

XCIND as a genetic disease of X-irradiation hypersensitivity and cancer susceptibility

Shuki Mizutani · Masatoshi Takagi

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Abstract The XCIND syndrome is named after distinct hypersensitivity to ionizing (X-ray) irradiation, cancer susceptibility, immunodeficiency, neurological abnormality, and double-strand DNA breakage. The disorders comprising XCIND syndrome are usually inherited in an autosomal recessive manner. Ataxia telangiectasia (A-T) is one such disease, and is caused by biallelic germline mutation of the Ataxia telangiectasia mutated (*ATM*) gene. Heterozygous carriers of the *ATM* mutation, who do not show A-T-like clinical symptoms, are estimated to comprise 1 % of the population. Thus, understanding the biological basis of XCIND, including A-T, should help shed light on the pathogenesis of genetic diseases with cancer susceptibility.

Keywords XCIND · Ataxia telangiectasia · ATM · DNA damage response

Introduction

Human beings are exposed to various insults to their cellular DNA in their everyday lives. However, the human body is equipped with DNA repair systems, which enable us to survive without developing devastating diseases due to the accumulation of DNA damage. There are two types of DNA damage: endogenous and exogenous. Endogenous DNA damage is caused by stresses associated with cellular oxygenation, DNA replication in cell proliferation, and

DNA recombination of antigen receptor genes, while exogenous insults are caused by such agents as UV irradiation, natural and/or medical ionizing radiation, and DNA-damaging agents, such as anti-cancer drugs or environmental toxic factors. Ionizing radiation causes double-strand breaks of DNA associated with the generation of oxygen radicals. DNA damage responses and repair systems are prerequisites for normal development and differentiation of embryonic stem cells. However, in some individuals these repair systems are genetically defective, causing genetic hypersensitivity to ionizing radiation and hence genomic instability.

One of the most reliable and traditional means for testing hypersensitivity to X-irradiation (X-IR) is colony-survival assay (CSA). But evaluating sensitivity to X-IR-induced DNA damage requires substantial experience. Instead of employing CSA, we can also use a foci formation assay for γ H2AX, 53BP1, or BRCA1 as surrogate marker, which is called the irradiation-induced foci (IRIF) kinetics method [1].

In this review we describe the X-IR hypersensitive syndrome XCIND, focusing on ataxia telangiectasia as a main representative disorder. Although XCIND disorders are rather rare, they may provide essential insights for understanding mechanisms of human diseases caused by concomitant effects of genetic and environmental factors, such as X-IR.

XCIND as a disease characterized by hypersensitivity to X-IR

XCIND is usually inherited in an autosomal recessive manner. Diseases belonging to the XCIND family are listed in Table 1 [2, 3]. Ruling out the possibility of XCIND is

S. Mizutani (✉) · M. Takagi
Department of Pediatrics and Developmental Biology,
Tokyo Medical and Dental University,
Yushima 1-5-45, Bunkyo-Ku, Tokyo 113-8519, Japan
e-mail: skkmiz@gmail.com

essential when patients are to be treated by chemotherapy and/or transplantation of hematopoietic stem cells for immunodeficiency or hematological malignancies, as hypersensitivity to X-IR leads to concomitant hypersensitivity to conditioning regimens, including chemotherapeutic agents which cause DNA damage.

The repair pathways for DNA double-strand breaks (DSB) include homologous recombination (HR) and non-homologous end joining (NHEJ) [4]. NHEJ repair requires KU70/80, DNA-PKcs (PRKDC), Artemis (DCLRE1C), DNA Ligase IV (LIG4), XRCC4, and Cernunnos/XLF (NHEJ1) proteins regardless of whether the source of the insult is endogenous or exogenous. One endogenous form of DNA DSB is that of antigen receptor genes [5], which is the reason defects in these proteins lead to lymphocyte deficiency as well as hypersensitivity to X-IR associated with frequent chromosomal abnormalities. A-T, Nijmegen breakage syndrome (NBS), and A-T-like disorder (ATLD)

are often associated with chromosomal translocation of chromosome 7 and 14, mainly involving the T cell receptor (TCR) or immunoglobulin (Ig) locus, while LIG4 deficiency is associated with random chromosomal translocations. To maintain chromosomal integrity, cell cycle checkpoints also play essential roles. This is because DNA replication or cell division should not be allowed to initiate until DNA repair is completed, as revealed by extensive studies on A-T [6].

More recently, RNF 168 deficiency, RAD50 deficiency, and DNA-PKcs deficiency have been found to be involved in human radiation sensitivity disorders. RNF168 is involved in a ubiquitin ligase cascade of histone chromatin H2A, a deficiency reported to cause RIDDLE syndrome, which is characterized by radiosensitivity, immunodeficiency, dysmorphic features and learning difficulties [7, 8]. RAD50 is one of the components of the MRE11/RAD50/NBS (MRN) complex, which functions in DSB repair by recognizing DSB sites. The loss of RAD50 function was found to lead to increased radiosensitivity, as well as microcephaly [9]. These clinical features are also shared by NBS, DNA LIG4 deficiency, and Cernunnos/XLF deficiency. In these cases, patients usually carry hypomorphic mutations of the genes; complete deficiency of the protein leads to embryonic lethality. DNA-PKcs deficiency has previously been shown to lead to severe combined immunodeficiency (SCID) in mice, horses, and dogs. It has long been a question whether germline mutations of DNA-PKcs are involved in human SCID. Hypomorphic mutation has recently been discovered in human SCID [10]. Details of the individual diseases are described below and shown in Table 2.

Ataxia telangiectasia, a representative XCIND disease

A-T is a progressive neurodegenerative disorder with onset of truncal ataxia usually before 3 years of age. Ocular

Table 1 Disorders included in XCIND

Disorder	Documented year	Gene mutated
Ataxia telangiectasia	1926	<i>ATM</i>
Fanconi anemia	1927	<i>FANC</i> genes
X-linked agammaglobulinemia	1952	<i>BTK</i>
SCID AD	1979	<i>ADA</i>
Nijmegen breakage syndrome	1981	<i>NBS1</i> (NBN)
ATLD	1999	<i>MRE11A</i>
Ligase IV deficiency	1999	<i>LIG4</i>
SCID-Artemis deficiency	2001	<i>Artemis</i> (<i>DCLRE1C</i>)
SCID-XLF/Cernunnos deficiency	2006	<i>XLF/Cernunnos</i> (<i>NHEJ1</i>)
RIDDLE syndrome	2009	<i>RNF168</i>
RAD50 deficiency	2009	<i>RAD50</i>
SCID	2009	<i>DNA-PKcs</i> (<i>PRKDC</i>)

Table 2 Phenotype of XCIND

	Ataxia	Telangiectasia	Immunodeficiency	Microcephaly	Cancer predisposition	Increased AFP	Chromosomal translocation including TCR or Ig
AT	+	+	+ T cell and immunoglobulin	–	+	+	+
NBS	–	–	+ T cell and immunoglobulin	+	+	–	+
ATLD	+	–	– Deficiency in specific functional antibody	–	–	–	+
	mild			or + various	or + various		
NBSLD	+	–	–	+	–	–	+
	mild						
RIDDLE syndrome	+	+	+ Decreased immunoglobulin level	+	–	+	No information

apraxia is also present in most patients. By the age of 10, patients are wheelchair-bound due to ataxia. One-third of patients have severe immunodeficiencies, accompanied by severe sino-pulmonary infections with non-opportunistic organisms. Conjunctival telangiectasia is a well-known characteristic feature that appears several years after the onset of neurologic symptoms [11].

Increase of serum alpha-fetoprotein (AFP), chromosomal translocations involving chromosomes 7 and 14, and a decrease in the lymphocytes of B cell and T cell compartments are the most useful test for the diagnosis of A-T. AFP is elevated in almost all the cases of A-T, and is the most useful clinical laboratory test for bedside diagnosis. T cell numbers are usually low, and many patients show only marginal deficiencies. γ/δ T cell numbers are usually elevated. B cell numbers are normal or slightly elevated. Deficiencies of IgE, IgG2, and IgA are often marked. Glucose intolerance is also frequently observed, suggesting that ATM may somehow be involved in the pathogenesis of DM in the general population.

Radiosensitivity is the most traditional and reliable test to confirm the clinical diagnosis of A-T. Colony-survival assay (CSA) and radioresistant DNA synthesis have been validated for clinical diagnostic testing and CSA was reported to be one of the most reliable assays to identify A-T patients [1], when tested in experienced laboratories.

The responsible gene in A-T is localized at chromosome at 11q22.3 and called the *ATM* gene, which contains 66 exons spanning approximately 150 kb of genomic DNA.

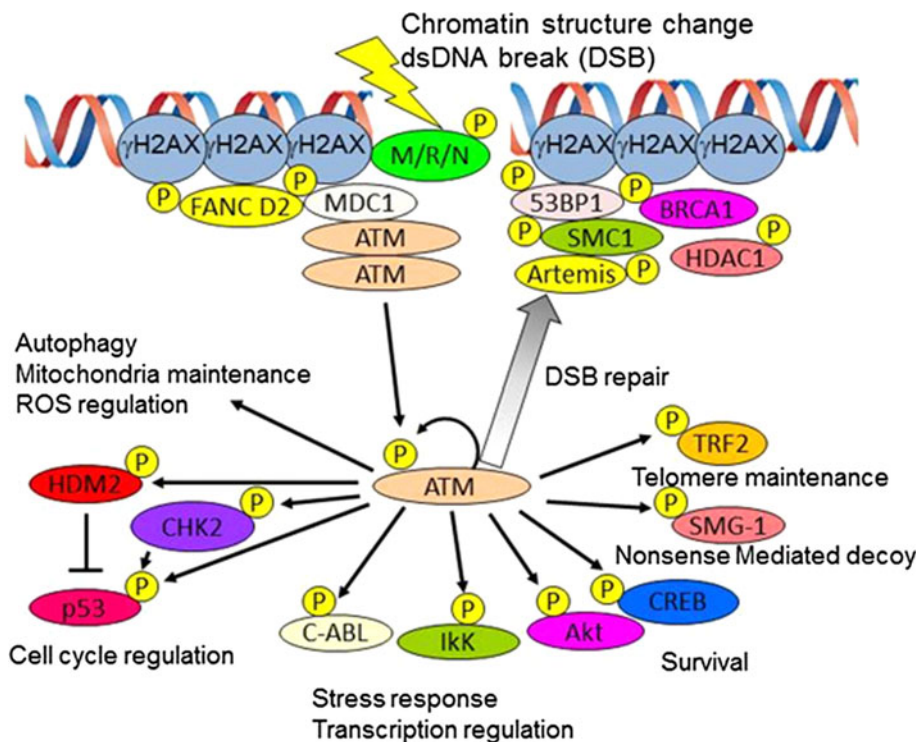
Its cDNA consists of 10140 bp. *ATM* mutation in A-T is inherited in an autosomal recessive manner. The frequency of A-T patients is estimated as 1 in 40000–300000, and heterozygous carriers of the *ATM* gene mutation reportedly comprise 0.5–1 % of the general population.

ATM protein and its function

The ATM protein is composed of 3056 amino acids encoded by a 9168-bp open reading frame [6]. The *ATM* gene mutation is not limited to A-T patients, and the *ATM* gene is mutated secondarily in various hematological malignancies, which lead to the speculation that *ATM* plays a role as a tumor suppressor gene [12]. The ATM protein encodes phosphoinositide 3-kinase-related kinase. When cells are exposed to genotoxic stresses, such as anticancer drugs or X-IR-induced DNA double-strand breaks, the ATM protein plays a central role in the DNA damage response (Fig. 1). ATM is present as a dimer or higher-order multimer in unstressed cells. During DNA damage response (DDR), the canonical pathway is activated in which ATM undergoes autophosphorylation on at least three sites (Ser367, Ser1893, and Ser1981). Autophosphorylation of these sites breaks apart the inactive, non-covalently bound ATM dimer, leading to the activation of the ATM-induced repair pathway [6, 13].

The activation of ATM protein kinase activity has been shown to be dependent on the presence of the MRN

Fig. 1 DNA damage response activates ATM



complex (Mre11/Rad50/Nbs1), which is a kind of sensor of DNA breakage. Once activated, ATM then phosphorylates a series of substrates that participate in signaling to the cell cycle checkpoints, DNA repair, or cell death. One of the most important substrates is the tumor suppressor protein p53 [14]. p53 is activated by phosphorylation in an ATM-dependent manner and prevents the passage of cells from G1 to S phase, or induces apoptosis when DNA damage is too profound to be repaired. Other well-characterized ATM substrates are the checkpoint kinase Chk2, the breast cancer susceptibility protein BRCA1, and Nbs1(NBN), which is mutated in the human genetic disorder Nijmegen breakage syndrome (NBS) [15], though there are many substrates yet not characterized.

ATM and cancer

The most frequent cancers in A-T are lymphoid tumors, which are observed in 15–30 % of A-T patients [11]. Lymphoid tumors frequently involve chromosomal translocation at chromosome 7 and 14 [16]. Recently, it was reported that TCR δ , but not the TCR α region, is involved in chromosome 14 translocation [17] and this is closely associated with recombination defects of the TCR β region in the double-negative 3a (DN3a) stage of thymocyte development. In the DN3a stage, RAG-dependent chromosomal breaks occur at the TCR δ loci, and aberrant chromosomal translocations are induced while developing to DN3b. Thus, T cell deficiency and chromosomal translocation are closely related in A-T [18].

A-T patients reportedly suffer from other cancers of the stomach, liver, ovaries, salivary glands, oral tissue, breast, and pancreas as well. A-T heterozygous carriers are also reported to be at relatively higher risk for breast cancer. Breast cancer is one of the most frequent secondary cancers in Hodgkin disease (HD), and risk increases especially after irradiation. It should be noted that in childhood HD, dominant negative mutations of the germline *ATM* gene are often seen, suggesting the possible involvement of a genetic background for childhood HD concomitantly with breast cancer [19].

Recent studies have indicated that the progression of preneoplastic lesions to neoplasia is inhibited by the existence of tumorigenesis barriers. Response to DNA replication stress is one such barrier, leading to activation of the DNA damage checkpoint and thereby to apoptosis or cell cycle arrest. It has been shown that oncogene-induced senescence is associated with signs of DNA replication stress, such as prematurely terminated DNA replication forks and DNA double-strand breaks. In ATM-deficient mice, the induction of senescence is absent, which leads to increased tumor proliferation. Thus, it has been suggested

that senescence in human preneoplastic lesions represents a barrier to malignant progression. This has been shown to be the case in early lesions of colon cancer. Precancerous colon adenoma expresses activated ATM protein, which corresponds to the autophosphorylation responding to replication stress of initial events in oncogene-induced cancer development [20–22]. It has also been shown that ATM is activated in myelodysplastic syndrome (MDS), which is a preleukemic condition caused by oncogenic stresses in hemopoietic stem cells, but is disrupted eventually when MDS evolves into overt leukemia [21]. Thus, ATM has a role in preventing the progression of cancer.

Other diseases in the XCIND syndrome

Nijmegen breakage syndrome

The Nijmegen breakage syndrome is an autosomal recessive chromosomal instability syndrome characterized by microcephaly, microgenia, “bird-like” face, growth retardation, mild mental retardation, immunodeficiency, and a predisposition to cancer with normal serum levels of alpha-fetoprotein. Cancer susceptibility is much higher than in AT. However, patients with NBS have neither ataxia nor telangiectasia. Almost all patients reportedly originate from Eastern Europe, and are homozygous for one founder hypomorphic NBN (NBS1) mutation c.657_661del5 originating from Slavic lines. Cells from NBS patients exhibit similar features with those of AT, namely hyper-radio-sensitivity, radio-resistant DNA synthesis, and chromosomal rearrangements involving TCR and Ig loci. [23].

Nijmegen breakage syndrome-like disorder (NBSLD)

Waltes et al. identified compound heterozygosity for mutations in the *RAD50* gene from NBS-like phenotype patients. The mutations were a combination of a nonsense mutation and a mutation that produces a larger polypeptide than the wild type. As a result, it produces a low level of unstable *RAD50* protein, presumably a hypomorphic mutation. Patients exhibit microcephaly, mild mental retardation, and mild ataxia. However, patients with NBSLD reportedly show neither immunodeficiency nor predisposition to cancer, although further study is needed. Cells from NBSLD patients exhibit moderately increased radiosensitivity and chromosomal rearrangements involving TCR and Ig loci. [9].

Ataxia telangiectasia-like disorder (ATLD)

Mutation in the *MRE11A* gene was identified from a clinically diagnosed AT-like patient. Clinical features are

much milder than AT, and the symptoms appear much more slowly than in AT. The appearance of ataxia, for example, is much later than AT, and telangiectasia and increased alpha-fetoprotein are absent. Although immunoglobulin levels are almost normal, the generation of specific functional antibodies is attenuated [24]. It has been thought that ATLD patients do not exhibit predisposition to cancer, but we have identified a Japanese ATLD family who developed lung cancer [25]. An original report described ATLD as a milder phenotype of AT. However, a more severe phenotype including microcephaly or NBS-like dimorphic features has been reported [25, 26]. MRE11A mutation affects M/R/N complex stability, and decreased NBS1 and/or RAD50 is observed. Residual activity of NBS1 and/or RAD50 may affect the difference in phenotypes. The cellular biological features are the same as in AT, including increased radiosensitivity and an increased level of spontaneously occurring chromosome aberrations involving the TCR or Ig region.

RIDDLE syndrome

RIDDLE syndrome is named after its clinical features, including increased radiosensitivity, immunodeficiency, dysmorphic features, and learning difficulties. Patients exhibit ataxia, telangiectasia, growth retardation, and microcephaly [8]. The gene responsible for RIDDLE syndrome is RNF168 [7]. Patients carry compound heterozygous nonsense mutations in the *RNF168* gene, resulting in aberrant RNF168 proteins. RNF168 is a ubiquitin ligase, and RNF168-dependent chromatin modifications orchestrate the accumulation of 53BP1 and BRCA1 to DNA lesions; their loss is the likely cause of the cellular and developmental phenotypes associated with RIDDLE syndrome. Cellular biological features include inability to recruit 53BP1 to sites of DNA double-strand breaks, moderately increased radiosensitivity, and dysregulation of cell cycle checkpoints after irradiation.

Severe combined immunodeficiency (SCID)

SCID is an immunodeficiency in which both B cell and T cell functions are impaired due to defects in one of several possible genes. One of the genes responsible for SCID is the gene encoding the common gamma chain (γ_c), shared by receptors for interleukins such as IL-2, IL-7, and IL-15. Attenuation of NHEJ during VDJ recombination also leads to SCID development. The molecules involved in NHEJ are known to be LIG4, Artemis (DCLRE1C), XLF/Cernunnos (NHEJ1), and DNA-PKcs (PKRDC) [5]. Although all of the mutated genes are directly responsible for SCID development, the phenotypes differ according to the gene

responsible. Only LIG4-deficient and XLF/Cernunnos-deficient SCID exhibits microcephaly and growth retardation.

Conflict of interest The authors declare no competing financial interests.

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