develop a combination therapy consisting of only direct-antiviral agents, Talamas and co-workers identified a bicyclic template with potent activity against the NS5B polymerase containing 3,5,6,8-tetrasubstituted quinoline core **67**, which was selected for advancement to clinical development (Talamas et al. 2014).

Isoquinoline-containing compound called asunaprevir (**68**) (Fig. 9), a tripeptidic acylsulfonamide inhibitor of the NS3/4A enzyme developed by Scola and co-workers (Scola et al. 2014a; b) went through phase III clinical trials for efficient treatment of HCV infection.

6 Derivatives with Anti-parasitic Activity

Parasitic diseases caused by protozoan species represent one of the major health challenges worldwide, affecting millions of people of all ages and social classes. The most threatened are children, pregnant women, immunocompromised individuals, and people living in developing countries. Consequences of illness burden not only patients and their families, their financial status, but also local and global economy. Taking into account the limitations of currently available drugs, the only proven strategy to fight against these diseases is development of the new therapeutics under clearly defined criteria (Nwaka and Hudson 2006).

6.1 Antimalarial Activity

According to WHO annual report, in 2017 approximately 219 million cases of malaria occurred worldwide, resulting in 435,000 deaths (World Health Organisation 2019). Most cases were registered in African regions, followed by South-East Asia and Eastern Mediterranean regions. Malaria is caused by five *Plasmodium* species with *Plasmodium falciparum* as the most dangerous one, since it causes cerebral malaria and is the major cause of death.

Although many different pharmacophores have been developed during past decades, quinoline derivatives (Figs. 1 and 10) are still a backbone for development of novel antimalarial drugs (Barnett and Guy 2014). Actually, one of the most successful antimalarial therapeutics, a well-known quinoline derivative chloroquine, has been used for decades to treat this illness. The main mechanisms of action of CQ and its congeners (Figs. 1 and 10) are inhibition of hemozoin polymerization and release of a free toxic haem (Egan and Marques 1999). Among diverse antimalarials used in clinics CQ, considered as safe even for children and pregnant women, is the easiest to synthesize. Unfortunately, malaria parasite has developed resistance

to most of antimalarial drugs including CQ. Structural modifications of this chemotype, which would include altering the nature of the quinoline core and side chain at C(4)-position, or synthesis of organic (Boudhar et al. 2016) or organo-metallic hybrids (Salas et al. 2013) present a good direction for obtaining novel derivatives with better activity against CQ resistant strains (Manohar et al. 2013).

Introducing of steroidal structure fragment, various thiophen, benzothiophen and adamantyl groups, numerous derivatives of 4-aminoquinoline have been synthesized and tested for their antimalarial activity. Steroidal compounds reported by (Videnovic et al. 2014), which were developed based on the results of the previous study (Solaja et al. 2008), showed low nanomolar IC_{50} (50% inhibition of β -haematin formation) activities against CQ resistant and MDR *P. falciparum* strains. Among them derivatives 69–71 were the most active in the series (Fig. 11), being more active than mefloquine (Fig. 10) against MDR strains. High potency of these derivatives was demonstrated with three times better inhibition of hemozoin production than CQ. Derivatives bearing electron-rich thiophen or benzothiophen groups were designed to enhance activity against CQ resistant and MDR strains of *P. falciparum*, by additional π -interactions with hem (Opsenica et al. 2015). 4-Fluoro derivatives, 72 and 73, were the most active in *in vivo* experiments showing curative activity (Fig. 11). Addition of aromatic ring, elongation of diaminoalkyl linker, and replacement of F-atom with CN-group (74–76, Fig. 11) enhance active against CQ resistant strain (Konstantinovic et al. 2017). The most active benzothiophene derivative 77 cured mice infected with *P. berghei* when dosed orally (Fig. 11). Derivatives with adamantyl-substituent, 78–81 (Fig. 11) (Terzic et al. 2016) were designed to increase lipophilicity of quinoline core and to provide amphiphilic structure (Wanka et al. 2013) and metabolic stability. These compounds demonstrated *in vitro* antimalarial activity at low nanomolar concentrations against CQ resistant and MDR strains, presenting the highest potency up to today. This modification increased

Fig. 10 The most-successful quinoline derived antimalarial drugs developed in the twentieth century

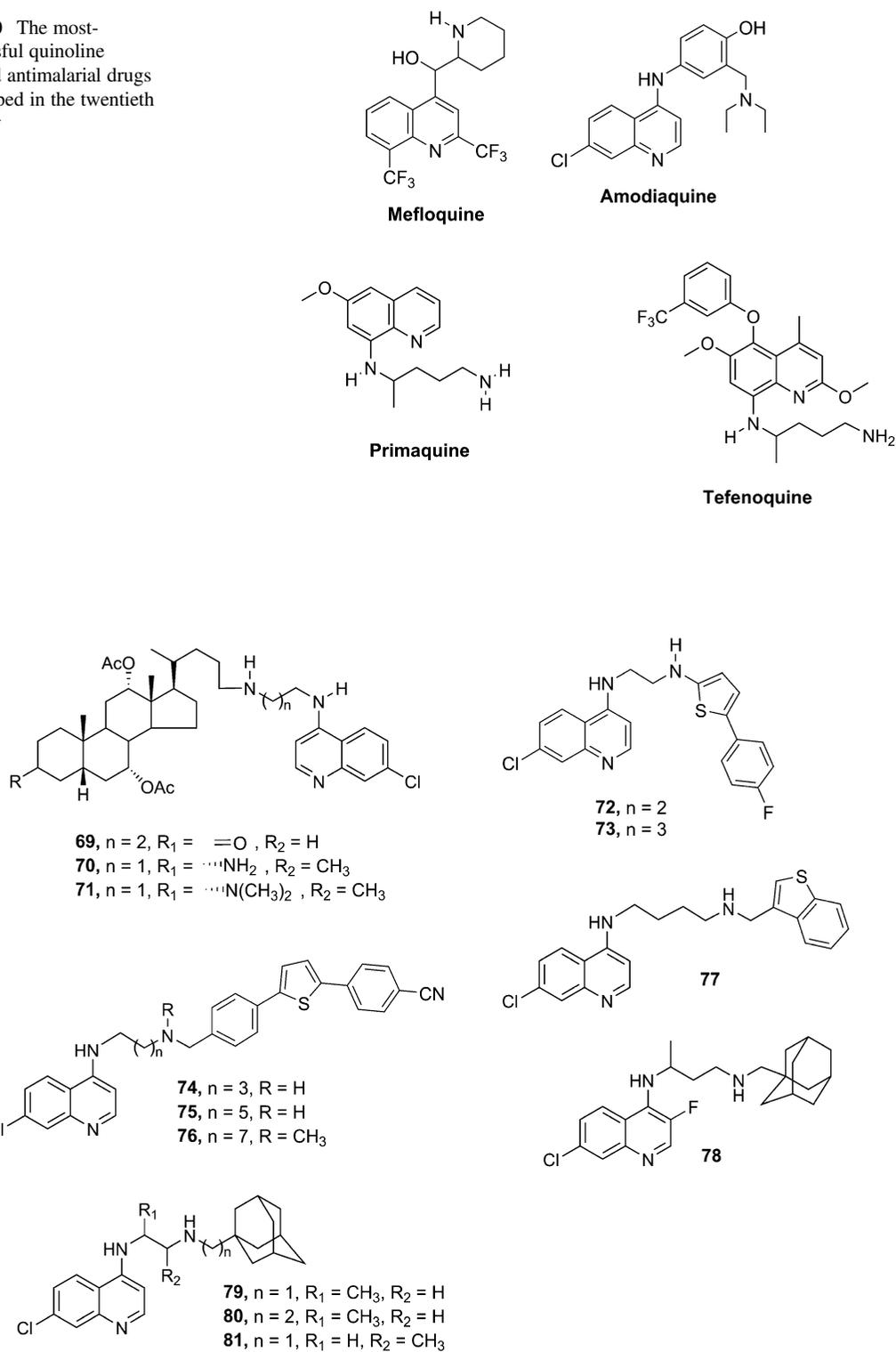


Fig. 11 Quinoline derivatives with antimalarial activity

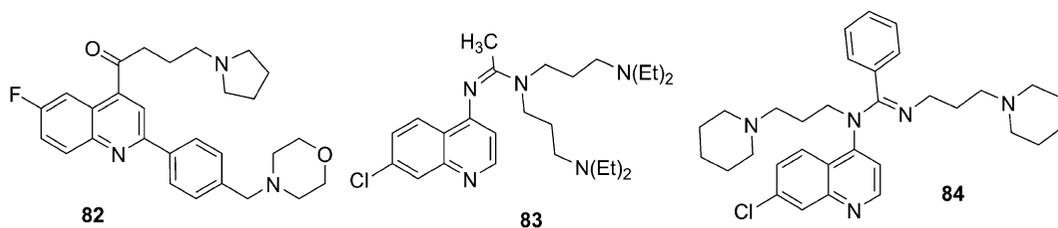


Fig. 12 Keto- and amidino-quinolines with antimalarial activity

microsomal metabolic stability over 60 min. Introducing the F-atom at C(3)-position attenuated *in vitro* activity against erythrocyte form of *P. falciparum*. However, five F-C(3) derivatives showed excellent *in vitro* and *in vivo* activity against liver stage of *P. berghei*, with **78** as the most active derivative with $IC_{50} = 0.1 \mu\text{M}$.

Changing the character of 4-amino group, or replacing it with different functional group, alters electronic properties of quinoline core. One of the such example is derivative **82** (Fig. 12) who contained carbonyl group attached to C(4) (Baragana et al. 2015). Compound **82** showed potent activity against multiple stages of the *Plasmodium* parasite life-cycle and exhibited excellent pharmacokinetic properties and an acceptable safety profile. This molecule having $EC_{90} = 2.4 \text{ nM}$ against MDR *P. falciparum* strain reduced parasitemia for 90% in mice infected with *P. berghei* after a single oral dose. Translation elongation factor 2, a protein responsible for the GTP-dependent translocation of the ribosome along mRNA and as such required for protein synthesis at all stages of the parasite life cycle was identified as the target. Therefore, with its potent activity against multiple stages of the parasite life cycle, high metabolically stability, novel mode of action, and excellent drug-like properties, **82** meets the key criteria for new antimalarial drug.

Few derivatives of yet another series of 4-amidinoquinolines, tested against CQ sensitive and CQ resistant strains, exhibited IC_{50} values less than 10 nM (Korotchenko et al. 2015). Introduction of the third substituent on the amidine-group provided both good activity and enhanced metabolic stability. Derivatives **83** and **84** (Fig. 12) showed activity with $IC_{50} = 1.9\text{--}34 \text{ nM}$.

However, those promising results were diminished with high resistant index (activity against resistant vs. activity against sensitive strains) ratio.

Amodiaquine (Fig. 10), well known and intensely studied drug, could replace CQ in anti-malarial therapy (Olliaro and Mussano 2003). It is successful in treatment of CQ resistant strains but unfortunately when used prophylactically toxic metabolites are produced (Schlitzer 2007). Therefore, a series of *N*-pyridyl, *N*-1H-benzo[d]imidazole, *N*-benzo[d]oxazole, and *N*-benzo[d]tiazole were tested (Ongarora et al. 2015). Three derivatives (**85–87**, Fig. 13) showed *in vivo* efficacies in dose-depend manner with 99.5% parasitemia reduction among which derivative **85** was the most successful. 8-Aminoquinolines primaquine and tefenoquine (Fig. 10) are also known and promising antimalarial compounds (Waters 2011). Shared common disadvantage of these compounds is biotransformation of terminal amino-groups into aldehydes, which are toxic. To prevent such transformation and to maintain good antimalarial activity a new derivatives that contained aminoxy and oxime-groups were synthesized (Leven et al. 2019). The compounds were evaluated *in vitro* against asexual blood stages, liver stages, and sexual stages of *P. falciparum*. 8-Aminoquinolines bearing a 2-alkoxy and a 5-phenoxy substituent (**88–92**, Fig. 13) were the most active compounds, with the IC_{50} values in the range 0.95–1.3 μM , comparable to primaquine and tefenoquine. Derivatives **93** and **94** showed significant activity against asexual blood stage of parasites (Fig. 13). However, identification and toxic profile of potential metabolites remained inexplicable.

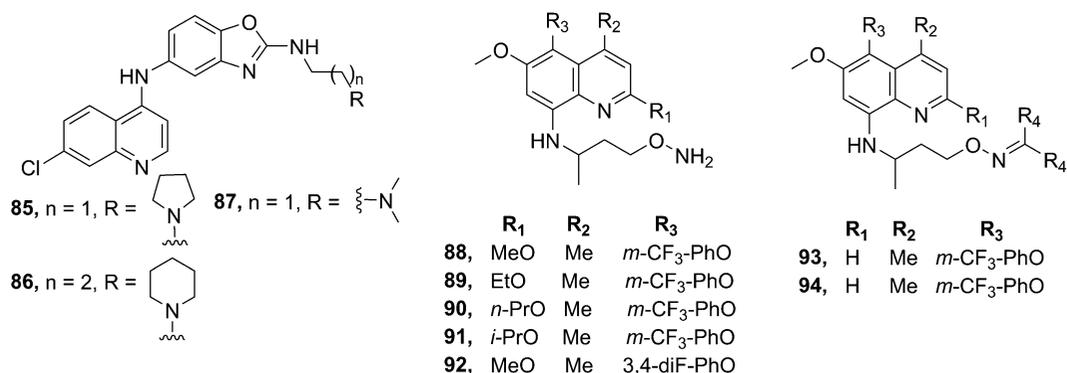


Fig. 13 4-Amino and 8-aminoquinolines with antimalarial activity

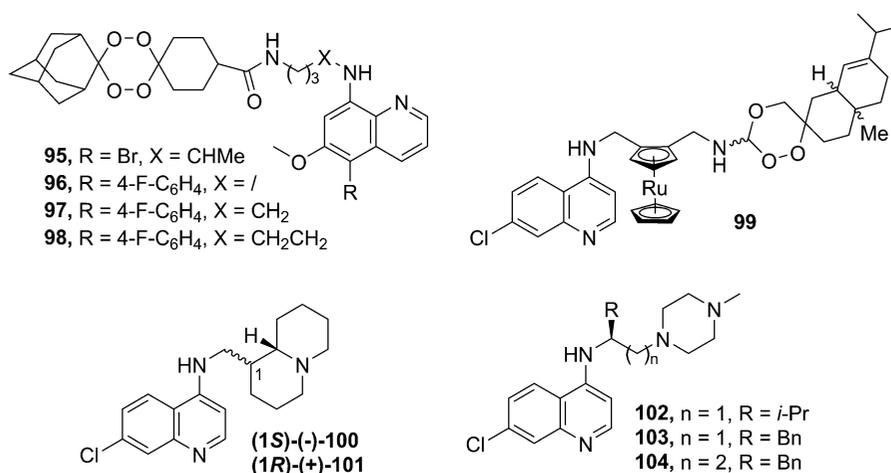


Fig. 14 Quinoline hybrids with antimalarial activity

Synthesis of hybrid compounds that contained two or more pharmacophore is a new promising approach in development of more successful antimalarials. It was expected that those hybrid-compounds cover distinct mechanism of actions, or affect different phase of plasmodium life cycle; hence, synergistic effect was anticipated.

1,2,4,5-Tetraoxane-8-aminoquinoline hybrids with various aryl or heteroaryl group as substituents at the metabolically labile C(5)-position of the 8-aminoquinoline moiety were tested *in vitro* as dual-stage antimalarial agents against *P. falciparum* CQ resistant strain (blood stage) and *P. berghei* (liver stage) (Capela et al. 2018). Derivatives **95–98** (Fig. 14) have improved metabolic stability, and proved to be efficient as dual-stage antiplasmodial agents with

good metabolic stability, inhibiting the development of intra-erythrocyte forms of *P. falciparum* ($EC_{50} = 15\text{--}17$ nM) and *P. berghei* liver stage ($EC_{50} = 1.11$ μM).

Hybrid compound **99** (Fig. 14) containing a ruthenocene, 4-aminoquinoline, and a 1,2,4-trioxane motifs exhibited $IC_{50} = 65.33$ nM and $IC_{50} = 62.0$ nM against CQ resistant and MDR strain of *P. falciparum*, respectively (Martínez et al. 2017). As such derivative **99** was better than CQ, and the parent metal-free and quinoline-free trioxane, indicating possible route for designing new derivatives that could overcome resistance problem.

Hybrid derivative **100** and **101** (Fig. 14), contained 4-aminoquinoline and a quinolizidine (Sparatore et al. 2005) fragments displayed a very

attractive bioactivity profile (Basilico et al. 2017). Both racemic (1*R*/1*S*)-AM1 and enantiomers exhibited an antimalarial activity in low nanomolar range against sensitive and CQ resistant strains, as well as against MDR strain of *P. falciparum*. Racemic (1*R*/1*S*)-mixture showed remarkable antimalarial activity against ten different isolates of *Plasmodium vivax* with three times higher potency than CQ. Enantiomer (1*S*)-**100** displayed an additive effect when applied in combination with dihydroartemisinin. All forms, racemate and enantiomers, were *in vivo* active with no significant differences in potency among them.

Enantiopure chiral 4-aminoquinoline derivatives that contain piperazine were synthesized (Dola et al. 2017) among which derivatives **102–104** (Fig. 14) showed activities in the range of 5.03–39.39 nM against *P. falciparum* strains. Compound **102**, showed the best activity *in vivo*, and successfully cured mice infected with CQ resistant *P. yoelii* strain. This derivative has been identified as a preclinical candidate molecule, due to excellent physico-chemical properties, an acceptable pharmacokinetic profile, and moderate metabolic clearance in liver microsomes.

Large number of examined quinoline derivatives has provided necessary information on structure-activity relationship and revealed new potential targets for the treatment of malarial parasite. However, although those results are highly valuable, new derivatives still have not brought much needed breakthrough in the therapy. Currently, in accordance with WHO recommendation, the most successful clinical treatments are combination therapies, which include established antimalarial drugs, such as artemisinin (Chico and Chandramohan 2011).

6.2 Antileishmanial Activity

Leishmaniasis is caused by *Leishmania* parasites and is transmitted to humans by the bite of infected female phlebotomine sandflies. The disease may occur in humans and animals, including those from the closest human surrounding. About

70 animal species and humans can be a reservoir of *Leishmania* and depending on the source it can be classified as zoonotic or anthroponotic. More than 20 different *Leishmania* species that infect mammals can cause infection in humans. There are four main forms of the disease: visceral leishmaniasis (also known as kala-azar), post-kala-azar dermal leishmaniasis, cutaneous leishmaniasis, and mucocutaneous leishmaniasis (World Health Organisation 2018). While cutaneous leishmaniasis is the most common form of the disease, visceral leishmaniasis is the most serious and almost always fatal if untreated. Based on the assumption that similar chemotypes could be active against related pathogens, broad series of quinoline-like structures (Woodring et al. 2015), and quinolines (Baragana et al. 2015; Devine et al. 2015; Konstantinovic et al. 2017; Solaja et al. 2008) were examined simultaneously as antimalarial, antileishmaniasis and antitrypanosomal agents, including application in combination therapy (Wijnant et al. 2017).

Ten out of thirty tested quinoline derivatives that contained adamantane or benzothiofene moiety exhibited IC₅₀ values less than 1 μM against *Leishmania infantum* and *Leishmania tropica* promastigote stage (Konstantinovic et al. 2018). Two derivatives, **105** and **106** (Fig. 15), exhibited dose-dependent *in vivo* activity, and a high level of mice survival and reduction of parasites level – up to 99% after 50 mg/kg/day and 5 days treatment. Those two compounds induced production of nitric oxide by IFNγ-primed macrophages, but only when highest doses were used. Also, both derivatives induced a persistent increase of reactive oxygen species (ROS) at all tested doses.

During the search for new scaffolds that would be able to inhibit three protozoan pathogens, quinoline derivatives **107–109** (Fig. 15) tested for inhibitory activity against two life stages (promastigote and amastigotes) of *Leishmania major* (Devine et al. 2015). While derivatives **107–109** showed high activities against promastigote life stage, exhibiting EC₅₀ values in the range 0.2–0.4 μM, only derivative **109** also showed activity against amastigotes life stage (EC₅₀ = 0.89 μM).

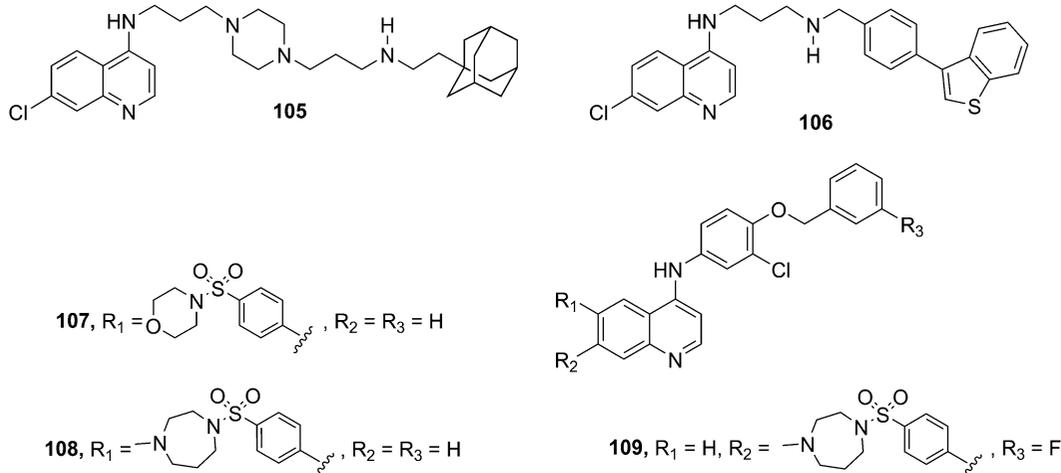


Fig. 15 4-Aminoquinolines with antileishmanial activity

Two derivatives of 4-aryloxy-7-chloroquinolines showed a strong inhibition of *Leishmania donovani* promastigote ($IC_{50} = 11.43 \mu\text{M}$ and $IC_{50} = 7.9 \mu\text{M}$) and amastigote proliferation ($IC_{50} = 1.01 \mu\text{M}$ and $IC_{50} = 1.02 \mu\text{M}$) (Valdivieso et al. 2018). Combinations of **110** or **111** (Fig. 16) with amphotericin B showed a synergistic anti-promastigote effect, while combinations of either **110** or **111** with miltefosine showed a synergistic anti-amastigote effect. Compounds showed deleterious effect on the mitochondrial electrochemical potential in IC_{50} concentrations, suggesting that rapid collapse of the parasite's mitochondrial membrane potential could be a mechanism of action.

A quinoline derivative **112** (Fig. 16) exhibited a strong effect against *Leishmania amazonensis*, inhibiting the growth of its promastigotes and intracellular amastigotes forms (Antinarelli et al. 2015; Antinarelli et al. 2018). The compound also affected the proliferation of amastigote-like forms of *L. amazonensis*, with $IC_{50} = 7.0 \mu\text{g/ml}$. Intramacrophage amastigotes treated with **112** showed a low capacity to reverse the effect of the compound in a drug-free medium. The strong effect on all parasite stages and the absence of reversibility of the action of **112** suggest that severe damage has been caused to the targets, and mechanism of its activity was preliminarily associated with mitochondrial dysfunction.

A quinoline derivative clioquinol (Fig. 5) inhibited *L. amazonensis* and *Leishmania infantum* promastigotes with $EC_{50} = 2.55 \mu\text{g/ml}$ and $EC_{50} = 1.44 \mu\text{g/ml}$, respectively, and also showed inhibitory activity against axenic amastigotes of *L. amazonensis* ($EC_{50} = 1.88 \mu\text{g/ml}$) and *L. infantum* ($EC_{50} = 0.98 \mu\text{g/ml}$) (Tavares et al. 2018). Significant reductions in the percentage of infected macrophages after treatment and in the pre-treated assay using clioquinol were observed. Clioquinol induces morphological and biochemical alterations in parasites, including reduction in cell volume, loss of mitochondrial membrane potential, increase in the ROS production and rupture of the plasma membrane.

N-Aryl derivatives of 2- and 3-aminoquinoline were evaluated as antiproliferative agents against *Leishmania mexicana*, the etiological agent of leishmaniasis and *Trypanosoma cruzi* (Chanquia et al. 2019). Fluorine-containing derivatives **113** and **114** (Fig. 16) were more than two-fold more potent than geneticin against intracellular promastigote form of *L. mexicana*, both exhibiting IC_{50} values of $41.9 \mu\text{M}$. The IC_{50} values of derivatives **113–115** (Fig. 16) were of the same order as drug benznidazole against epimastigote form of *T. cruzi*. The compounds were capable to inhibit the degradation of haem, inducing intracellular

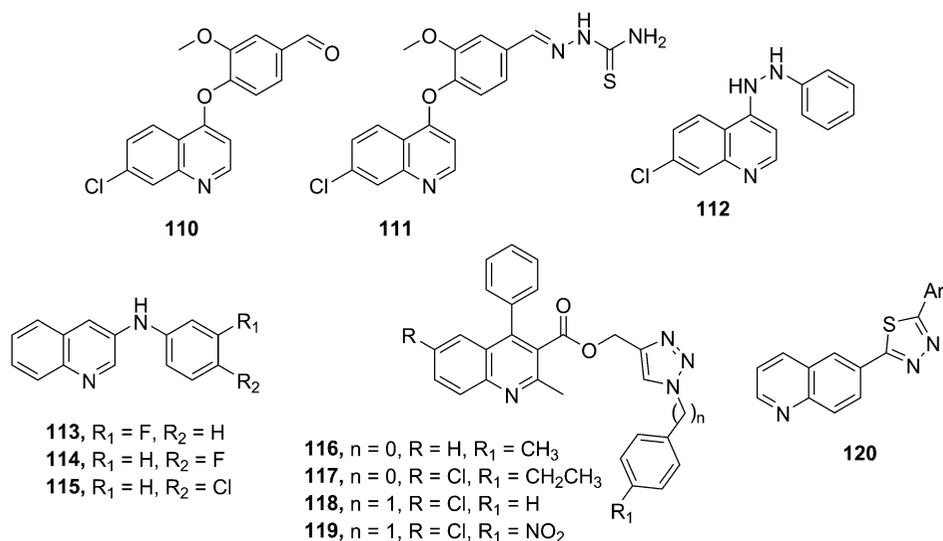


Fig. 16 Quinoline derivatives with antileishmanial activity

oxidative damage, which is not countered by the antioxidative defence system of the parasite.

Series of triazolyl esters of 2-methyl-4-phenylquinoline-3-carboxylic acid have been evaluated *in vitro* against *L. donovani* (Upadhyay et al. 2018). Most of the derivatives exhibited significant antileishmanial activity against promastigotes and intracellular amastigotes, with less cytotoxicity in comparison to the sodium stibogluconate. Four compounds **116–119** (Fig. 16) proved to be active derivatives, with lower toxicity and better selectivity index and were selected for *in vivo* evaluation. Compound **118** showed consistent activity up to day 28 post-treatment in a hamster model, which is promising finding as the hamster model of visceral leishmaniasis closely resembles the human condition.

Sixteen derivatives of 6-substituted quinoline analogues **120** (Fig. 16) showed high inhibitory potency against *L. major* promastigotes with IC₅₀ = 0.04–5.60 μM, and all were more active in comparison with pentamidine (IC₅₀ = 7.02 μM) (Almandil et al. 2019). Presence of either electron donating or electron withdrawing group on phenyl ring plays an important role in inhibition, but electron donating group showed better activity.

Leishmaniasis did not take much attention in R&D programs in comparison to other parasitic disease, like malaria. However, since the disease

is continuously increasing in Eastern Mediterranean Region, with 5 times growth in the period 1998–2015 (World Health Organisation 2017), there is an urgent need for changing of such a treatment. Although many quinolines exhibited good antileishmanial activity, their targets are still unknown.

Identification of the targets is prerequisite to direct further structural modifications of quinoline scaffold in order to optimize their therapeutic activity.

6.3 Antitrypanosomal Activity

Trypanosoma is unicellular parasitic protozoa which causes infections in humans in the tropical and sub-tropical areas. Trypanosomes infect a variety of hosts and cause fatal disease, such as sleeping sickness in humans. More than 19 different *Trypanosoma* species were identified as causative agents of diseases that differ in symptoms, action and main host. *Trypanosoma cruzi* causes American trypanosomiasis, or Chagas disease, when it infects a wide variety of wild and domestic mammals, rodents, many species of bloodsucking triatomid insects – the usual vectors of disease, and humans. The other members of the genus *Trypanosoma* that cause disease in humans

are two subspecies of African trypanosomes, *T. brucei* subspecies *gambiense* and *T. brucei* subspecies *rhodesiense*. These organisms cause West African and East African trypanosomiasis, respectively. The main limitation of the drugs that are currently in use against *Trypanosoma* is toxicity and requirement for intravenous dosing, thus novel drugs are needed. One approach is to develop compounds that show activity against molecular targets that are common for related pathogens. For example, *T. brucei* (human African trypanosomiasis), *T. cruzi* (Chagas' disease), *Leishmania* spp. (leishmaniasis), and *Plasmodium* spp. (malaria) express kinases and phosphodiesterases which are involved in the process of cellular signalling (Gould and de Koning 2011; Parsons et al. 2005). Recently it was found that certain human kinase inhibitors could be also effective against *T. brucei* with EC₅₀ in the low micromolar range (Katiyar et al. 2013), including tyrosine kinase inhibitors and new compounds developed on that basis (Patel et al. 2013).

Quinoline and quinoline-like compounds were tested for their antiprotozoan activity (Devine et al. 2015). From 4-aminoquinoline scaffold, derivatives **121–123** (Fig. 17), and derivatives **108** and **109** (Fig. 15) showed medium nanomolar activities against *Trypanosoma brucei* and *Trypanosoma cruzi*. In general, derivatives showed better activity against *T. brucei* (ED₅₀ = 79–450 nM), than against *T. cruzi* (ED₅₀ = 730–950 nM), with exception of derivative **122**, which is more active against *T. cruzi* (EC₅₀ = 90 nM vs. EC₅₀ = 350 nM).

The TbrPDEB1 enzyme, a member of cyclic nucleotide phosphodiesterases was used as a

target for series of 3-carbamido-4-aminoquinolines (Ochiana et al. 2015). Derivatives **124–127** (Fig. 18) exhibited the highest activity, inhibiting >80% of TbrPDEB1 and exhibiting IC₅₀ in the range 3.5–6.4 μM.

Another derivatives of chalocone (**128**; general structure **A**, Fig. 18) and corresponding 4,5-dihydro-1H-pyrazole derivatives (**129**; general structure **B**, Fig. 18) were screened against intracellular amastigotes of *T. cruzi* and intracellular amastigotes of *Leishmania (Vianna) panamensis* (Ramirez-Prada et al. 2017). Compound **129** was highly active against *T. cruzi* (EC₅₀ = 0.70 μg/ml), while the best antileishmanial activity exhibited compound **128**, which was active at 0.79 μg/ml.

Novel 7-phenyl-quinolines **130–133** (Fig. 18) were evaluated against bloodstream forms of *T. cruzi*. Nine quinolines were more effective against amastigotes than benznidazole (EC₅₀ = 2.7 μM) and they showed EC₅₀ values ranging from 0.6 to 0.1 μM (Nefertiti et al. 2018). All examined quinolines were highly active *in vitro* against African trypanosomes, showing EC₅₀ values ≤0.25 μM. The most potent *in vitro* candidates **130** were tested in *in vivo* models of *T. b. rhodesiense* infection, and showed a more than 98% reduction of the parasitemia and curing a half number of infected mice, after 3 doses of 40 mg/kg intraperitoneal administration. However, derivative **131** was even more efficacious, showed complete reduction of the parasitemia and cured a half of infected mice.

A series of sulfone derivatives were tested for their *in vitro* activity against *T. brucei*. From 22 examined compounds, derivatives obtained from 8-aminoquinoline **132** and **133** show the

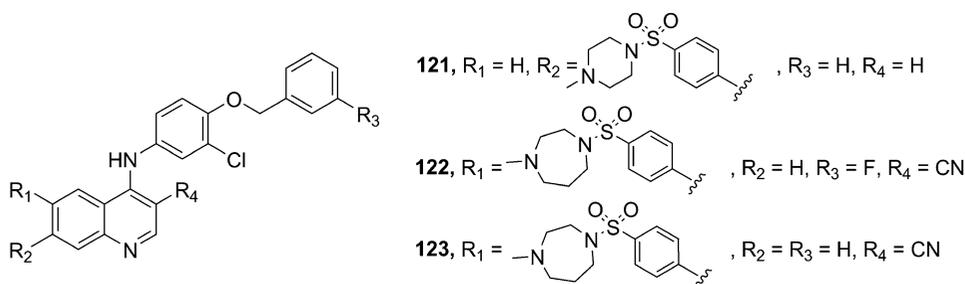


Fig. 17 4-Aminoquinolines with antitrypanosomal activity

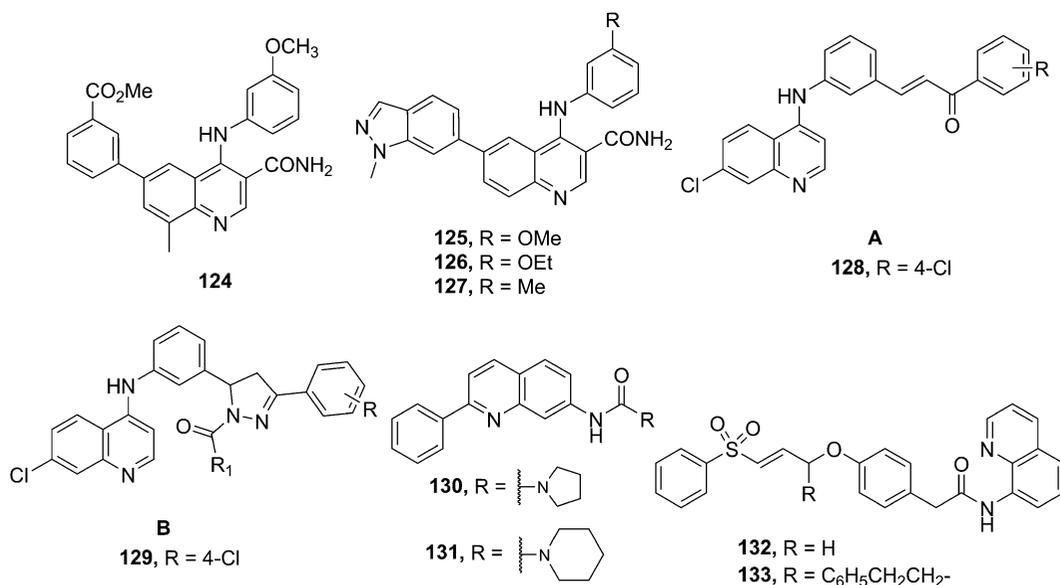


Fig. 18 Quinoline derivatives as antitrypanosomal agents

best inhibitory activity $IC_{50} = 0.76 \mu\text{M}$ (Zhang et al. 2018). However, during in vivo evaluation against *T. brucei*, when infected mice were treated for four consecutive days intraperitoneally, derivative **132** did not show activity, and the infected mice were positive for parasites 24 h post-treatment. However, second 8-quinoline derivative **133** was able to completely inactivate rhodesain, a cysteine protease essential for parasite survival and infectivity (Ettari et al. 2013), at $20 \mu\text{M}$ after 1 h of incubation with estimated IC_{50} value of 800 nM.

Pursuit for efficacious clinical therapy based on quinoline-based compounds still continues. Although there are increasing numbers of reports on trypanosoma growth inhibitors, and identification of potential targets, the mechanism of action (s) remain unknown.

7 Conclusion

As emergence and spread of the resistance to existing therapeutics are inevitable the novel advanced drugs and strategies for treatments of numerous infective diseases, affecting tens of millions of people worldwide, are continuously

being demanded. Among large number of various pharmacophores used for development of effective antimicrobials, quinoline/quinolone scaffold can be considered a privileged structure since it displays wide range of bioactivities. The diversity of synthesized quinoline/quinolone derivatives provides high and selective activity through different modes of action and low cytotoxicity on human cells.

The drugs based on quinoline/quinolone structure are well-known and in use for over half a century in the treatment of bacterial, parasitic, and viral infections. Besides, in the last two decades quinoline/quinolone derivatives emerged as efficient anti-virulence agents targeting bacterial and fungal virulence factors, including formation of biofilms, characteristic of chronic infections. Of particular importance is the ability of certain quinoline-based compounds to target both bacteria and fungi, either their planktonic or biofilm life form, suggesting that these compounds are promising chemical matter for the development of therapeutics against mixed-species and/or biofilm-associated infections. Quinoline/quinolone derivatives exhibit strong activity by themselves and can increase efficacy of commonly used drugs when applied together. Recently, a

new promising approach was employed in the development of more successful quinoline-derived therapeutics based on the synthesis of hybrid compounds that contain two or more pharmacophores affecting different targets and/or employing distinct mechanisms of action. Hence, there are still plenty of possibilities for development of novel more effective antimicrobials.

The high potency of quinoline/quinolone derivatives is demonstrated by their activities in nanomolar and low micromolar concentrations against wide range of infective agents including resistant strains. The importance of quinoline/quinolone as a scaffold for drugs' development is emphasized by recently granted accelerated approval for clinical use to a new quinoline-based antibiotic developed to fight MDR mycobacteria.

Therefore, there are more than enough reasons to continue quest for efficacious clinical therapeutics based on quinoline/quinolone structures since they are still proving to be unprecedented source of much needed solutions for infections treatments.

Acknowledgments This study has been funded by the Ministry of Education, Science and Technological Development, Republic of Serbia (Grants No. 173048 and No.172008).

References

- Abouelhassan Y, Garrison AT, Burch GM, Wong W, Norwood VM, Huigens RW 3rd (2014) Discovery of quinoline small molecules with potent dispersal activity against methicillin-resistant *Staphylococcus aureus* and *Staphylococcus epidermidis* biofilms using a scaffold hopping strategy. *Bioorg Med Chem Lett* 24 (21):5076–5080. <https://doi.org/10.1016/j.bmcl.2014.09.009>
- Abouelhassan Y, Garrison AT, Bai F, Norwood VM, Nguyen MT, Jin S, Huigens RW 3rd (2015) A phytochemical-halogenated Quinoline combination therapy strategy for the treatment of pathogenic bacteria. *ChemMedChem* 10(7):1157–1162. <https://doi.org/10.1002/cmdc.201500179>
- Al-Bari MA (2015) Chloroquine analogues in drug discovery: new directions of uses, mechanisms of actions and toxic manifestations from malaria to multifarious diseases. *J Antimicrob Chemother* 70(6):1608–1621. <https://doi.org/10.1093/jac/dkv018>
- Albuquerque P, Casadevall A (2012) Quorum sensing in fungi—a review. *Med Mycol* 50(4):337–345. <https://doi.org/10.3109/13693786.2011.652201>
- Aleksić I, Šegan S, Andrić F, Zlatović M, Moric I, Opsenica DM, Senerovic L (2017) Long-chain 4-Aminoquinolines as quorum sensing inhibitors in *Serratia marcescens* and *Pseudomonas aeruginosa*. *ACS Chem Biol* 12(5):1425–1434. <https://doi.org/10.1021/acscchembio.6b01149>
- Almandil NB, Taha M, Rahim F, Wadood A, Imran S, Alqahtani MA, Bamarouf YA, Ibrahim M, Mosaddik A, Gollapalli M (2019) Synthesis of novel quinoline-based thiazadiazole, evaluation of their antileishmanial potential and molecular docking studies. *Bioorg Chem* 85:109–116. <https://doi.org/10.1016/j.bioorg.2018.12.025>
- Anderson RJ, Groundwater PW, Todd A, Worsley A (2012) Antibacterial agents: chemistry, mode of action, mechanisms of resistance and clinical applications. Wiley, Chichester
- Andries K, Verhasselt P, Guillemont J, Gohlmann HW, Neefs JM, Winkler H, Van Gestel J, Timmerman P, Zhu M, Lee E, Williams P, de Chaffoy D, Huitric E, Hoffner S, Cambau E, Truffot-Pernot C, Lounis N, Jarlier V (2005) A diarylquinoline drug active on the ATP synthase of *Mycobacterium tuberculosis*. *Science* 307(5707):223–227. <https://doi.org/10.1126/science.1106753>
- Antinarelli LM, Dias RM, Souza IO, Lima WP, Gameiro J, da Silva AD, Coimbra ES (2015) 4-Aminoquinoline derivatives as potential antileishmanial agents. *Chem Biol Drug Des* 86(4):704–714. <https://doi.org/10.1111/cbdd.12540>
- Antinarelli LMR, de Oliveira Souza I, Zabala CPV, Gameiro J, Britta EA, Nakamura CV, Lima WP, da Silva AD, Coimbra ES (2018) Antileishmanial activity of a 4-hydrazinoquinoline derivative: induction of autophagy and apoptosis-related processes and effectiveness in experimental cutaneous leishmaniasis. *Exp Parasitol* 195:78–86
- Baragana B, Hallyburton I, Lee MC, Norcross NR, Grimaldi R, Otto TD, Proto WR, Blagborough AM, Meister S, Wirjanata G, Ruecker A, Upton LM, Abraham TS, Almeida MJ, Pradhan A, Porzelle A, Luksch T, Martinez MS, Luksch T, Bolscher JM, Woodland A, Norval S, Zuccotto F, Thomas J, Simeons F, Stojanovski L, Osuna-Cabello M, Brock PM, Churcher TS, Sala KA, Zakutansky SE, Jimenez-Diaz MB, Sanz LM, Riley J, Basak R, Campbell M, Avery VM, Sauerwein RW, Dechering KJ, Noviyanti R, Campo B, Frearson JA, Angulo-Barturen I, Ferrer-Bazaga S, Gamo FJ, Wyatt PG, Leroy D, Siegl P, Delves MJ, Kyle DE, Wittlin S, Marfurt J, Price RN, Sinden RE, Winzeler EA, Charman SA, Bebrevska L, Gray DW, Campbell S, Fairlamb AH, Willis PA, Rayner JC, Fidock DA, Read KD, Gilbert IH (2015) A novel multiple-stage

- antimalarial agent that inhibits protein synthesis. *Nature* 522(7556):315–320. <https://doi.org/10.1038/nature14451>
- Barbosa-Lima G, Moraes AM, Araujo ADS, da Silva ET, de Freitas CS, Vieira YR, Marttorelli A, Neto JC, Bozza PT, de Souza MVN, Souza TML (2017) 2,8-bis(trifluoromethyl)quinoline analogs show improved anti-Zika virus activity, compared to mefloquine. *Eur J Med Chem* 127:334–340. <https://doi.org/10.1016/j.ejmech.2016.12.058>
- Barnett DS, Guy RK (2014) Antimalarials in development in 2014. *Chem Rev* 114(22):11221–11241. <https://doi.org/10.1021/cr500543f>
- Basak A, Abouelhassan Y, Huigens RW 3rd (2015) Halogenated quinolines discovered through reductive amination with potent eradication activities against MRSA, MRSE and VRE biofilms. *Org Biomol Chem* 13(41):10290–10294. <https://doi.org/10.1039/c5ob01883h>
- Basak A, Abouelhassan Y, Norwood VM, Bai F, Nguyen MT, Jin S, Huigens RW 3rd (2016) Synthetically tuning the 2-position of halogenated Quinolines: optimizing antibacterial and biofilm eradication activities via alkylation and reductive amination pathways. *Chemistry* 22(27):9181–9189. <https://doi.org/10.1002/chem.201600926>
- Basak A, Abouelhassan Y, Kim YS, Norwood VM, Jin S, Huigens RW 3rd (2018) Halogenated quinolines bearing polar functionality at the 2-position: identification of new antibacterial agents with enhanced activity against *Staphylococcus epidermidis*. *Eur J Med Chem* 155:705–713. <https://doi.org/10.1016/j.ejmech.2018.06.045>
- Basilico N, Parapini S, Sparatore A, Romeo S, Misiano P, Vivas L, Yardley V, Croft SL, Habluetzel A, Lucantoni L, Renia L, Russell B, Suwanarusk R, Nosten F, Dondio G, Bigogno C, Jabes D, Taramelli D (2017) In vivo and in vitro activities and ADME-Tox profile of a quinolizidine-modified 4-Aminoquinoline: a potent anti-*P. falciparum* and anti-*P. vivax* blood-stage antimalarial. *Molecules* 22(12). <https://doi.org/10.3390/molecules22122102>
- Ben Yaakov D, Shadkchan Y, Albert N, Kontoyiannis DP, Osherov N (2017) The quinoline bromoquinol exhibits broad-spectrum antifungal activity and induces oxidative stress and apoptosis in *aspergillus fumigatus*. *J Antimicrob Chemother* 72(8):2263–2272. <https://doi.org/10.1093/jac/dkx117>
- Bernier SP, Surette MG (2013) Concentration-dependent activity of antibiotics in natural environments. *Front Microbiol* 4:20. <https://doi.org/10.3389/fmicb.2013.00020>
- Bisacchi GS (2015) Origins of the quinolone class of antibacterials: an expanded “discovery story”. *J Med Chem* 58(12):4874–4882. <https://doi.org/10.1021/jm501881c>
- Boudhar A, Ng XW, Loh CY, Chia WN, Tan ZM, Nosten F, Dymock BW, Tan KS (2016) Overcoming chloroquine resistance in malaria: design, synthesis, and structure-activity relationships of novel hybrid compounds. *Antimicrob Agents Chemother* 60(5):3076–3089. <https://doi.org/10.1128/AAC.02476-15>
- Capela R, Magalhaes J, Miranda D, Machado M, Sanches-Vaz M, Albuquerque IS, Sharma M, Gut J, Rosenthal PJ, Frade R, Perry MJ, Moreira R, Prudencio M, Lopes F (2018) Endoperoxide-8-aminoquinoline hybrids as dual-stage antimalarial agents with enhanced metabolic stability. *Eur J Med Chem* 149:69–78. <https://doi.org/10.1016/j.ejmech.2018.02.048>
- Cecchetti V, Parolin C, Moro S, Pecere T, Filipponi E, Calistri A, Tabarrini O, Gatto B, Palumbo M, Fravolini A, Palu G (2000) 6-Aminoquinolones as new potential anti-HIV agents. *J Med Chem* 43(20):3799–3802
- Chander S, Ashok P, Zheng YT, Wang P, Raja KS, Taneja A, Murugesan S (2016) Design, synthesis and in-vitro evaluation of novel tetrahydroquinoline carbamates as HIV-1 RT inhibitor and their antifungal activity. *Bioorg Chem* 64:66–73. <https://doi.org/10.1016/j.bioorg.2015.12.005>
- Chanquia SN, Larregui F, Puente V, Labriola C, Lombardo E, Garcia Linares G (2019) Synthesis and biological evaluation of new quinoline derivatives as antileishmanial and antitrypanosomal agents. *Bioorg Chem* 83:526–534. <https://doi.org/10.1016/j.bioorg.2018.10.053>
- Chico RM, Chandramohan D (2011) Azithromycin plus chloroquine: combination therapy for protection against malaria and sexually transmitted infections in pregnancy. *Expert Opin Drug Metab Toxicol* 7(9):1153–1167. <https://doi.org/10.1517/17425255.2011.598506>
- Chu XM, Wang C, Liu W, Liang LL, Gong KK, Zhao CY, Sun KL (2019) Quinoline and quinolone dimers and their biological activities: an overview. *Eur J Med Chem* 161:101–117. <https://doi.org/10.1016/j.ejmech.2018.10.035>
- Costa-Orlandi CB, Sardi JCO, Pitangui NS, de Oliveira HC, Scorzoni L, Galeane MC, Medina-Alarcon KP, Melo W, Marcelino MY, Braz JD, Fusco-Almeida AM, Mendes-Giannini MJS (2017) Fungal biofilms and Polymicrobial diseases. *J Fungi* 3(2):pii: E22. <https://doi.org/10.3390/jof3020022>
- de Azambuja Carvalho PH, Duval AR, Manzolli Leite FR, Nedel F, Cunico W, Lund RG (2016) (7-Chloroquinolin-4-yl)arylhydrazones: *Candida albicans* enzymatic repression and cytotoxicity evaluation, part 2. *J Enzyme Inhib Med Chem* 31(1):126–131. <https://doi.org/10.3109/14756366.2015.1010527>
- de Souza IO, Schrekker CM, Lopes W, Orru RV, Hranjec M, Perin N, Machado M, Oliveira LF, Donato RK, Stefani V, Fuentefria AM, Schrekker HS (2016) Bifunctional fluorescent benzimidazo[1,2- α]quinolines for *Candida* spp. biofilm detection and biocidal activity. *J Photochem Photobiol B* 163:319–326. <https://doi.org/10.1016/j.jphotobiol.2016.08.037>

- Defoidt T (2018) Quorum-sensing systems as targets for Antivirulence therapy. *Trends Microbiol* 26 (4):313–328. <https://doi.org/10.1016/j.tim.2017.10.005>
- Delattin N, Bardiou D, Marchand A, Chaltin P, De Brucker K, Cammue BP, Thevissen K (2012) Identification of fungicidal 2,6-disubstituted quinolines with activity against *Candida* biofilms. *Molecules* 17 (10):12243–12251. <https://doi.org/10.3390/molecules171012243>
- Delvecchio R, Higa LM, Pezzuto P, Valadao AL, Garcez PP, Monteiro FL, Loiola EC, Dias AA, Silva FJ, Aliota MT, Caine EA, Osorio JE, Bellio M, O'Connor DH, Rehen S, de Aguiar RS, Savarino A, Campanati L, Tanuri A (2016) Chloroquine, an endocytosis blocking agent, inhibits Zika virus infection in different cell models. *Viruses* 8(12). <https://doi.org/10.3390/v8120322>
- Devine W, Woodring JL, Swaminathan U, Amata E, Patel G, Erath J, Roncal NE, Lee PJ, Leed SE, Rodriguez A, Mensa-Wilmot K, Sciotti RJ, Pollastri MP (2015) Protozoan parasite growth inhibitors discovered by cross-screening yield potent scaffolds for Lead discovery. *J Med Chem* 58(14):5522–5537. <https://doi.org/10.1021/acs.jmedchem.5b00515>
- Deziel E, Lepine F, Milot S, He J, Mindrinos MN, Tompkins RG, Rahme LG (2004) Analysis of *Pseudomonas aeruginosa* 4-hydroxy-2-alkylquinolines (HAQs) reveals a role for 4-hydroxy-2-heptylquinoline in cell-to-cell communication. *Proc Natl Acad Sci U S A* 101(5):1339–1344. <https://doi.org/10.1073/pnas.0307694100>
- Diggle SP, Lumjiaktase P, Dipilato F, Winzer K, Kunakorn M, Barrett DA, Chhabra SR, Camara M, Williams P (2006) Functional genetic analysis reveals a 2-Alkyl-4-quinolone signaling system in the human pathogen *Burkholderia pseudomallei* and related bacteria. *Chem Biol* 13(7):701–710. <https://doi.org/10.1016/j.chembiol.2006.05.006>
- Dola VR, Soni A, Agarwal P, Ahmad H, Raju KS, Rashid M, Wahajuddin M, Srivastava K, Haq W, Dwivedi AK, Puri SK, Katti SB (2017) Synthesis and evaluation of Chirally defined side chain variants of 7-Chloro-4-Aminoquinoline to overcome drug resistance in malaria chemotherapy. *Antimicrob Agents Chemother* 61(3). <https://doi.org/10.1128/AAC.01152-16>
- Dolan N, Gavin DP, Eshwika A, Kavanagh K, McGinley J, Stephens JC (2016) Synthesis, antibacterial and anti-MRSA activity, in vivo toxicity and a structure-activity relationship study of a quinoline thiourea. *Bioorg Med Chem Lett* 26(2):630–635. <https://doi.org/10.1016/j.bmcl.2015.11.058>
- Duval AR, Carvalho PH, Soares MC, Gouvea DP, Siqueira GM, Lund RG, Cunico W (2011) 7-chloroquinolin-4-yl arylhydrazones derivatives: synthesis and antifungal activity. *Sci World J* 11:1489–1495. <https://doi.org/10.1100/tsw.2011.141>
- Egan TJ, Marques HM (1999) The role of haem in the activity of chloroquine and related antimalarial drugs. *Coord Chem Rev* 190–192:493–517
- El Shehry MF, Ghorab MM, Abbas SY, Fayed EA, Shedid SA, Ammar YA (2018) Quinolone derivatives bearing pyrazole moiety: synthesis and biological evaluation as possible antibacterial and antifungal agents. *Eur J Med Chem* 143:1463–1473. <https://doi.org/10.1016/j.ejmech.2017.10.046>
- Espinosa-Valdes MP, Borbolla-Alvarez S, Delgado-Espinosa AE, Sanchez-Tejeda JF, Ceron-Nava A, Quintana-Romero OJ, Ariza-Castolo A, Garcia-Del Rio DF, Loza-Mejia MA (2019) Synthesis, in silico, and in vitro evaluation of long chain alkyl amides from 2-Amino-4-quinolone derivatives as biofilm inhibitors. *Molecules* 24(2). <https://doi.org/10.3390/molecules24020327>
- Ettari R, Tamborini L, Angelo IC, Micale N, Pinto A, De Micheli C, Conti P (2013) Inhibition of rhodesain as a novel therapeutic modality for human African trypanosomiasis. *J Med Chem* 56(14):5637–5658. <https://doi.org/10.1021/jm301424d>
- Fernández-Piñar R, Cámara M, Dubern J-F, Ramos JL, Espinosa-Urgel M (2011) The *Pseudomonas aeruginosa* quinolone quorum sensing signal alters the multicellular behaviour of *Pseudomonas putida* KT2440. *Res Microbiol* 162(8):773–781. <https://doi.org/10.1016/j.resmic.2011.06.013>
- Gama N, Kumar K, Ekengard E, Haukka M, Darkwa J, Nordlander E, Meyer D (2016) Gold(I) complex of 1,1'-bis(diphenylphosphino) ferrocene-quinoline conjugate: a virostatic agent against HIV-1. *Biomaterials* 29 (3):389–397. <https://doi.org/10.1007/s10534-016-9921-9>
- Garrison AT, Abouelhassan Y, Yang H, Yousof HH, Nguyen TJ, Huigens Iii RW (2017) Microwave-enhanced Friedlander synthesis for the rapid assembly of halogenated quinolines with antibacterial and biofilm eradication activities against drug resistant and tolerant bacteria. *Med Chem Commun* 8(4):720–724. <https://doi.org/10.1039/c6md000381h>
- Gould MK, de Koning HP (2011) Cyclic-nucleotide signalling in protozoa. *FEMS Microbiol Rev* 35 (3):515–541. <https://doi.org/10.1111/j.1574-6976.2010.00262.x>
- Gualerzi CO, Brandi L, Fabbretti A, Pon CL (2013) Antibiotics: targets, mechanisms and resistance. Wiley-VCH, Weinheim. <https://doi.org/10.1002/9783527659685>
- Hajimadhi Z, Zabihollahi R, Aghasadeghi MR, Hosseini Ashtiani S, Zargh A (2016) Novel quinolone-3-carboxylic acid derivatives as anti-HIV-1 agents: design, synthesis, and biological activities. *Med Chem Res* 25:1861–1876
- Hall-Stoodley L, Costerton JW, Stoodley P (2004) Bacterial biofilms: from the natural environment to infectious diseases. *Nat Rev Microbiol* 2(2):95–108
- Hamama WS, Ibrahim ME, Gooda AA, Zoorob HH (2018) Efficient synthesis, antimicrobial, antioxidant

- assessments and geometric optimization calculations of azoles- incorporating Quinoline moiety. *J Heterocyclic Chem* 55(11):2623–2634
- He QQ, Zhang X, Yang LM, Zheng YT, Chen F (2013) Synthesis and biological evaluation of 5-fluoroquinolone-3-carboxylic acids as potential HIV-1 integrase inhibitors. *J Enzyme Inhib Med Chem* 28(4):671–676. <https://doi.org/10.3109/14756366.2012.668540>
- Heeb S, Fletcher MP, Chhabra SR, Diggle SP, Williams P, Camara M (2011) Quinolones: from antibiotics to autoinducers. *FEMS Microbiol Rev* 35(2):247–274. <https://doi.org/10.1111/j.1574-6976.2010.00247.x>
- Hucke O, Coulombe R, Bonneau P, Bertrand-Laperle M, Brochu C, Gillard J, Joly MA, Landry S, Lepage O, Llinas-Brunet M, Pesant M, Poirier M, Poirier M, McKercher G, Marquis M, Kukolj G, Beaulieu PL, Stammers TA (2014) Molecular dynamics simulations and structure-based rational design lead to allosteric HCV NS5B polymerase thumb pocket 2 inhibitor with picomolar cellular replicon potency. *J Med Chem* 57(5):1932–1943. <https://doi.org/10.1021/jm4004522>
- Huigens RW 3rd (2018) The path to new halogenated Quinolines with enhanced activities against *Staphylococcus epidermidis*. *Microbiol Insights* 11:1178636118808532. <https://doi.org/10.1177/1178636118808532>
- Ilangovan A, Fletcher M, Rampioni G, Pustelny C, Rumbaugh K, Heeb S, Cámara M, Truman A, Chhabra SR, Emsley J, Williams P (2013) Structural basis for native agonist and synthetic inhibitor recognition by the *Pseudomonas aeruginosa* quorum sensing regulator PqsR (MvFR). *PLoS Pathog* 9(7):e1003508. <https://doi.org/10.1371/journal.ppat.1003508>
- Irfan M, Aneja B, Yadava U, Khan SI, Manzoor N, Daniliuc CG, Abid M (2015) Synthesis, QSAR and anticandidal evaluation of 1,2,3-triazoles derived from naturally bioactive scaffolds. *Eur J Med Chem* 93:246–254. <https://doi.org/10.1016/j.ejmech.2015.02.007>
- Irfan M, Alam S, Manzoor N, Abid M (2017) Effect of quinoline based 1,2,3-triazole and its structural analogues on growth and virulence attributes of *Candida albicans*. *PLoS One* 12(4):e0175710. <https://doi.org/10.1371/journal.pone.0175710>
- Jentsch NG, Hart AP, Hume JD, Sun J, McNeely KA, Lama C, Pigza JA, Donahue MG, Kessl JJ (2018) Synthesis and evaluation of aryl Quinolines as HIV-1 integrase Multimerization inhibitors. *ACS Med Chem Lett* 9(10):1007–1012. <https://doi.org/10.1021/acsmchemlett.8b00269>
- Katiyar S, Kufareva I, Behera R, Thomas SM, Ogata Y, Pollastri M, Abagyan R, Mensa-Wilmot K (2013) Lapatinib-binding protein kinases in the African trypanosome: identification of cellular targets for kinase-directed chemical scaffolds. *PLoS One* 8(2):e56150. <https://doi.org/10.1371/journal.pone.0056150>
- Khan FAK, Kaduskar RN, Patil R, Patil RH, Ansari SA, Alkahtani HM, Almezhiia AA, Shinde DB, Sangshetti JN (2019) Synthesis, biological evaluations and computational studies of N-(3-(2-(7-Chloroquinolin-2-yl)vinyl) benzylidene)anilines as fungal biofilm inhibitors. *Bioorg Med Chem Lett* 29(4):623–630. <https://doi.org/10.1016/j.bmcl.2018.12.046>
- Konstantinovic J, Videnovic M, Srbljanovic J, Djurkovic-Djakovic O, Bogojevic K, Sciotti R, Solaja B (2017) Antimalarials with benzothioephene moieties as aminoquinoline partners. *Molecules* 22(3). <https://doi.org/10.3390/molecules22030343>
- Konstantinovic J, Videnovic M, Orsini S, Bogojevic K, D'Alessandro S, Scaccabarozzi D, Terzic Jovanovic N, Gradoni L, Basilico N, Solaja BA (2018) Novel Aminoquinoline derivatives significantly reduce parasite load in Leishmania infantum infected mice. *ACS Med Chem Lett* 9(7):629–634. <https://doi.org/10.1021/acsmchemlett.8b00053>
- Korotchenko V, Sathunuru R, Gerena L, Caridha D, Li Q, Kreishman-Deitrick M, Smith PL, Lin AJ (2015) Antimalarial activity of 4-amidinquinoline and 10-amidinobenzonaphthyridine derivatives. *J Med Chem* 58(8):3411–3431. <https://doi.org/10.1021/jm501809x>
- Krauss J, Muller C, Kiessling J, Richter S, Staudacher V, Bracher F (2014) Synthesis and biological evaluation of novel N-alkyl tetra- and decahydroisoquinolines: novel antifungals that target ergosterol biosynthesis. *Arch Pharm* 347(4):283–290. <https://doi.org/10.1002/ardp.201300338>
- Krauss J, Hornacek M, Muller C, Staudacher V, Stadler M, Bracher F (2015) Synthesis and antifungal evaluation of novel N-alkyl tetra- and perhydroquinoline derivatives. *Sci Pharm* 83(1):1–14. <https://doi.org/10.3797/scipharm.1409-13>
- Kumar DV, Rai R, Brameld KA, Riggs J, Somoza JR, Rajagopalan R, Janc JW, Xia YM, Ton TL, Hu H, Lehoux I, Ho JD, Young WB, Hart B, Green MJ (2012) 3-heterocyclyl quinolone inhibitors of the HCV NS5B polymerase. *Bioorg Med Chem Lett* 22(1):300–304. <https://doi.org/10.1016/j.bmcl.2011.11.013>
- Leon B, Haeckl FP, Linington RG (2015) Optimized quinoline amino alcohols as disruptors and dispersal agents of *Vibrio cholerae* biofilms. *Org Biomol Chem* 13(31):8495–8499. <https://doi.org/10.1039/c5ob01134e>
- Leven M, Held J, Duffy S, Alves Avelar LA, Meister S, Delves M, Plouffe D, Kuna K, Tschan S, Avery VM, Winzeler EA, Mordmuller B, Kurz T (2019) 8-aminoquinolines with an aminoxyalkyl side chain exert in vitro dual-stage antiparasitic activity. *ChemMedChem* 14(4):501–511. <https://doi.org/10.1002/cmdc.201800691>
- Loregian A, Mercorelli B, Muratore G, Sinigalia E, Pagni S, Massari S, Gribaudo G, Gatto B, Palumbo M, Tabarrini O, Cecchetti V, Palu G (2010) The 6-aminoquinolone WC5 inhibits human cytomegalovirus replication at an early stage by interfering with the transactivating activity of viral immediate-early

- 2 protein. *Antimicrob Agents Chemother* 54 (5):1930–1940. <https://doi.org/10.1128/AAC.01730-09>
- Lu C, Kirsch B, Zimmer C, de Jong JC, Henn C, Maurer CK, Musken M, Haussler S, Steinbach A, Hartmann RW (2012) Discovery of antagonists of PqsR, a key player in 2-alkyl-4-quinolone-dependent quorum sensing in *Pseudomonas aeruginosa*. *Chem Biol* 19 (3):381–390. <https://doi.org/10.1016/j.chembiol.2012.01.015>
- Lu C, Kirsch B, Maurer CK, de Jong JC, Braunschauen A, Steinbach A, Hartmann RW (2014a) Optimization of anti-virulence PqsR antagonists regarding aqueous solubility and biological properties resulting in new insights in structure–activity relationships. *Eur J Med Chem* 79:173–183. <https://doi.org/10.1016/j.ejmech.2014.04.016>
- Lu C, Maurer CK, Kirsch B, Steinbach A, Hartmann RW (2014b) Overcoming the unexpected functional inversion of a PqsR antagonist in *Pseudomonas aeruginosa*: an in vivo potent antivirulence agent targeting pqs quorum sensing. *Angew Chem Int Ed* 53 (4):1109–1112. <https://doi.org/10.1002/anie.201307547>
- Luthra P, Liang J, Pietzsch CA, Khadka S, Edwards MR, Wei S, De S, Posner B, Bukreyev A, Ready JM, Basler CF (2018) A high throughput screen identifies benzoquinoline compounds as inhibitors of Ebola virus replication. *Antivir Res* 150:193–201. <https://doi.org/10.1016/j.antiviral.2017.12.019>
- Manfroni G, Cannalire R, Barreca ML, Kaushik-Basu N, Leyssen P, Winquist J, Iraci N, Manvar D, Paeshuysse J, Guhamazumder R, Basu A, Sabatini S, Tabarrini O, Danielson UH, Neyts J, Cecchetti V (2014) The versatile nature of the 6-aminoquinolone scaffold: identification of submicromolar hepatitis C virus NS5B inhibitors. *J Med Chem* 57 (5):1952–1963. <https://doi.org/10.1021/jm401362f>
- Manohar S, Khan SI, Rawat DS (2013) 4-aminoquinoline-triazine-based hybrids with improved in vitro antimalarial activity against CQ-sensitive and CQ-resistant strains of *Plasmodium falciparum*. *Chem Biol Drug Des* 81(5):625–630. <https://doi.org/10.1111/cbdd.12108>
- Mao TQ, He QQ, Wan ZY, Chen WX, Chen FE, Tang GF, De Clercq E, Daelemans D, Pannecouque C (2015) Anti-HIV diarylpyrimidine-quinolone hybrids and their mode of action. *Bioorg Med Chem* 23 (13):3860–3868. <https://doi.org/10.1016/j.bmc.2015.03.037>
- Martínez A, Deregnacourt C, Sinou V, Latour C, Roy D, Schrével J, Sánchez-Delgado RA (2017) Synthesis of an organo-ruthenium aminoquinoline-trioxane hybrid and evaluation of its activity against plasmodium falciparum and its toxicity toward normal mammalian cells. *Med Chem Res* 26(2):473–483
- Massoud AA, Langer V, Gohar YM, Abu-Youssef MA, Janis J, Lindberg G, Hansson K, Ohlstrom L (2013) Effects of different substituents on the crystal structures and antimicrobial activities of six Ag (I) quinoline compounds. *Inorg Chem* 52 (7):4046–4060. <https://doi.org/10.1021/ic400081v>
- Mercorelli B, Luginini A, Muratore G, Massari S, Terlizzi ME, Tabarrini O, Gribaudo G, Palu G, Loregian A (2014) The 6-Aminoquinolone WC5 inhibits different functions of the immediate-early 2 (IE2) protein of human cytomegalovirus that are essential for viral replication. *Antimicrob Agents Chemother* 58 (11):6615–6626. <https://doi.org/10.1128/AAC.03309-14>
- Miquel S, Lagrèfeuille R, Souweine B, Forestier C (2016) Anti-biofilm activity as a health issue. *Front Microbiol* 7:592–592. <https://doi.org/10.3389/fmicb.2016.00592>
- Montoya A, Quiroga J, Abonia R, Derita M, Sortino M, Ornelas A, Zacchino S, Insuasty B (2016) Hybrid molecules containing a 7-Chloro-4-aminoquinoline nucleus and a substituted 2-pyrazoline with antiproliferative and antifungal activity. *Molecules* 21(8). <https://doi.org/10.3390/molecules21080969>
- Moradali MF, Ghods S, Rehm BH (2017) *Pseudomonas aeruginosa* lifestyle: a paradigm for adaptation, survival, and persistence. *Front Cell Infect Microbiol* 7:39. <https://doi.org/10.3389/fcimb.2017.00039>
- Musiol R, Serda M, Hensel-Bielowka S, Polanski J (2010) Quinoline-based antifungals. *Curr Med Chem* 17 (18):1960–1973
- Nefertiti ASG, Batista MM, Da Silva PB, Batista DGJ, Da Silva CF, Peres RB, Torres-Santos EC, Cunha-Junior EF, Holt E, Boykin DW, Brun R, Wenzler T, Soeiro MNC (2018) In vitro and in vivo studies of the Trypanocidal effect of novel Quinolines. *Antimicrob Agents Chemother* 62(2). <https://doi.org/10.1128/AAC.01936-17>
- Nikolić S, Ospenica DM, Filipović V, Dojčinović B, Arandelović S, Radulović S, Grgurić-Šipka S (2015) Strong *in vitro* cytotoxic potential of new ruthenium-cymene complexes. *Organometallics* 34 (14):3464–3473. <https://doi.org/10.1021/acs.organomet.5b00041>
- Nwaka S, Hudson A (2006) Innovative lead discovery strategies for tropical diseases. *Nat Rev Drug Discov* 5(11):941–955. <https://doi.org/10.1038/nrd2144>
- Ochiana SO, Bland ND, Settimo L, Campbell RK, Pollastri MP (2015) Repurposing human PDE4 inhibitors for neglected tropical diseases. Evaluation of analogs of the human PDE4 inhibitor GSK-256066 as inhibitors of PDEB1 of *trypanosoma brucei*. *Chem Biol Drug Des* 85(5):549–564. <https://doi.org/10.1111/cbdd.12443>
- Olliaro P, Mussano P (2003) Amodiaquine for treating malaria. *Cochrane Database Syst Rev* 2:CD000016. <https://doi.org/10.1002/14651858.CD000016>
- Ongarora DS, Strydom N, Wicht K, Njoroge M, Wiesner L, Egan TJ, Wittlin S, Jurva U, Masimirembwa CM, Chibale K (2015) Antimalarial benzoheterocyclic 4-aminoquinolines: structure-activity relationship, in vivo evaluation, mechanistic and bioactivation studies. *Bioorg Med Chem* 23 (17):5419–5432. <https://doi.org/10.1016/j.bmc.2015.07.051>

- Ospenica I, Filipovic V, Nuss JE, Gomba LM, Ospenica D, Burnett JC, Gussio R, Solaja BA, Bavari S (2012) The synthesis of 2,5-bis(4-amidinophenyl) thiophene derivatives providing submicromolar-range inhibition of the botulinum neurotoxin serotype A metalloprotease. *Eur J Med Chem* 53:374–379. <https://doi.org/10.1016/j.ejmech.2012.03.043>
- Ospenica IM, Verbic TZ, Tot M, Sciotti RJ, Pybus BS, Djurkovic-Djakovic O, Slavic K, Solaja BA (2015) Investigation into novel thiophene- and furan-based 4-amino-7-chloroquinolines afforded antimalarials that cure mice. *Bioorg Med Chem* 23(9):2176–2186. <https://doi.org/10.1016/j.bmc.2015.02.061>
- Parsons M, Worthey EA, Ward PN, Mottram JC (2005) Comparative analysis of the kinomes of three pathogenic trypanosomatids: *Leishmania* major, *Trypanosoma brucei* and *Trypanosoma cruzi*. *BMC Genomics* 6:127. <https://doi.org/10.1186/1471-2164-6-127>
- Patel G, Karver CE, Behera R, Guyett PJ, Sullenberger C, Edwards P, Roncal NE, Mensa-Wilmot K, Pollastri MP (2013) Kinase scaffold repurposing for neglected disease drug discovery: discovery of an efficacious, lapatinib-derived lead compound for trypanosomiasis. *J Med Chem* 56(10):3820–3832. <https://doi.org/10.1021/jm400349k>
- Phillips B, Cai R, Delaney W, Du Z, Ji M, Jin H, Lee J, Li J, Niedziela-Majka A, Mish M, Pyun HJ, Saugier J, Tirunagari N, Wang J, Yang H, Wu Q, Sheng C, Zonte C (2014) Highly potent HCV NS4B inhibitors with activity against multiple genotypes. *J Med Chem* 57(5):2161–2166. <https://doi.org/10.1021/jm401646w>
- Pierre F, O'Brien SE, Haddach M, Bourbon P, Schwaebé MK, Stefan E, Darjania L, Stansfield R, Ho C, Siddiqui-Jain A, Streiner N, Rice WG, Anderes K, Ryckman DM (2011) Novel potent pyrimido[4,5-c]quinoline inhibitors of protein kinase CK2: SAR and preliminary assessment of their analgesic and anti-viral properties. *Bioorg Med Chem Lett* 21(6):1687–1691. <https://doi.org/10.1016/j.bmcl.2011.01.091>
- Pippi B, Reginatto P, Machado G, Bergamo VZ, Lana DFD, Teixeira ML, Franco LL, Alves RJ, Andrade SF, Fuentefria AM (2017) Evaluation of 8-hydroxyquinoline derivatives as hits for antifungal drug design. *Med Mycol* 55(7):763–773. <https://doi.org/10.1093/mmy/myx003>
- Pippi B, Machado G, Bergamo VZ, Alves RJ, Andrade SF, Fuentefria AM (2018) Clioquinol is a promising preventive morphological switching compound in the treatment of *Candida* infections linked to the use of intrauterine devices. *J Med Microbiol* 67(11):1655–1663. <https://doi.org/10.1099/jmm.0.000850>
- Plantone D, Koudriavtseva T (2018) Current and future use of chloroquine and hydroxychloroquine in infectious, immune, neoplastic, and neurological diseases: a mini-review. *Clin Drug Investig* 38(8):653–671. <https://doi.org/10.1007/s40261-018-0656-y>
- Ramirez-Prada J, Robledo SM, Velez ID, Crespo MDP, Quiroga J, Abonia R, Montoya A, Svetaz L, Zacchino S, Insuasty B (2017) Synthesis of novel quinoline-based 4,5-dihydro-1H-pyrazoles as potential anticancer, antifungal, antibacterial and antiprotozoal agents. *Eur J Med Chem* 131:237–254. <https://doi.org/10.1016/j.ejmech.2017.03.016>
- Reen FJ, Mooij MJ, Holcombe LJ, McSweeney CM, McGlacken GP, Morrissey JP, O'Gara F (2011) The *Pseudomonas* quinolone signal (PQS), and its precursor HHQ, modulate interspecies and interkingdom behaviour. *FEMS Microbiol Ecol* 77(2):413–428. <https://doi.org/10.1111/j.1574-6941.2011.01121.x>
- Reen FJ, Clarke SL, Legendre C, McSweeney CM, Eccles KS, Lawrence SE, O'Gara F, McGlacken GP (2012) Structure–function analysis of the C-3 position in analogues of microbial behavioural modulators HHQ and PQS. *Org Biomol Chem* 10(44):8903–8910. <https://doi.org/10.1039/C2OB26823J>
- Reen FJ, Shanahan R, Cano R, O'Gara F, McGlacken GP (2015) A structure activity-relationship study of the bacterial signal molecule HHQ reveals swarming motility inhibition in *Bacillus atrophaeus*. *Org Biomol Chem* 13(19):5537–5541. <https://doi.org/10.1039/C5OB00315F>
- Salas PF, Herrmann C, Cawthray JF, Nimphius C, Kenkel A, Chen J, de Kock C, Smith PJ, Patrick BO, Adam MJ, Orvig C (2013) Structural characteristics of chloroquine-bridged ferrocenophane analogues of ferroquine may obviate malaria drug-resistance mechanisms. *J Med Chem* 56(4):1596–1613. <https://doi.org/10.1021/jm301422h>
- Sato M, Motomura T, Aramaki H, Matsuda T, Yamashita M, Ito Y, Kawakami H, Matsuzaki Y, Watanabe W, Yamataka K, Ikeda S, Kodama E, Matsuoka M, Shinkai H (2006) Novel HIV-1 integrase inhibitors derived from quinolone antibiotics. *J Med Chem* 49(5):1506–1508. <https://doi.org/10.1021/jm0600139>
- Savarino A, Shytaj IL (2015) Chloroquine and beyond: exploring anti-rheumatic drugs to reduce immune hyperactivation in HIV/AIDS. *Retrovirology* 12:51. <https://doi.org/10.1186/s12977-015-0178-0>
- Savarino A, Boelaert JR, Cassone A, Majori G, Cauda R (2003) Effects of chloroquine on viral infections: an old drug against today's diseases? *Lancet Infect Dis* 3(11):722–727
- Schlitzer M (2007) Malaria chemotherapeutics part I: history of antimalarial drug development, currently used therapeutics, and drugs in clinical development. *ChemMedChem* 2(7):944–986. <https://doi.org/10.1002/cmdc.200600240>
- Scola PM, Sun LQ, Wang AX, Chen J, Sin N, Venables BL, Sit SY, Chen Y, Cocuzza A, Bilder DM, D'Andrea SV, Zheng B, Hewawasam P, Tu Y, Friborg J, Falk P,

- Hernandez D, Levine S, Chen C, Yu F, Sheaffer AK, Zhai G, Barry D, Knipe JO, Han YH, Schartman R, Donoso M, Masure K, Sinz MW, Zvyaga T, Good AC, Rajamani R, Kish K, Tredup J, Klei HE, Gao Q, Mueller L, Colonna RJ, Grasela DM, Adams SP, Loy J, Levesque PC, Sun H, Shi H, Sun L, Warner W, Li D, Zhu J, Meanwell NA, McPhee F (2014a) The discovery of asunaprevir (BMS-650032), an orally efficacious NS3 protease inhibitor for the treatment of hepatitis C virus infection. *J Med Chem* 57(5):1730–1752. <https://doi.org/10.1021/jm500297k>
- Scola PM, Wang AX, Good AC, Sun LQ, Combrink KD, Campbell JA, Chen J, Tu Y, Sin N, Venables BL, Sit SY, Chen Y, Cocuzza A, Bilder DM, D'Andrea S, Zheng B, Hewawasam P, Ding M, Thuring J, Li J, Hernandez D, Yu F, Falk P, Zhai G, Sheaffer AK, Chen C, Lee MS, Barry D, Knipe JO, Li W, Han YH, Jenkins S, Gesenberg C, Gao Q, Sinz MW, Santone KS, Zvyaga T, Rajamani R, Klei HE, Colonna RJ, Grasela DM, Hughes E, Chien C, Adams S, Levesque PC, Li D, Zhu J, Meanwell NA, McPhee F (2014b) Discovery and early clinical evaluation of BMS-605339, a potent and orally efficacious tripeptidic acylsulfonamide NS3 protease inhibitor for the treatment of hepatitis C virus infection. *J Med Chem* 57(5):1708–1729. <https://doi.org/10.1021/jm401840s>
- Shaikh SKJ, Kamble RR, Somagond SM, Devarajegowda HC, Dixit SR, Joshi SD (2017) Tetrazolylmethyl quinolines: design, docking studies, synthesis, anticancer and antifungal analyses. *Eur J Med Chem* 128:258–273. <https://doi.org/10.1016/j.ejmech.2017.01.043>
- Shang XF, Morris-Natschke SL, Liu YQ, Guo X, Xu XS, Goto M, Li JC, Yang GZ, Lee KH (2018) Biologically active quinoline and quinazoline alkaloids part I. *Med Res Rev* 38(3):775–828. <https://doi.org/10.1002/med.21466>
- Shinde RB, Raut JS, Chauhan NM, Karuppaiyl SM (2013) Chloroquine sensitizes biofilms of *Candida albicans* to antifungal azoles. *Braz J Infect Dis* 17(4):395–400. <https://doi.org/10.1016/j.bjid.2012.11.002>
- Solaja BA, Opsenica D, Smith KS, Milhous WK, Terzic N, Opsenica I, Burnett JC, Nuss J, Gussio R, Bavari S (2008) Novel 4-aminoquinolines active against chloroquine-resistant and sensitive *P. falciparum* strains that also inhibit botulinum serotype A. *J Med Chem* 51(15):4388–4391. <https://doi.org/10.1021/jm800737y>
- Soukariéh F, Vico Oton E, Dubern J-F, Gomes J, Halliday N, de Pilar CM, Ramírez-Prada J, Insuasty B, Abonia R, Quiroga J, Heeb S, Williams P, Stocks MJ, Cámara M (2018) In silico and in vitro-guided identification of inhibitors of Alkylquinolone-dependent quorum sensing in *Pseudomonas aeruginosa*. *Molecules* (Basel, Switzerland) 23(2):257. <https://doi.org/10.3390/molecules23020257>
- Sparatore A, Basilico N, Parapini S, Romeo S, Novelli F, Sparatore F, Taramelli D (2005) 4-Aminoquinoline quinolizidinyl- and quinolizidinylalkyl-derivatives with antimalarial activity. *Bioorg Med Chem* 13(18):5338–5345. <https://doi.org/10.1016/j.bmc.2005.06.047>
- Szczepaniak J, Cieslik W, Romanowicz A, Musiol R, Krasowska A (2017) Blocking and dislocation of *Candida albicans* Cdr1p transporter by styrylquinolines. *Int J Antimicrob Agents* 50(2):171–176. <https://doi.org/10.1016/j.ijantimicag.2017.01.044>
- Talamas FX, Abbot SC, Anand S, Brameld KA, Carter DS, Chen J, Davis D, de Vicente J, Fung AD, Gong L, Harris SF, Inbar P, Labadie SS, Lee EK, Lemoine R, Le Pogam S, Leveque V, Li J, McIntosh J, Najera I, Park J, Railkar A, Rajyaguru S, Sangi M, Schoenfeld RC, Staben LR, Tan Y, Taygerly JP, Villasenor AG, Weller PE (2014) Discovery of N-[4-[6-tert-butyl-5-methoxy-8-(6-methoxy-2-oxo-1H-pyridin-3-yl)-3-quinolyl]phenyl]methanesulfonamide (RG7109), a potent inhibitor of the hepatitis C virus NS5B polymerase. *J Med Chem* 57(5):1914–1931. <https://doi.org/10.1021/jm401329s>
- Tavares GSV, Mendonca DVC, Lage DP, Granato JD, Ottoni FM, Ludolf F, Chavez-Fumagalli MA, Duarte MC, Tavares CAP, Alves RJ, Coimbra ES, Coelho EAF (2018) Antileishmanial activity, cytotoxicity and mechanism of action of Clioquinol against leishmania infantum and leishmania amazonensis species. *Basic Clin Pharmacol Toxicol* 123(3):236–246. <https://doi.org/10.1111/bcpt.12990>
- Terzic N, Konstantinovic J, Tot M, Burojevic J, Djurkovic-Djakovic O, Srbijanovic J, Stajner T, Verbic T, Zlatovic M, Machado M, Albuquerque IS, Prudencio M, Sciotti RJ, Pecic S, D'Alessandro S, Taramelli D, Solaja BA (2016) Reinvestigating old pharmacophores: are 4-aminoquinolines and tetraoxanes potential two-stage antimalarials? *J Med Chem* 59(1):264–281. <https://doi.org/10.1021/acs.jmedchem.5b01374>
- Upadhyay A, Kushwaha P, Gupta S, Dodda RP, Ramalingam K, Kant R, Goyal N, Sashidhara KV (2018) Synthesis and evaluation of novel triazolyl quinoline derivatives as potential antileishmanial agents. *Eur J Med Chem* 154:172–181. <https://doi.org/10.1016/j.ejmech.2018.05.014>
- Valdivieso E, Mejias F, Torrealba C, Benaim G, Kouznetsov VV, Sojo F, Rojas-Ruiz FA, Arvelo F, Dagger F (2018) In vitro 4-Aryloxy-7-chloroquinoline derivatives are effective in mono- and combined therapy against *Leishmania donovani* and induce mitochondrial membrane potential disruption. *Acta Trop* 183:36–42. <https://doi.org/10.1016/j.actatropica.2018.03.023>
- Vandekerckhove S, Van Herreweghe S, Willems J, Danneels B, Desmet T, de Kock C, Smith PJ, Chibale K, D'Hooghe M (2015) Synthesis of functionalized 3-, 5-, 6- and 8-aminoquinolines via intermediate (3-pyrrolin-1-yl)- and (2-oxopyrrolidin-1-yl)quinolines and evaluation of their antiplasmodial

- and antifungal activity. *Eur J Med Chem* 92:91–102. <https://doi.org/10.1016/j.ejmech.2014.12.020>
- Vial L, Lepine F, Milot S, Groleau MC, Dekimpe V, Woods DE, Deziel E (2008) *Burkholderia pseudomallei*, *B. thailandensis*, and *B. ambifaria* produce 4-hydroxy-2-alkylquinoline analogues with a methyl group at the 3 position that is required for quorum-sensing regulation. *J Bacteriol* 190 (15):5339–5352. <https://doi.org/10.1128/JB.00400-08>
- Videnovic M, Opsenica DM, Burnett JC, Gomba L, Nuss JE, Selakovic Z, Konstantinovic J, Krstic M, Segan S, Zlatovic M, Sciotti RJ, Bavari S, Solaja BA (2014) Second generation steroidal 4-aminoquinolines are potent, dual-target inhibitors of the botulinum neurotoxin serotype A metalloprotease and *P. falciparum* malaria. *J Med Chem* 57(10):4134–4153. <https://doi.org/10.1021/jm500033r>
- Villa P, Arumugam N, Almansour AI, Suresh Kumar R, Mahalingam SM, Maruoka K, Thangamani S (2019) Benzimidazole tethered pyrrolo[3,4-b]quinoline with broad-spectrum activity against fungal pathogens. *Bioorg Med Chem Lett* 29(5):729–733. <https://doi.org/10.1016/j.bmcl.2019.01.006>
- Wadhwa P, Jain P, Rudrawar S, Jadhav HRA (2018) Quinoline, coumarin and other heterocyclic analogs based HIV-1 integrase inhibitors. *Curr Drug Discov Technol* 15(1):2–19. <https://doi.org/10.2174/1570163814666170531115452>
- Wanka L, Iqbal K, Schreiner PR (2013) The lipophilic bullet hits the targets: medicinal chemistry of adamantane derivatives. *Chem Rev* 113 (5):3516–3604. <https://doi.org/10.1021/cr100264t>
- Waters NC, Edstein MD (2011) 8-Aminoquinolines: primaquine and tafenoquine. In: Treatment and prevention of malaria. Springer, Basel, pp 69–94
- Wijnant GJ, Van Bocxlaer K, Yardley V, Murdan S, Croft SL (2017) Efficacy of paromomycin-chloroquine combination therapy in experimental cutaneous leishmaniasis. *Antimicrob Agents Chemother* 61(8):pii: e00358-17
- Wilson TA, Koneru PC, Rebensburg SV, Lindenberger JJ, Kobe MJ, Cockroft NT, Adu-Ampratwum D, Larue RC, Kvaratskhelia M, Fuchs JR (2019) An Isoquinoline scaffold as a novel class of allosteric HIV-1 integrase inhibitors. *ACS Med Chem Lett* 10 (2):215–220. <https://doi.org/10.1021/acsmchemlett.8b00633>
- Woodring JL, Patel G, Erath J, Behera R, Lee PJ, Leed SE, Rodriguez A, Sciotti RJ, Mensa-Wilmot K, Pollastra MP (2015) Evaluation of aromatic 6-substituted Thienopyrimidines as scaffolds against parasites that cause trypanosomiasis, Leishmaniasis, and malaria. *Med Chem Commun* 6(2):339–346. <https://doi.org/10.1039/C4MD000441H>
- World Health Organization (2013). https://www.who.int/tb/features_archive/bedaquilinelaunch/en/. Accessed 20 Apr 2019
- World Health Organisation (2017). <https://apps.who.int/iris/bitstream/handle/10665/258973/WER9238.pdf?sequence=1>. Accessed 22 Mar 2019
- World Health Organisation (2018) Leishmaniasis facts. <http://www.who.int/en/news-room/fact-sheets/detail/leishmaniasis>. Accessed 22 Mar 2019
- World Health Organisation (2019). <https://apps.who.int/iris/bitstream/handle/10665/275867/9789241565653-eng.pdf?ua=1>. Accessed 21 Mar 2019
- Xu Z, Zhao SJ, Lv ZS, Gao F, Wang Y, Zhang F, Bai L, Deng JL (2019) Fluoroquinolone-isatin hybrids and their biological activities. *Eur J Med Chem* 162:396–406. <https://doi.org/10.1016/j.ejmech.2018.11.032>
- Zablotskaya A, Segal I, Geronikaki A, Shestakova I, Nikolajeva V, Makarenkova G (2017) N-heterocyclic choline analogues based on 1,2,3,4-tetrahydro(iso)quinoline scaffold with anticancer and anti-infective dual action. *Pharmacol Rep* 69(3):575–581. <https://doi.org/10.1016/j.pharep.2017.01.028>
- Zhang H, Collins J, Nyamwihura R, Ware S, Kaiser M, Ogungbe IV (2018) Discovery of a quinoline-based phenyl sulfone derivative as an antitrypanosomal agent. *Bioorg Med Chem Lett* 28(9):1647–1651. <https://doi.org/10.1016/j.bmcl.2018.03.039>
- Zuo R, Garrison AT, Basak A, Zhang P, Huigens RW 3rd, Ding Y (2016) In vitro antifungal and antibiofilm activities of halogenated quinoline analogues against *Candida albicans* and *Cryptococcus neoformans*. *Int J Antimicrob Agents* 48(2):208–211. <https://doi.org/10.1016/j.ijantimicag.2016.04.019>